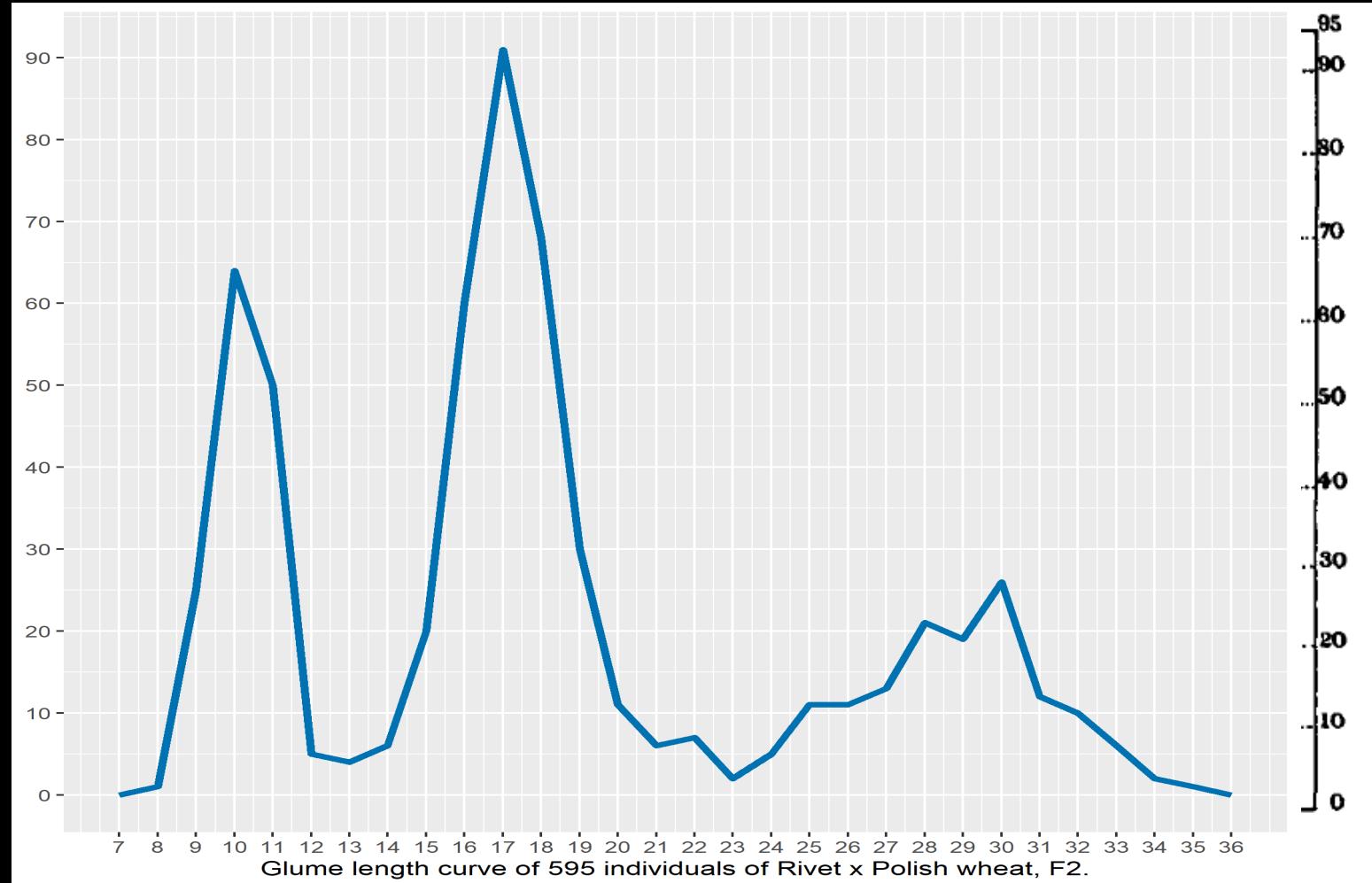


Solving a 100-year old mystery: Cloning the *P1* locus in *Triticum polonicum*

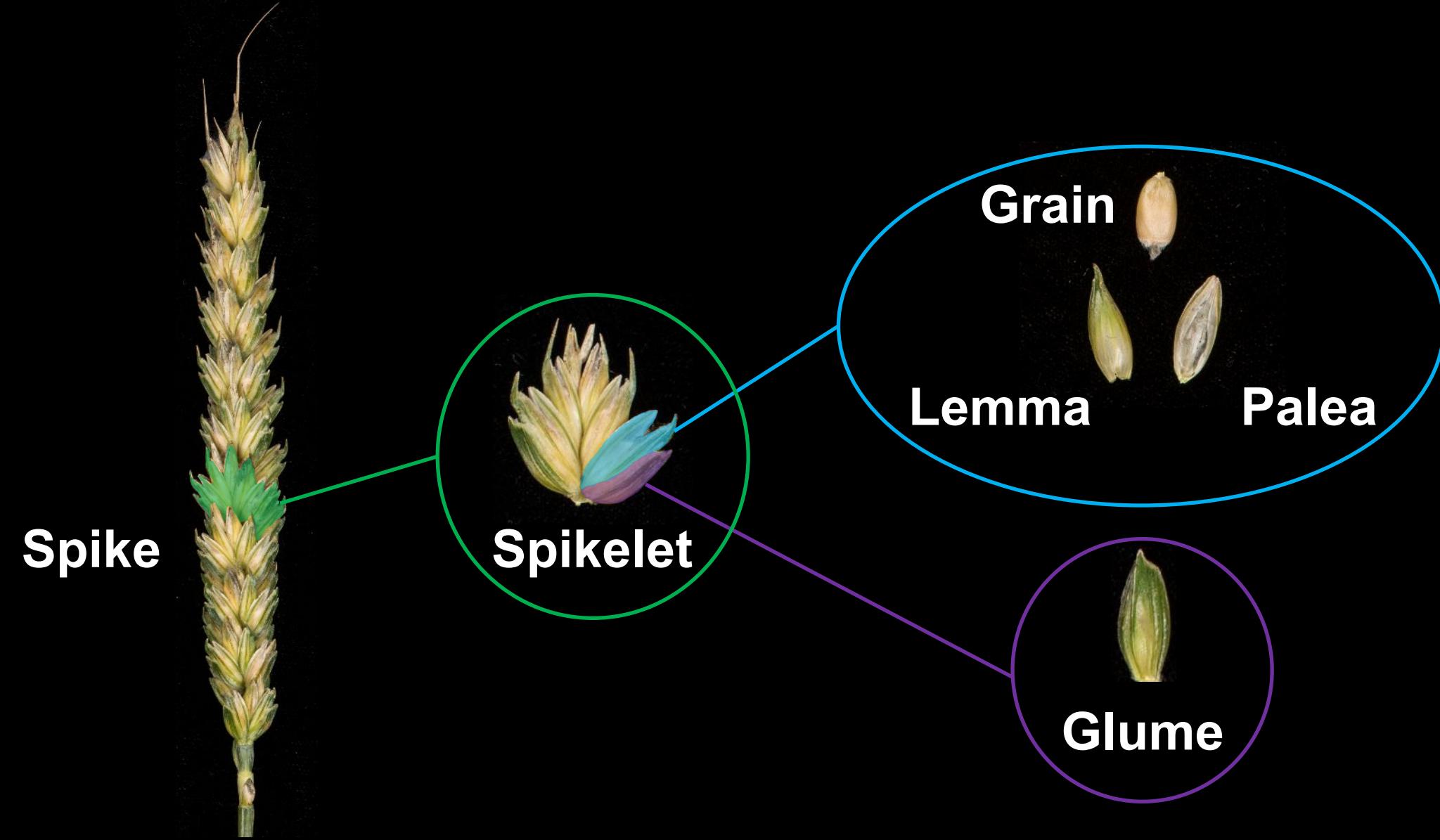


The *P1* locus of *Triticum polonicum* confers long glumes and grains



Biffen 1905; Matsumura, 1950; Watanabe *et al.*, 1996; Kosuge *et al.*, 2010; Okamoto and Takumi, 2013

Short Terminology



Spikelet composition



Near Isogenic Lines (NILs)



X



→

Backcross by phenotype (long glumes)



BC 4 / BC 6 NILs

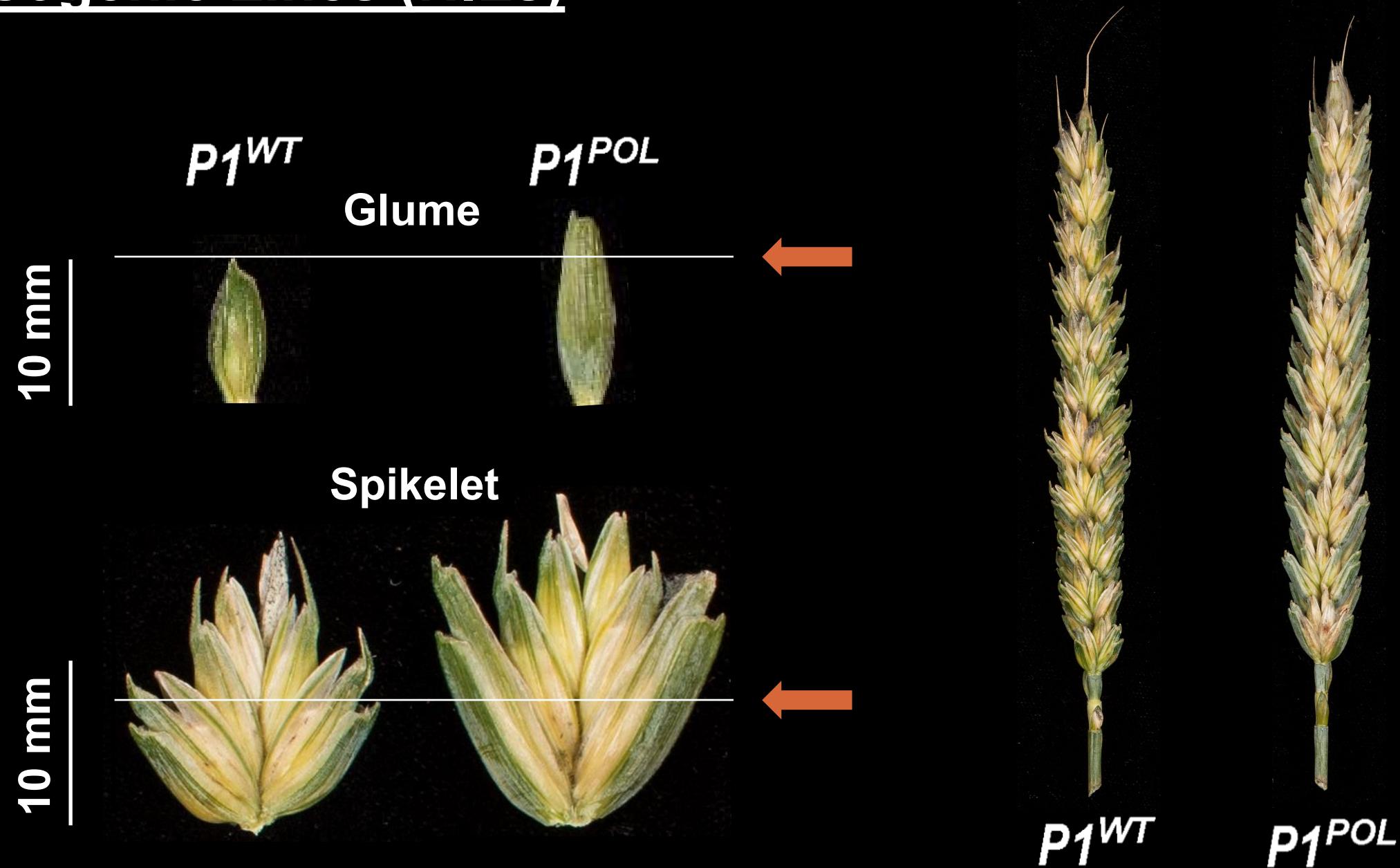
T. polonicum

T. aestivum
cv. Paragon



James Simmonds

Near Isogenic Lines (NILs)



Phenotypic effects of *P1* in field-grown NILs



P1^{WT}

P1^{POL}

+5.0% *** Grain length

+5.5% *** Thousand Grain Weight

+0.5% NS Yield

6 environments over 5 years; all data in Adamski *et al.*, 2021

Phenotypic effects of *P1* in field-grown NILs



$P1^{WT}$

$P1^{POL}$

+6 cm ***

Crop Height

+1.6cm **

Spike length

-0.8 days ***

Time to Heading

Breeder's toolkit



6 environments over 5 years; all data in Adamski *et al.*, 2021

Mapping the *P1* locus - BC4 recombinants

Markers ordered via POPSEQ (genetic; cM)

| POPSEQ [cM] | 98.56 | - | 99.25 | 99.82 | 100.43 | 102.14 | 107.1 |
|-------------|------------|-----------|------------|------------|------------|------------|------------|
| Axiom SNP | BS00022435 | Phenotype | BS00022469 | BS00022306 | BS00021964 | BS00033578 | BS00077445 |
| Paragon | Paragon | Paragon | Paragon | Paragon | Paragon | Paragon | Paragon |

Polonicum

T-pol

T-pol

T-pol

T-pol

T-pol

T-pol

T-pol

Mapping the *P1* locus - BC4 recombinants

Markers ordered via POPSEQ (genetic; cM)

Mapping the *P1* locus - BC4 recombinants

Markers ordered with RefSeqv0.4 (physical; bp)

Mapping the P1 locus - BC4 recombinants



Markers ordered with RefSeqv0.4 (physical; bp)

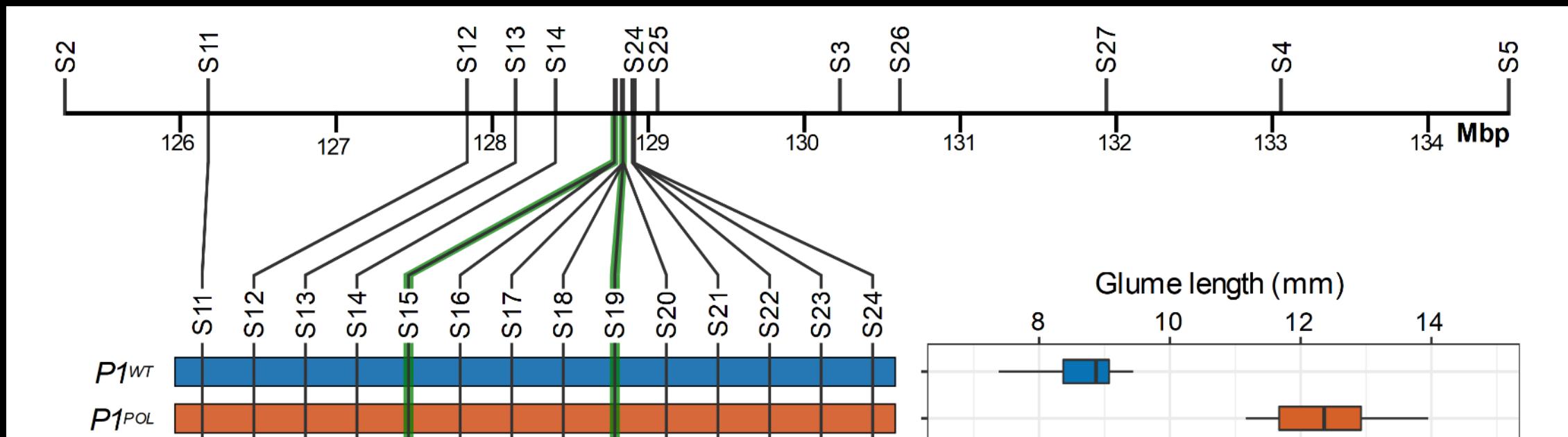
| | | | | | | | |
|-----------------|-------------|-------------|-------------|-------------|-------------|-----------|-------------|
| POPSEQ [cM] | 99.82 | 100.43 | 99.25 | 102.14 | 98.56 | - | 107.1 |
| Axiom SNP | BS00022306 | BS00021964 | BS00022469 | BS00033578 | BS00022435 | - | BS00077445 |
| RefSeqv0.4 [bp] | 117,276,239 | 118,073,697 | 120,105,578 | 120,899,237 | 127,502,160 | Phenotype | 544,983,113 |

Mapping the *P1* locus - BC4 recombinants

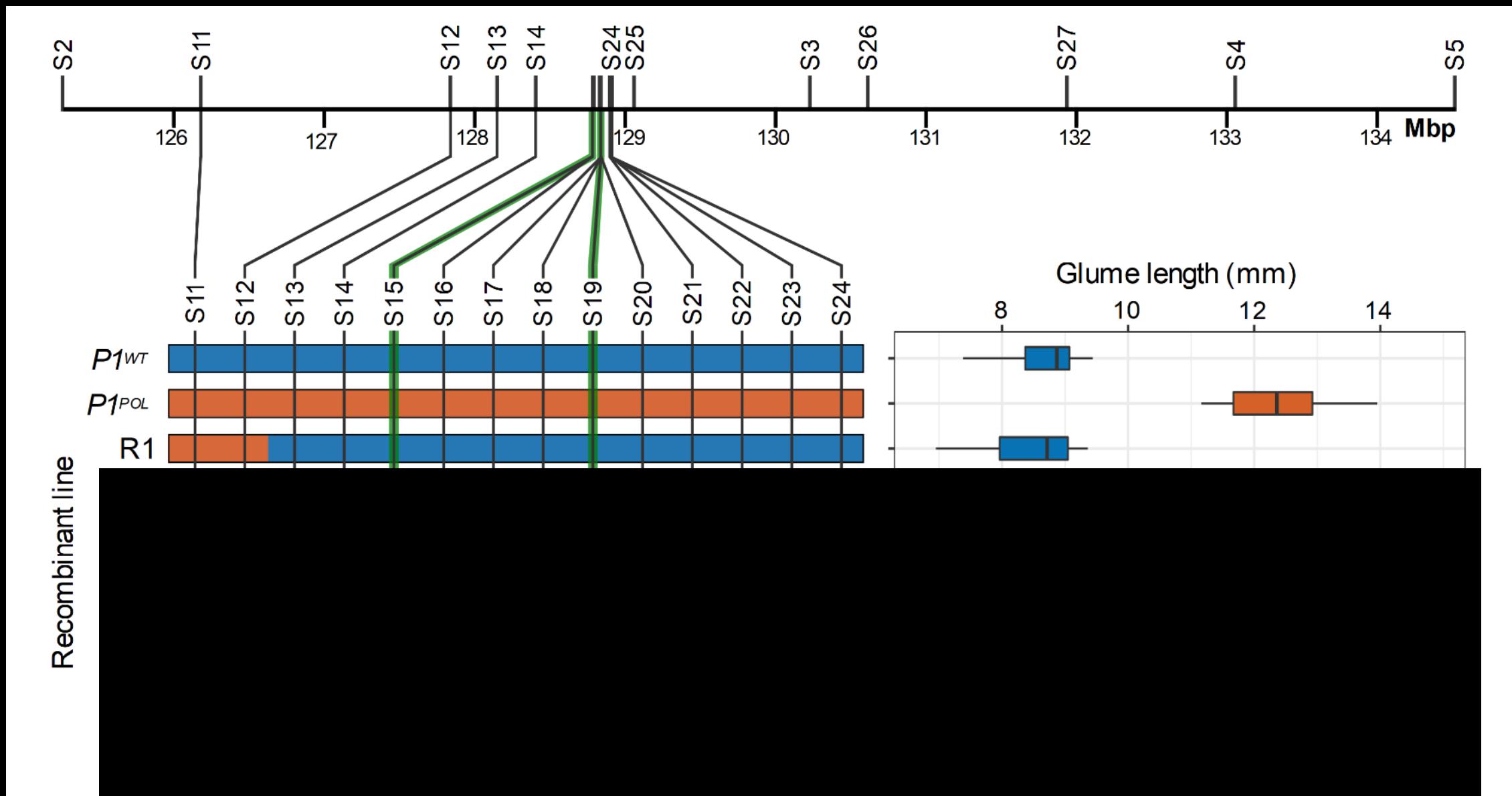
Markers ordered with RefSeqv0.4 (physical; bp)



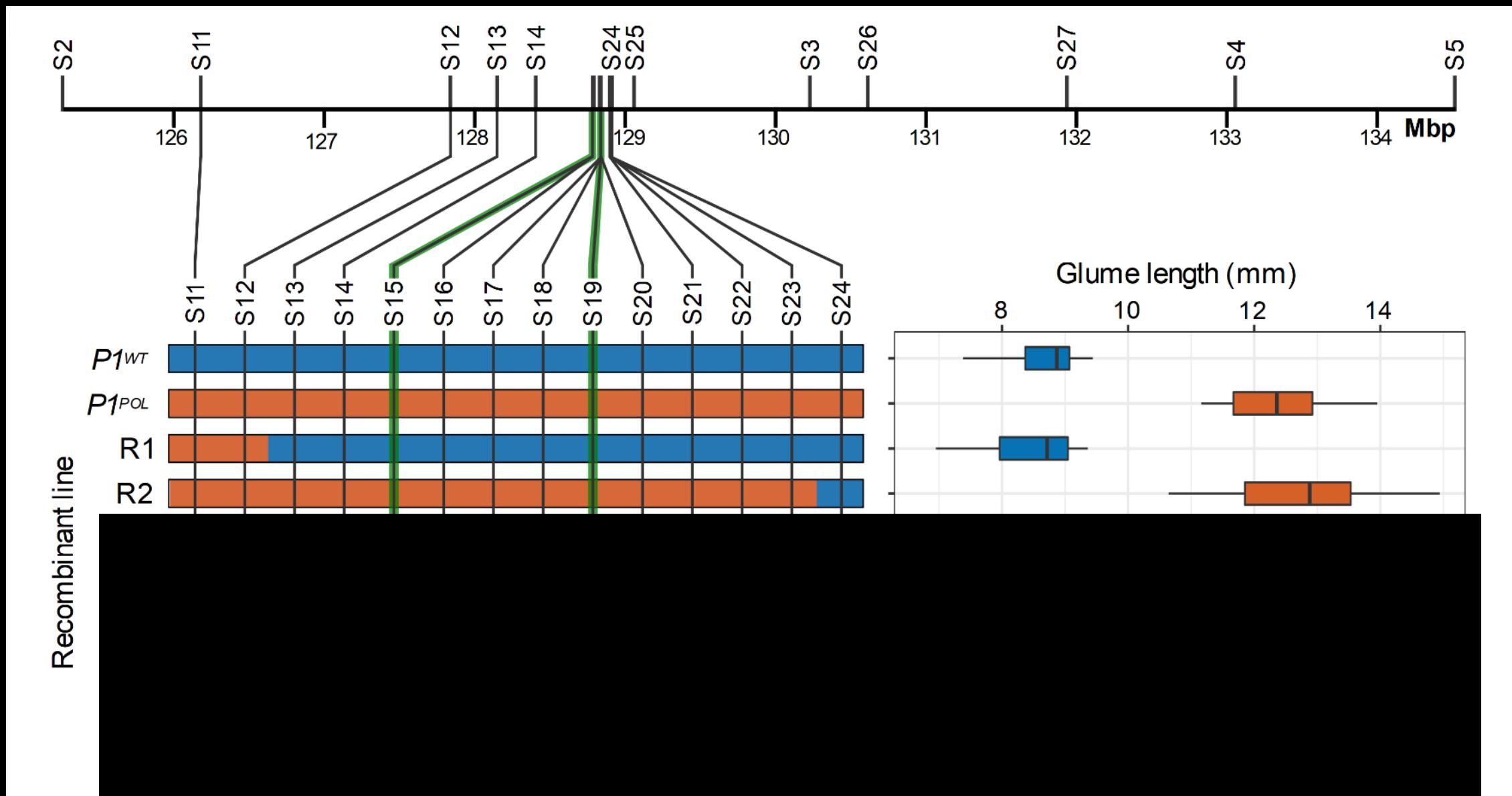
Mapping the *P1* locus - BC6 recombinants



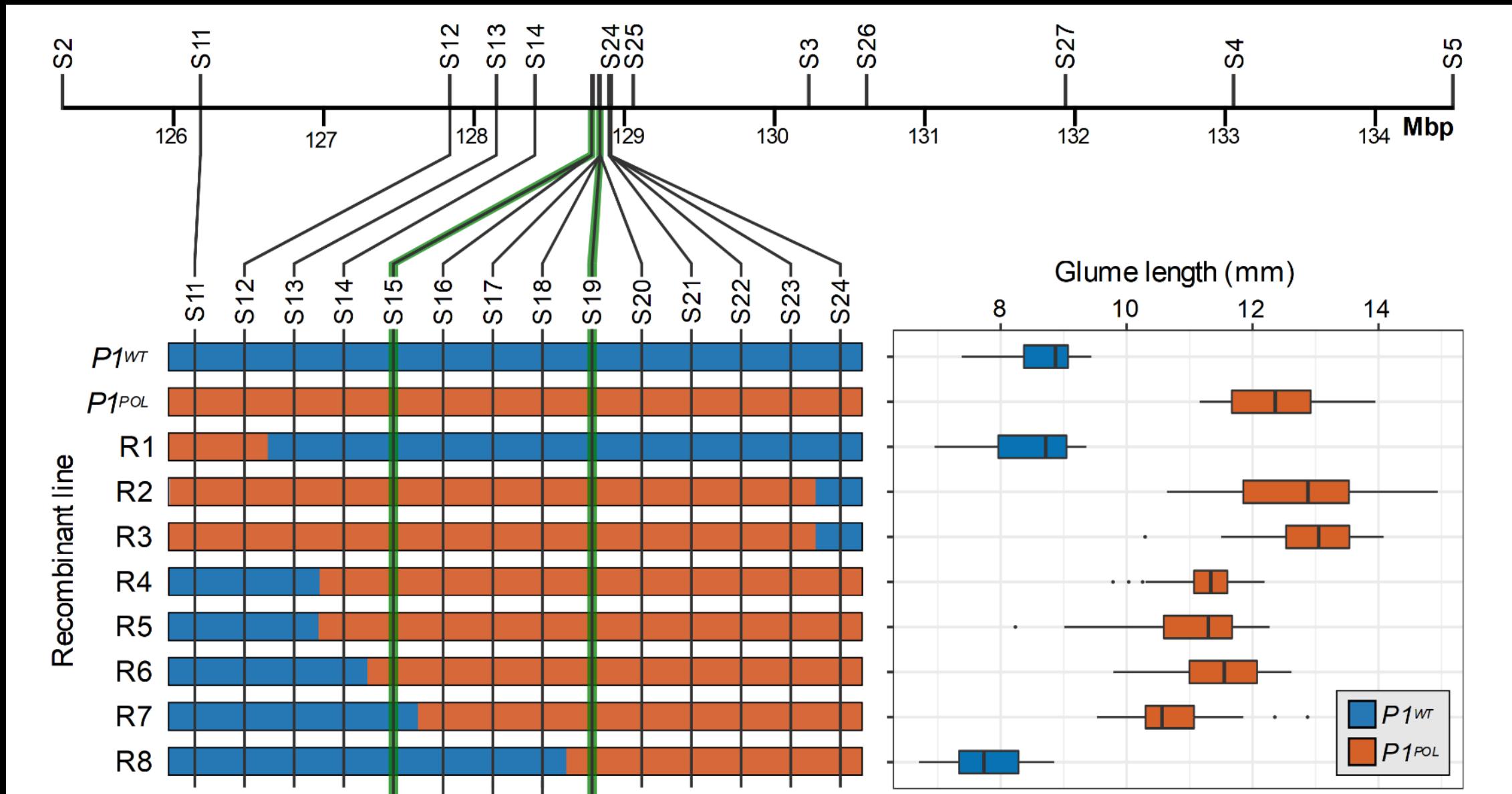
Mapping the *P1* locus - BC6 recombinants



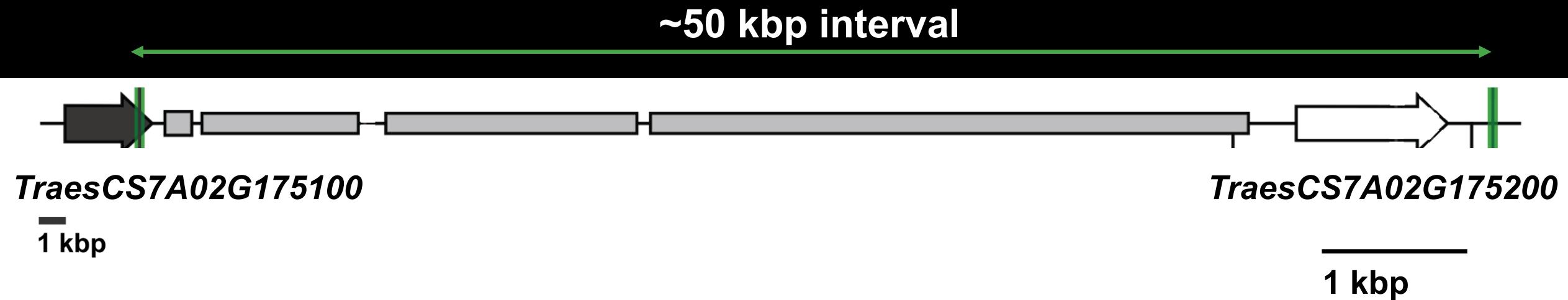
Mapping the *P1* locus - BC6 recombinants



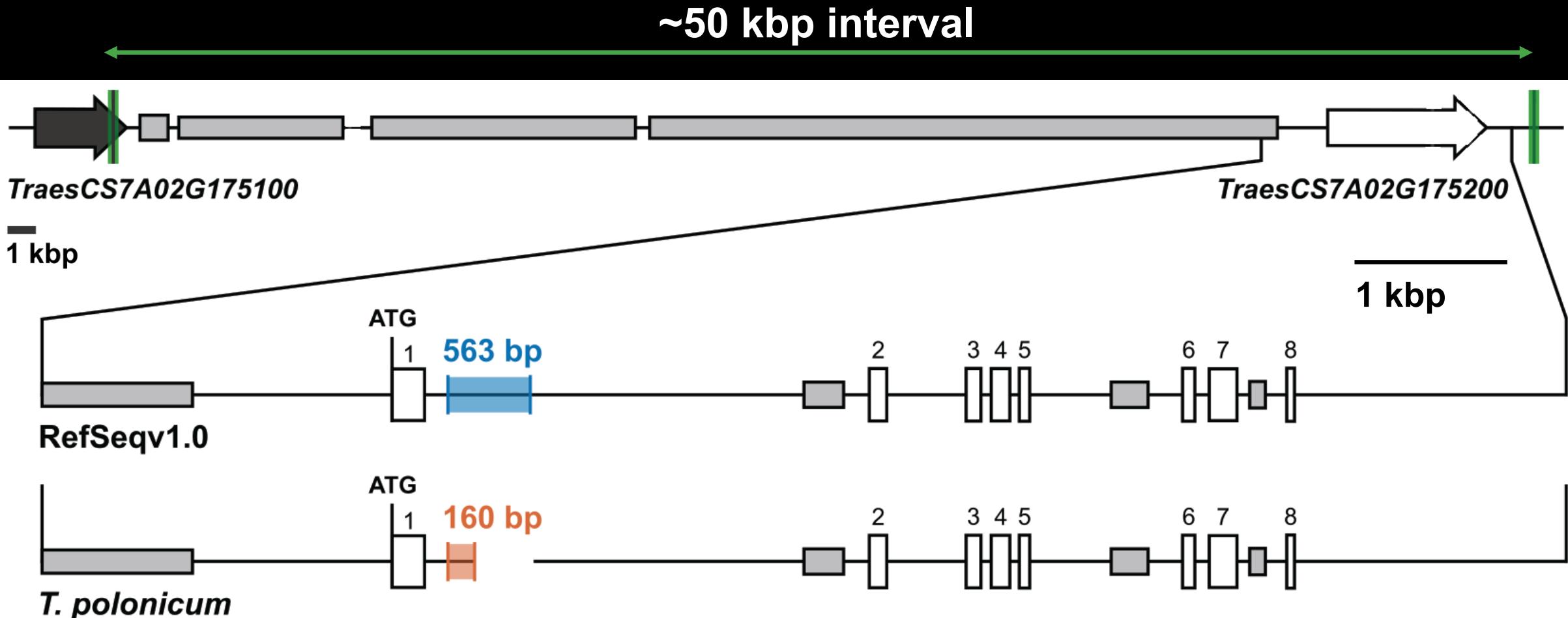
Mapping the *P1* locus - BC6 recombinants



Fine-mapping the *P1* locus



Fine-mapping the *P1* locus



Allele classification

TraesCS7A02G175200 is a *Short Vegetative Phase (SVP)* homolog

Allele classification

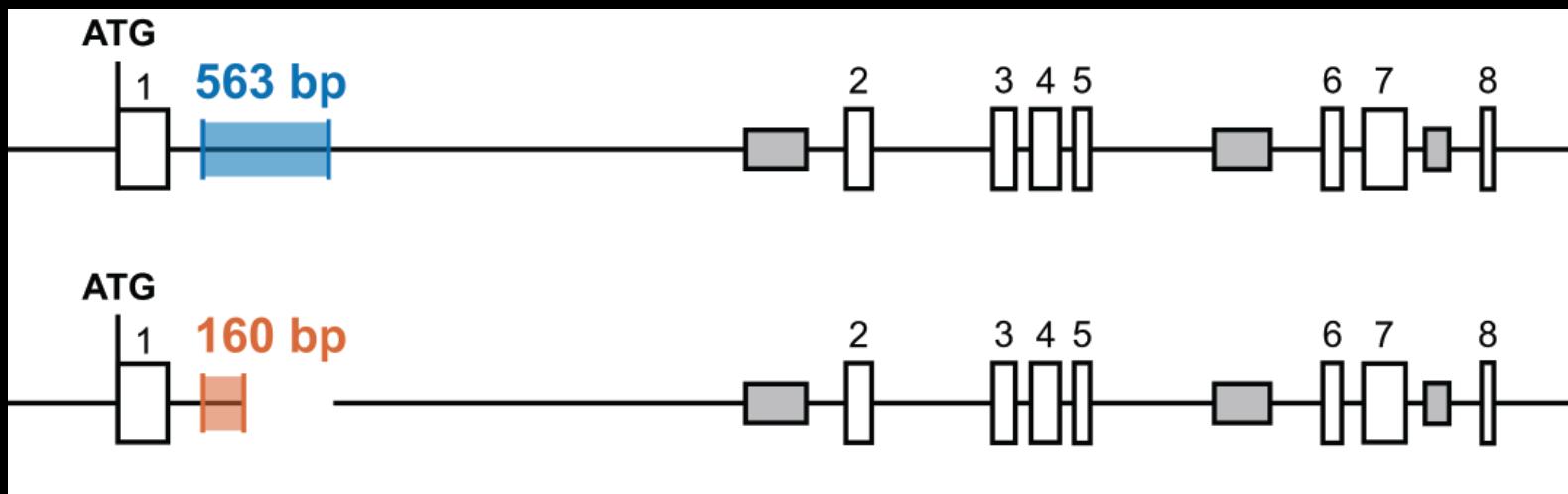
TraesCS7A02G175200 is a *Short Vegetative Phase (SVP)* homolog

VEGETATIVE TO REPRODUCTIVE TRANSITION 2 (VRT2)

Allele classification

TraesCS7A02G175200 is a *Short Vegetative Phase (SVP)* homolog

VEGETATIVE TO REPRODUCTIVE TRANSITION 2 (VRT2)



VRT-A2a allele (wildtype)

VRT-A2b allele (*polonicum*)

Potential origin of the VRT-A2b allele

CTTTCTCTTACTCCAGATCTGTCCGTTCTCTCGTGTCCCCGACTCGATGC GGATTGGGATCCTCTTGCTGCACGGGCTAGCTCTTGCA CGCAAGCAGTAGGATAAGAGTAGTAGTAGTTTCTCTATACTCTTGATCTGTCCGTTCTCTC
CAGATCTGTCCGTTCTCCAGATCTGTCTACTCCAGATCTGTCCGTTCTCCAGATCTGTCCGTTCTCTCCACTAGCAGTAGGATAAGAGTAGTAGTTTCTGGGCTA

Potential origin of the VRT-A2b allele

CTTTCTCTCTACTCCAGATCTGTCCGTTCTTCTCGTGTCCCCGACTCGATGCGGATTGGGATCCTCTTGCTGCACGGCTAGCTTTGCACGCAAGCAGTAGGATAAGAGTAGTAGTTTCTCCTAATCTTGATCTGCCGTTCTCTC
CAGATCTGTCCGTTCTTCTCAGATCTGTCCCTACTCCAGATCCTACTCCAGATCTGTCCGTTCTCCAGATCTGTCCGTTCTCTOCACTAGCAGTAGGATAAGAGTAGTAGTTTCTGGGCTA

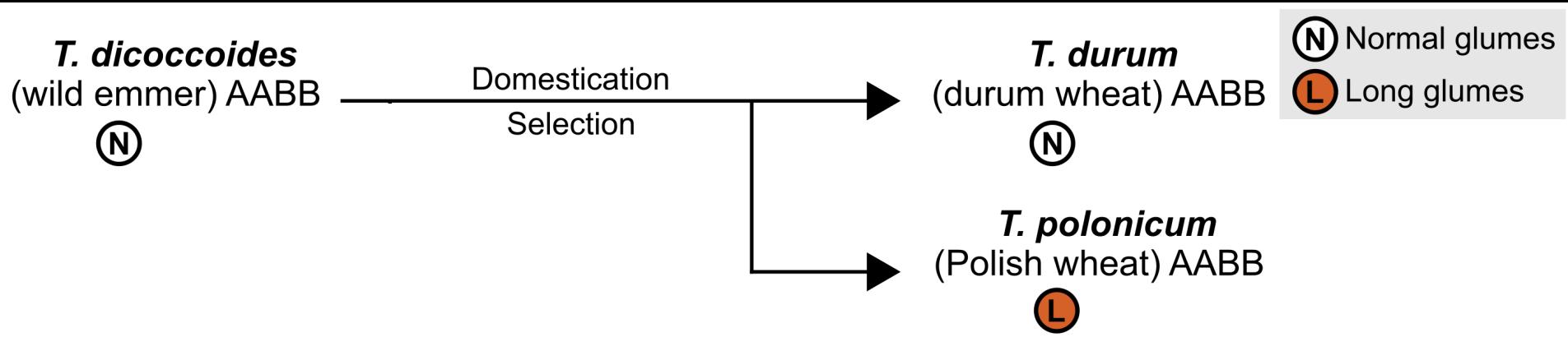
Potential origin of the *VRT-A2b* allele

CTTTCTCTCTACTCCAGATCTGTCCGTTCTTCTCGTGTCCCCGACTCGATGC GGATT CGGGAT CCTCTTGCTGCACGGCTAGCTTTGCACGCAAGCAGTAGGATAAGAGTAGTAGTAGTTTCTCCTA ACTCTTGATCTGTCCGTTCTC
CAGATCTGTCCGTTCTTCTCAGATCTGTCCCTACTCCAGATCCTACTCCAGATCTGTCCGTTCTCCAGATCTGTCCGTTCTTCTCCTACTAGCAGTAGGATAAGAGTAGTAGTTTCTGGGCTA

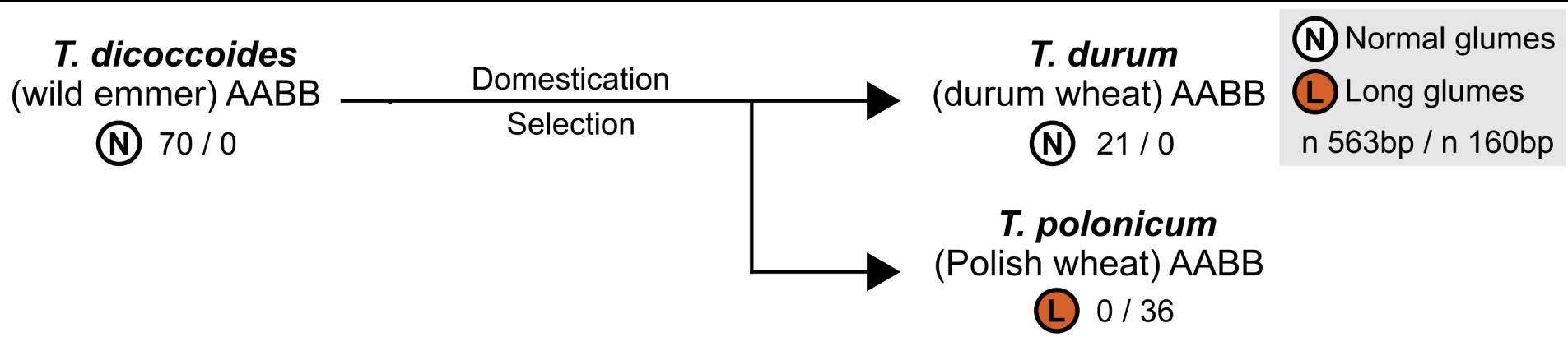
CTTGACGCAAGCAGTAGGATAAGAGTAGTAGTTTCTCCTA ACTCTTGATCTGTCCGTTCTTCTCCAGATCTGTCCGTTCTTCTCCAGATCTGTCCCTACTCCAGATCCTACTCCAGATCTGTCCGTTCTTCTCCAGATCTGTCCGTTCTTCTC
AGCAGTAGGATAAGAGTAGTAGTAGTTTCT

TACTCTT GATCTGTCCGTTCTTCTC
CAGATCTGTCCGTTCTTCTC
CAGATCTGTCC
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TACTCCAGATCTGTCCGTTCTTCTC
CAGATCTGTCCGTTCTTCTC

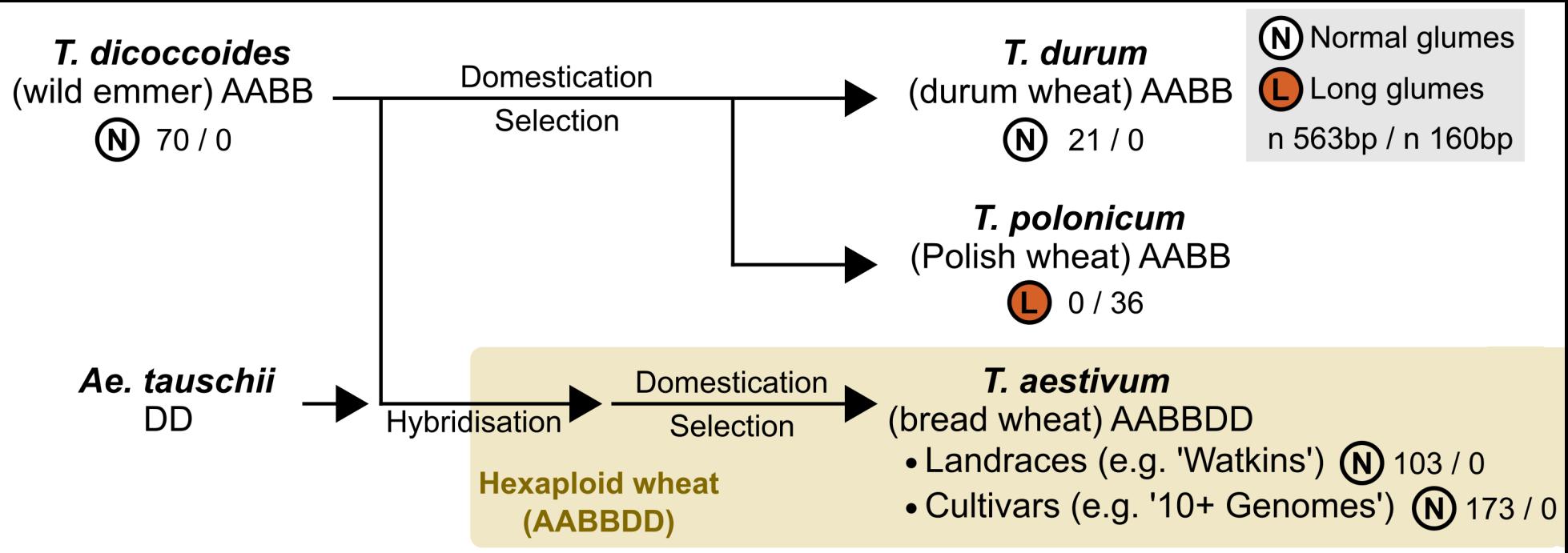
Distribution of *P1* allele



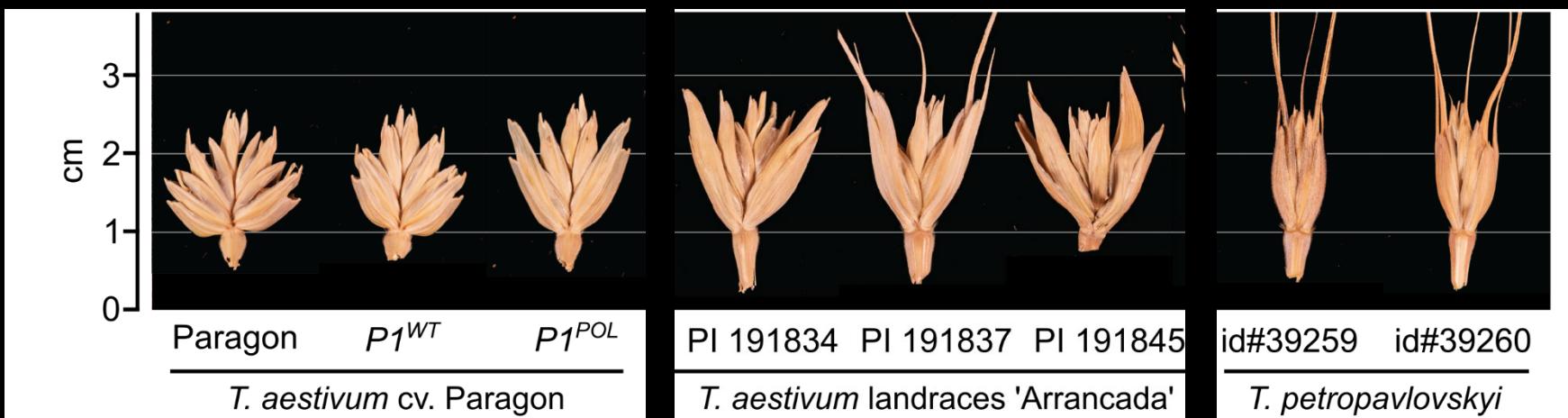
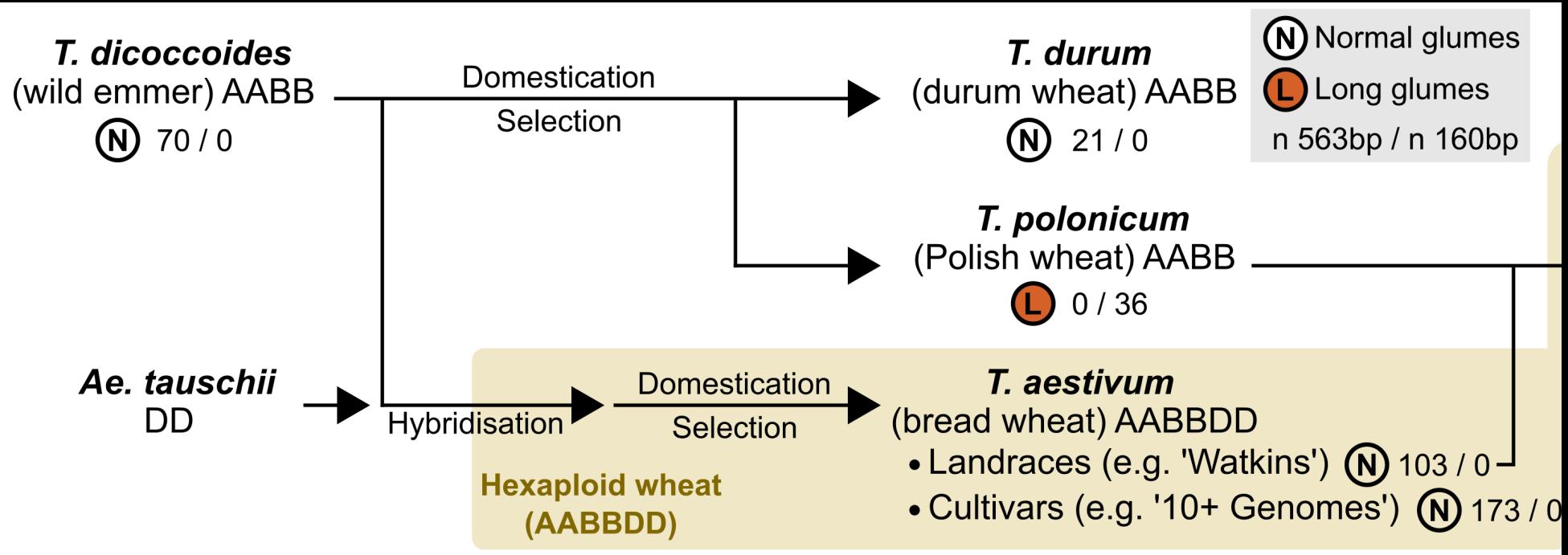
Distribution of *P1* allele



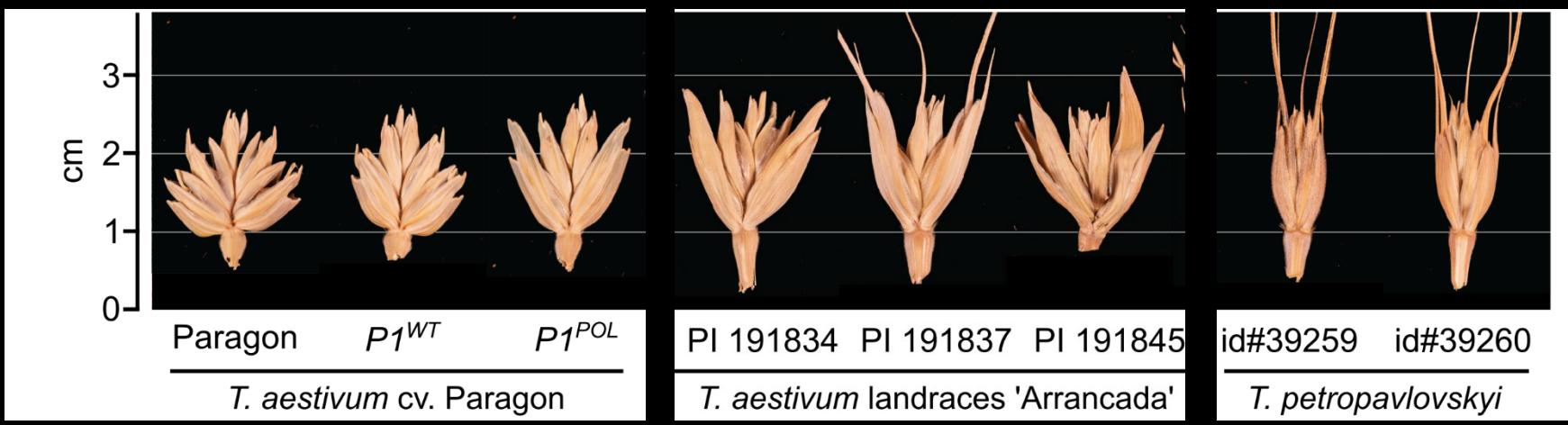
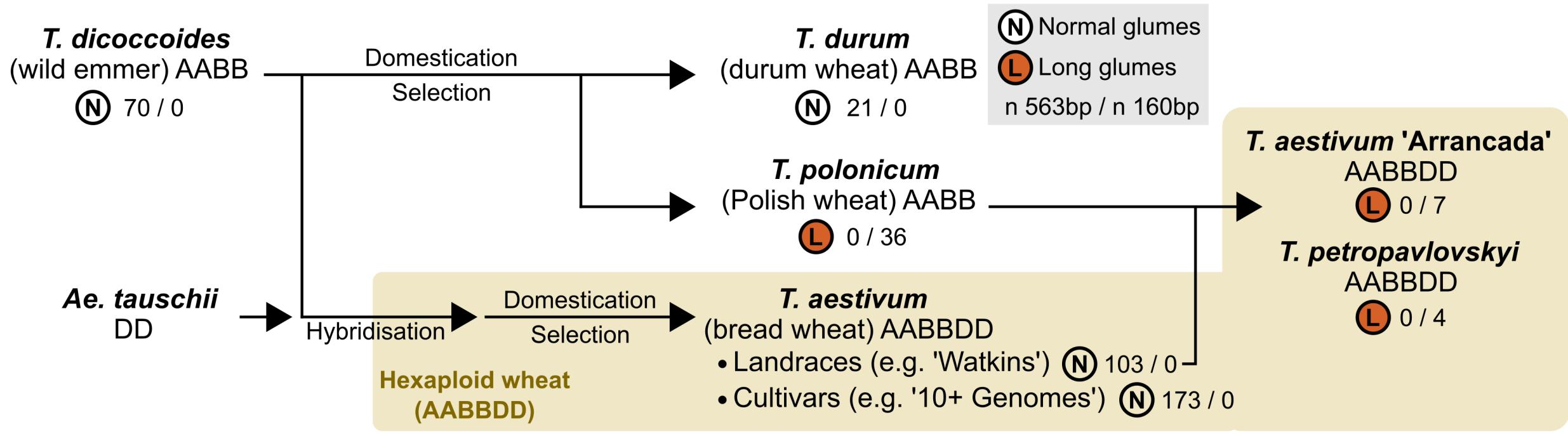
Distribution of *P1* allele



Distribution of *P1* allele

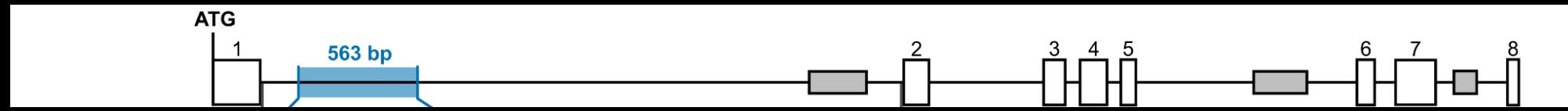


Distribution of *P1* allele

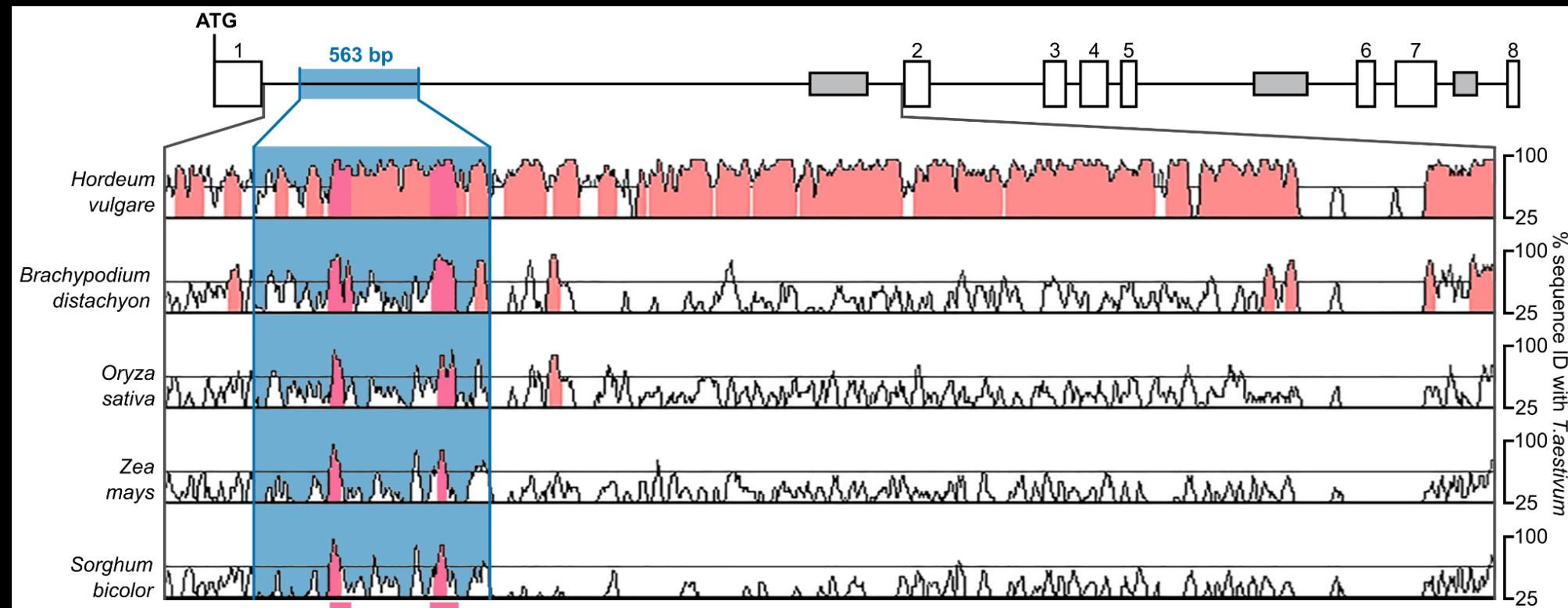


Adamski et al., 2021
Liu et al., 2021
Chai et al., 2021
Xiao et al., 2021

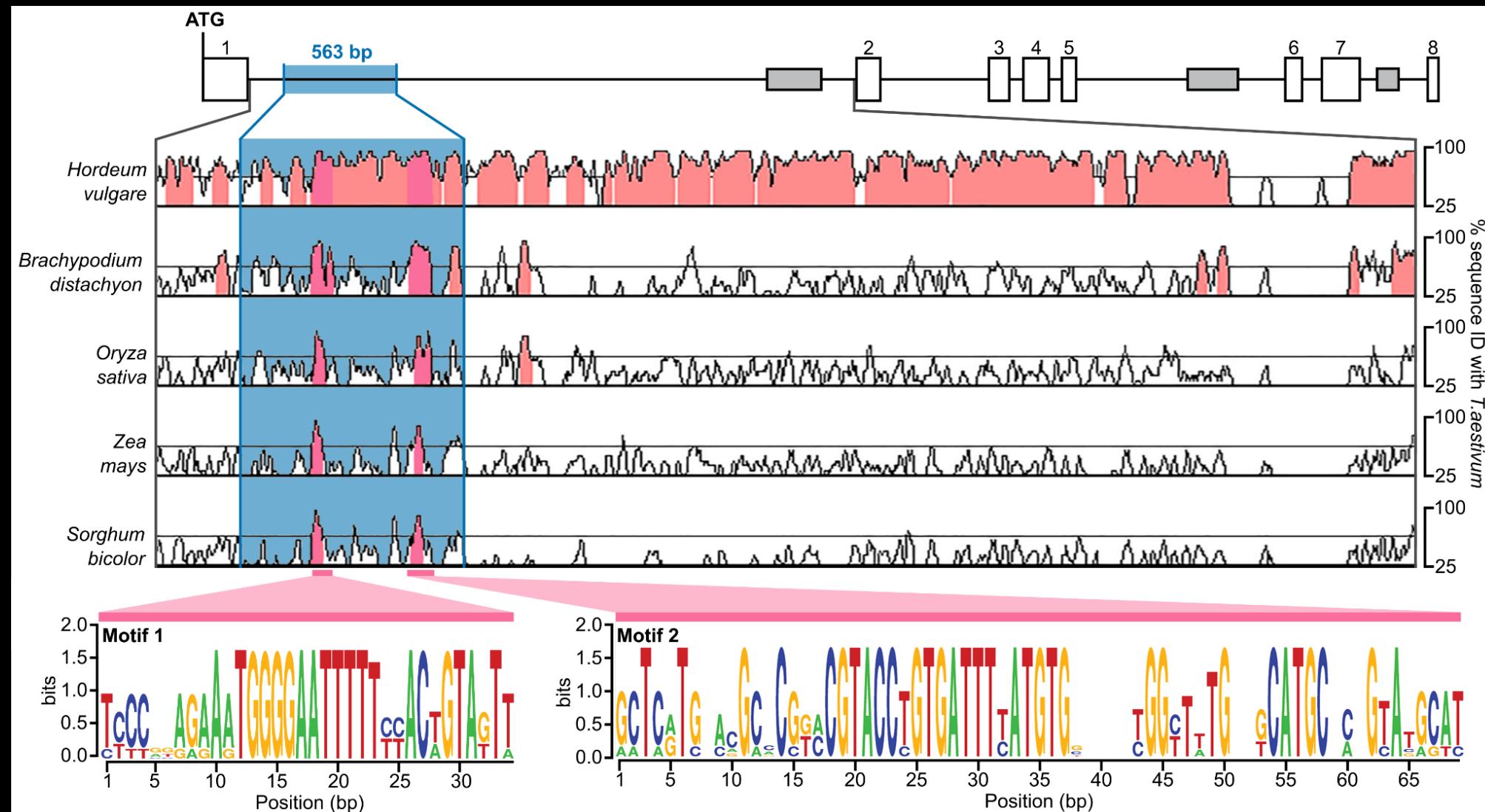
Conservation of VRT-A2a intron 1 sequence across grasses



Conservation of VRT-A2a intron 1 sequence across grasses

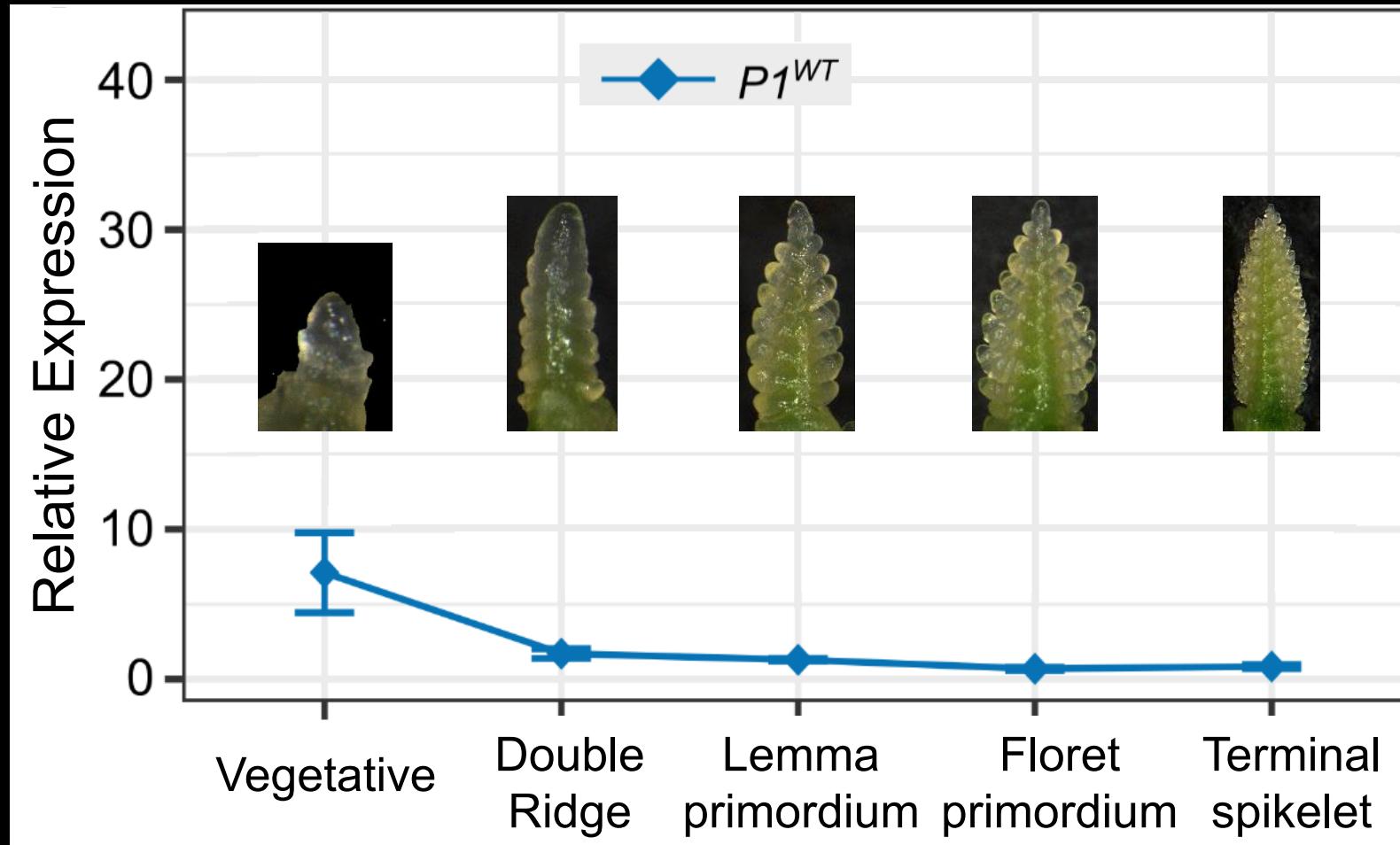


Conservation of VRT-A2a intron 1 sequence across grasses

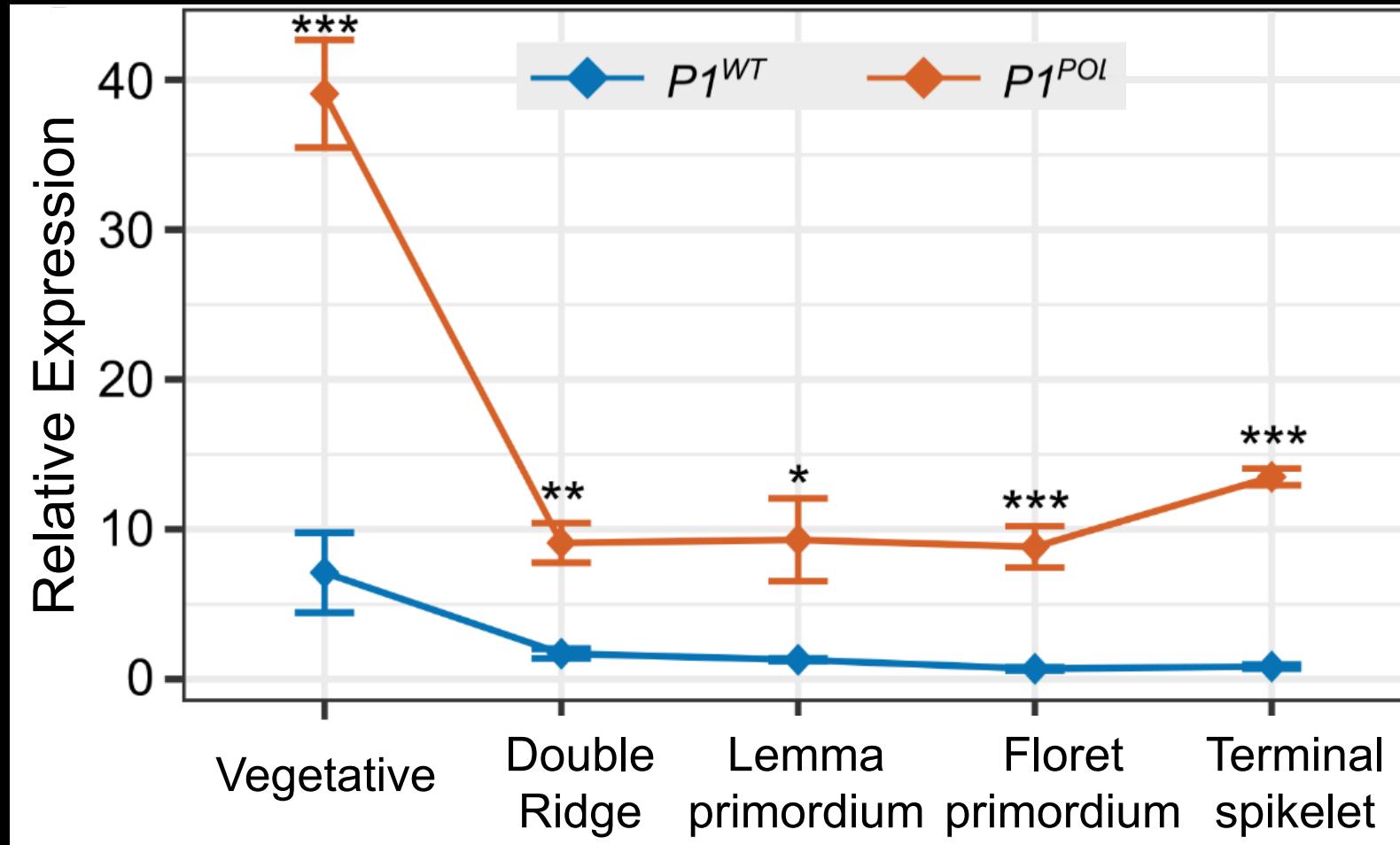


Motifs absent from *T. polonicum*

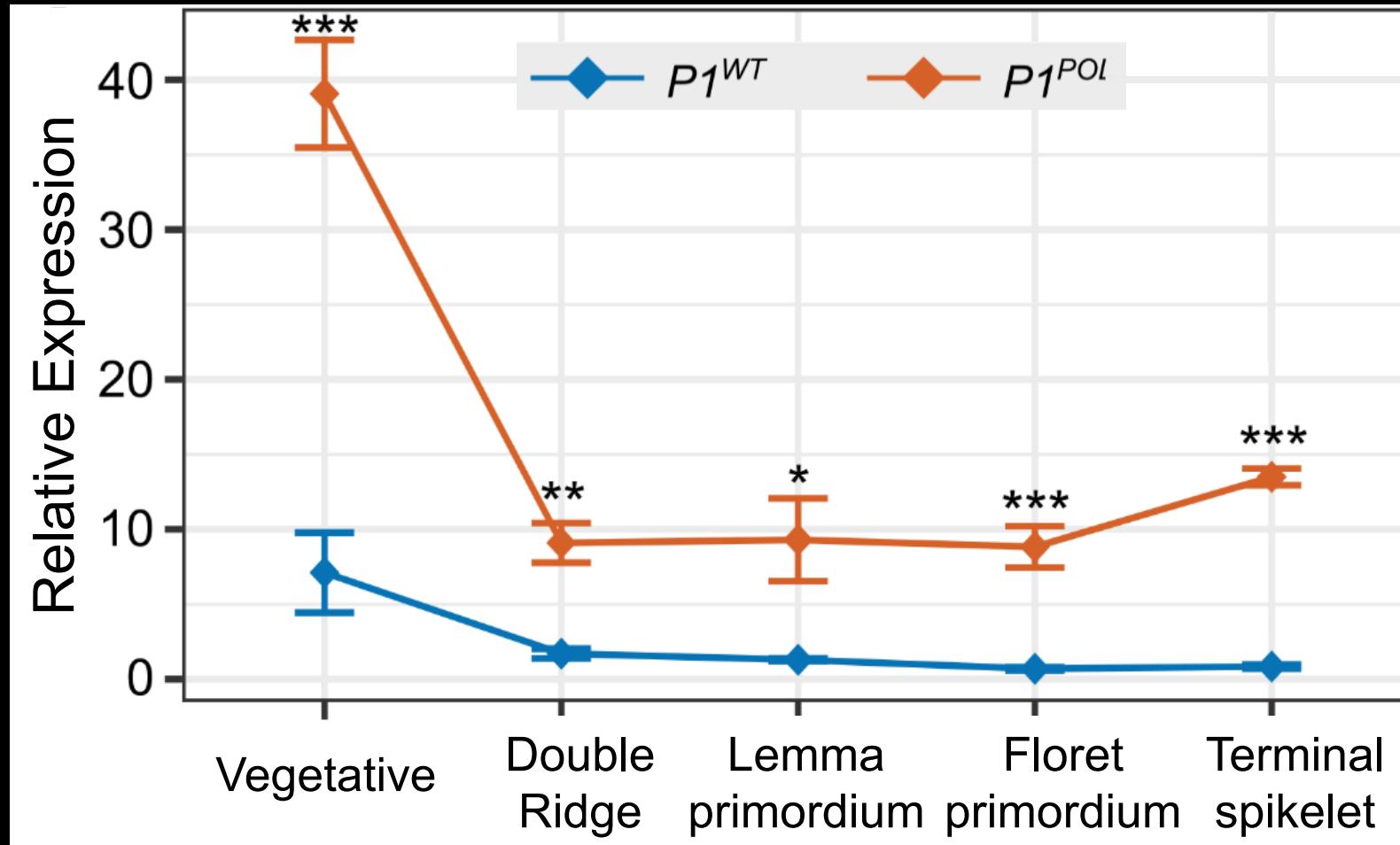
The *VRT-A2b* allele is misexpressed in developing spikes



The *VRT-A2b* allele is misexpressed in developing spikes

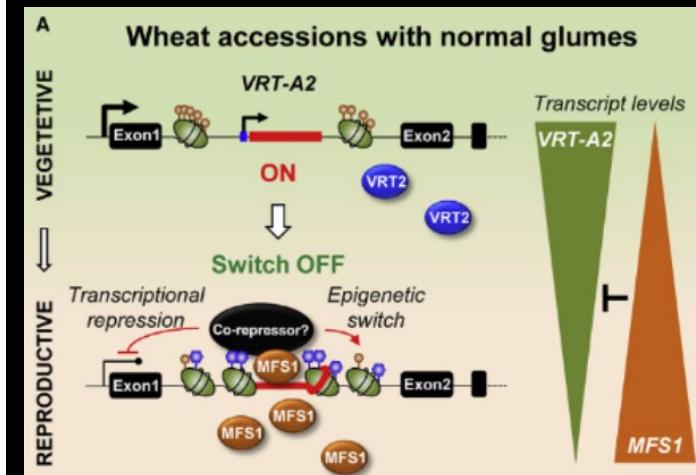
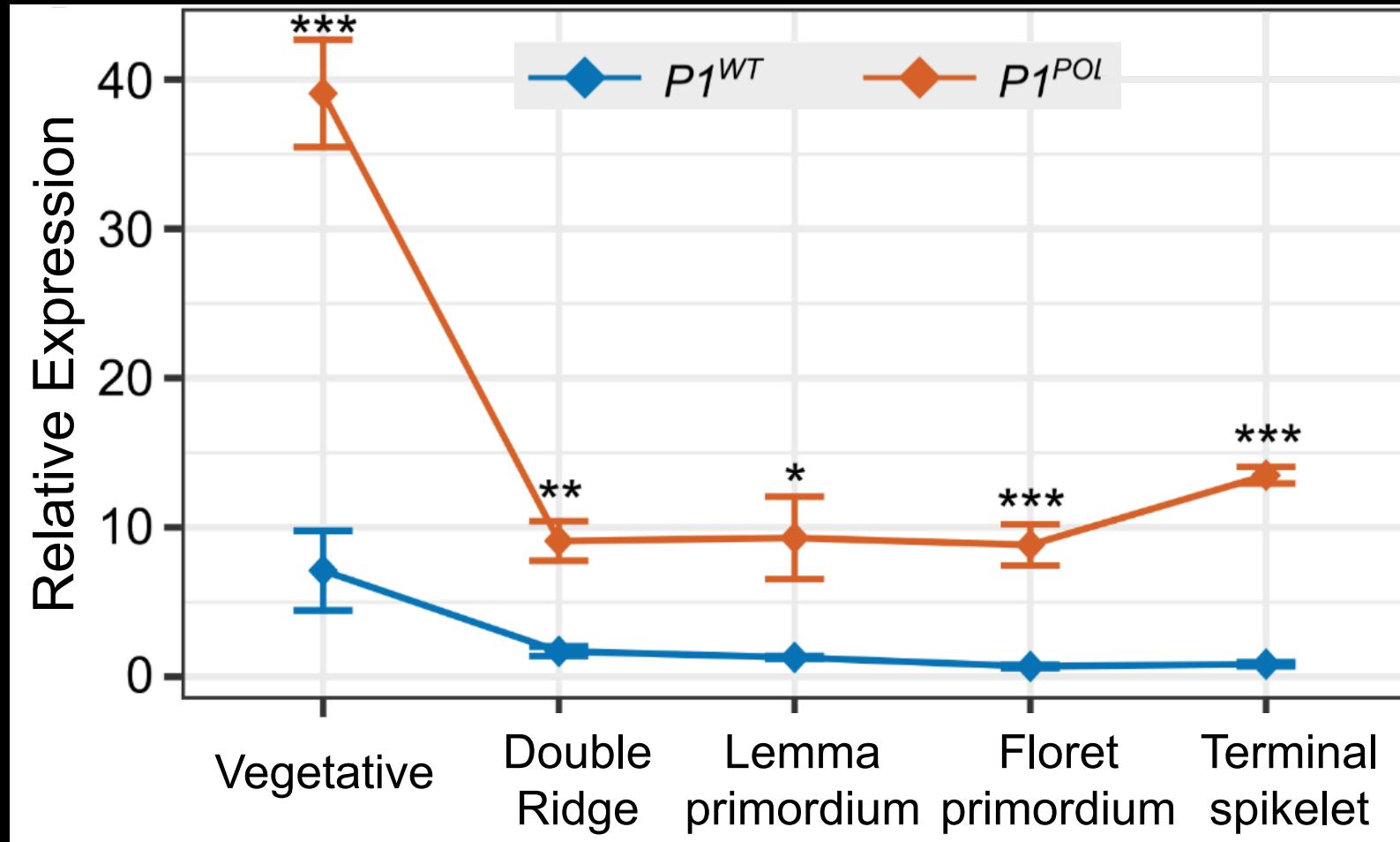


The *VRT-A2b* allele is misexpressed in developing spikes



The deleted motifs seem to act as binding sites for repressors of *VRT-A2* transcription.

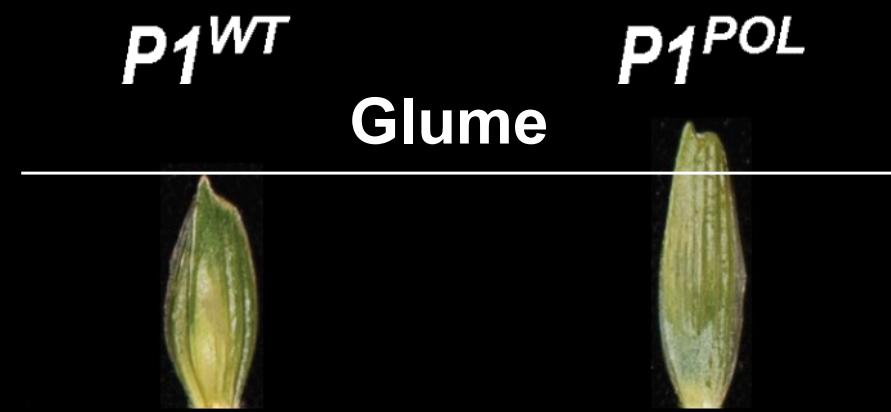
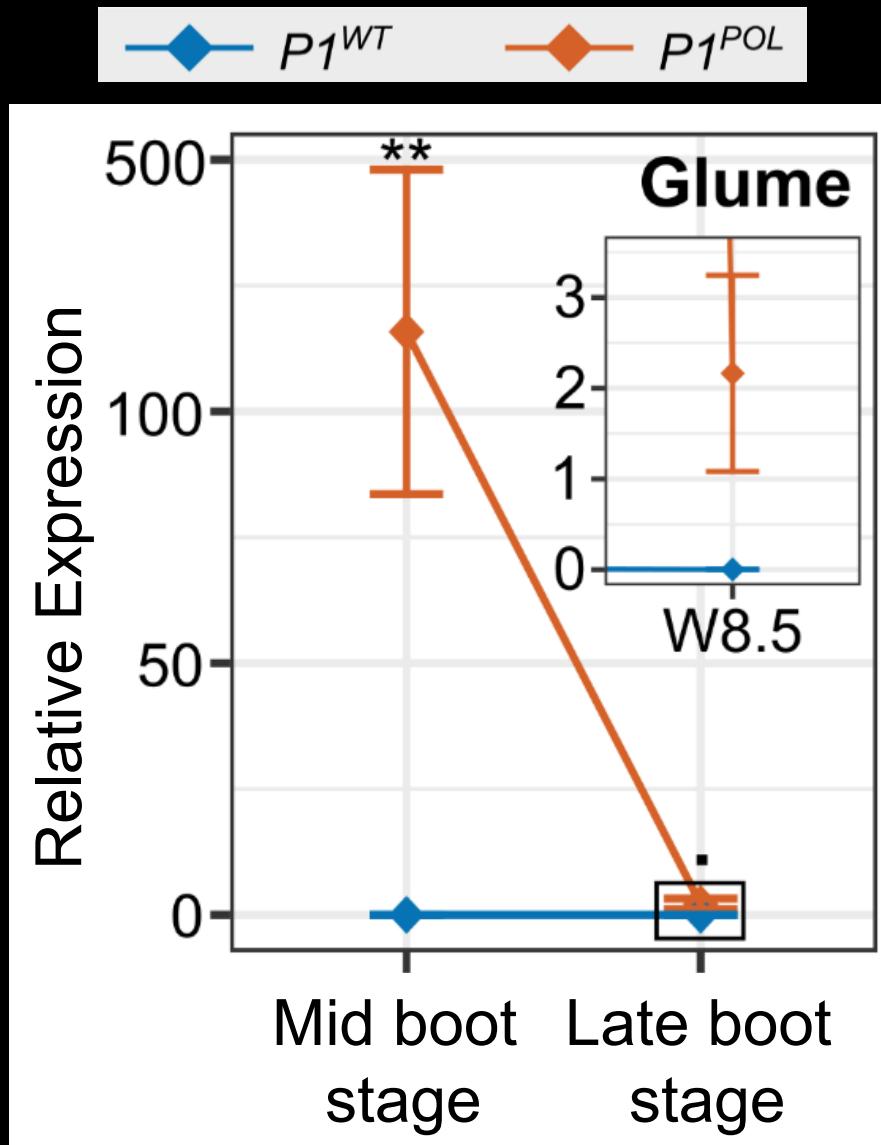
The *VRT-A2b* allele is misexpressed in developing spikes



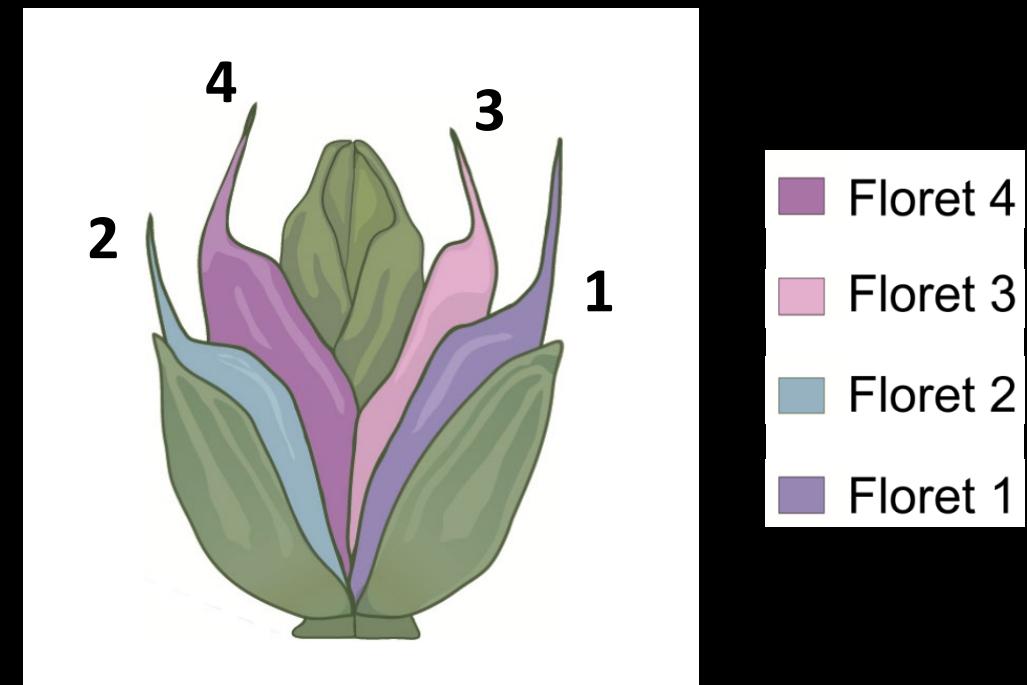
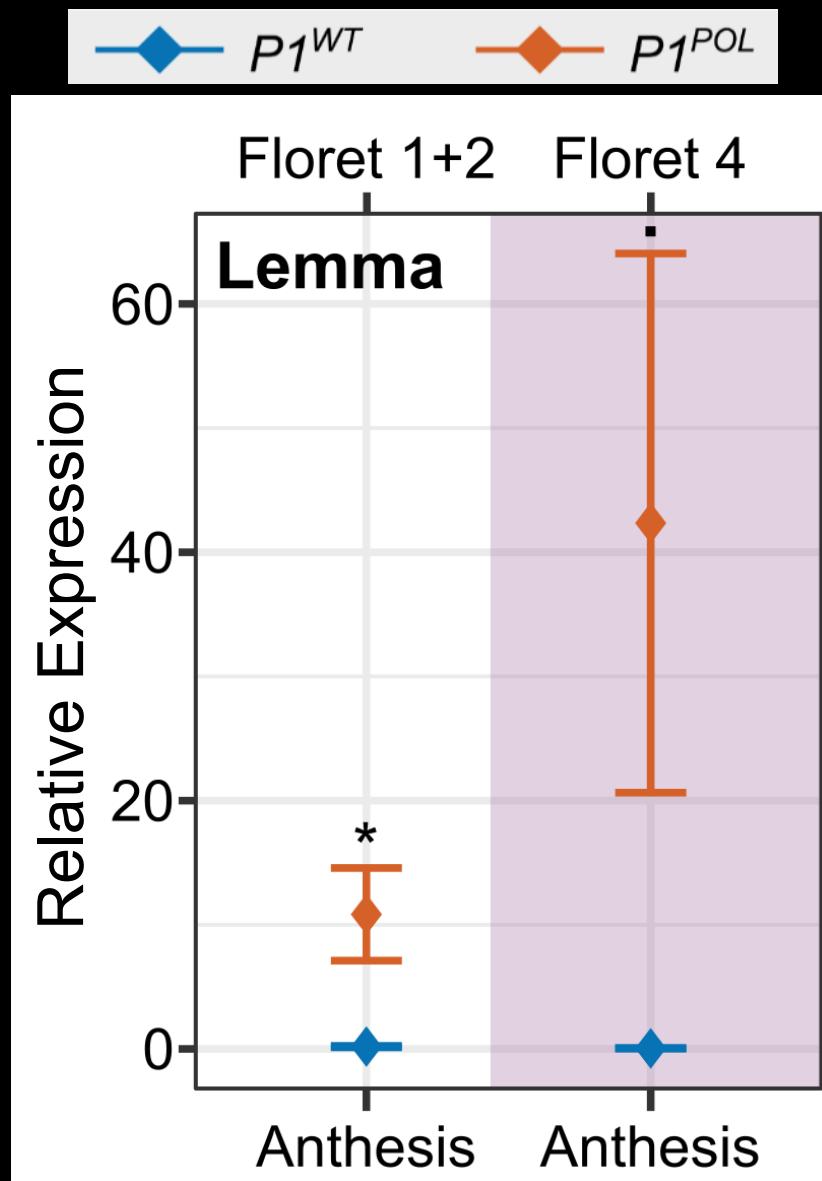
Liu et al., 2021

The deleted motifs seem to act as binding sites for repressors of *VRT-A2* transcription.

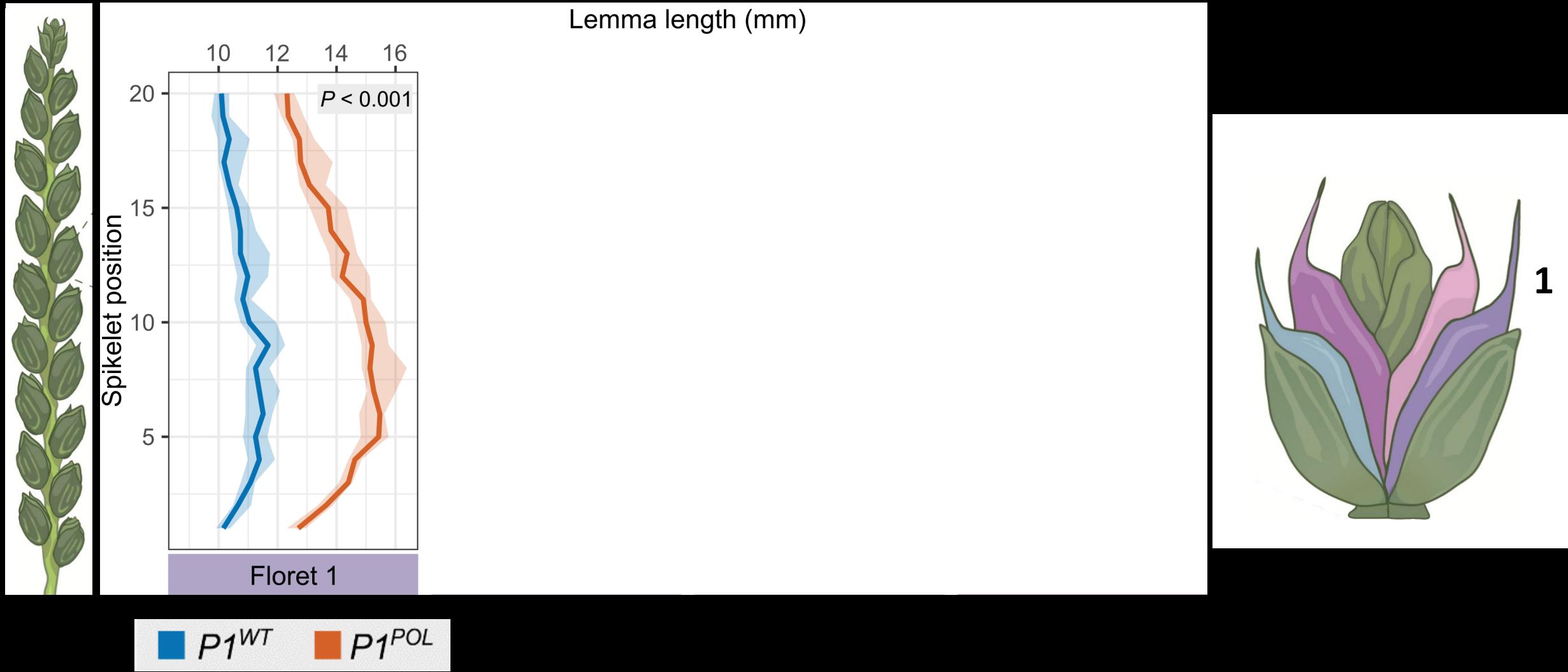
VRT-A2b is ectopically expressed in glumes



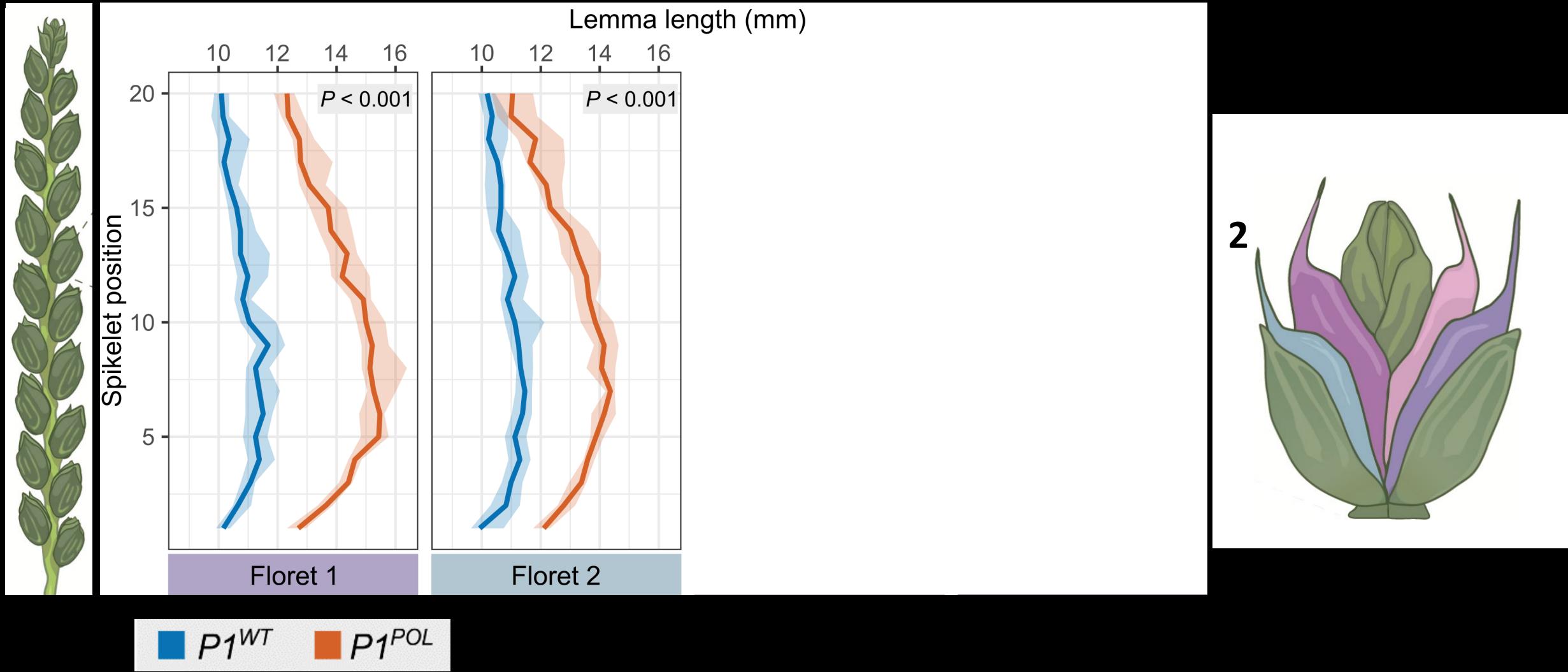
VRT-A2b is ectopically expressed in lemmas



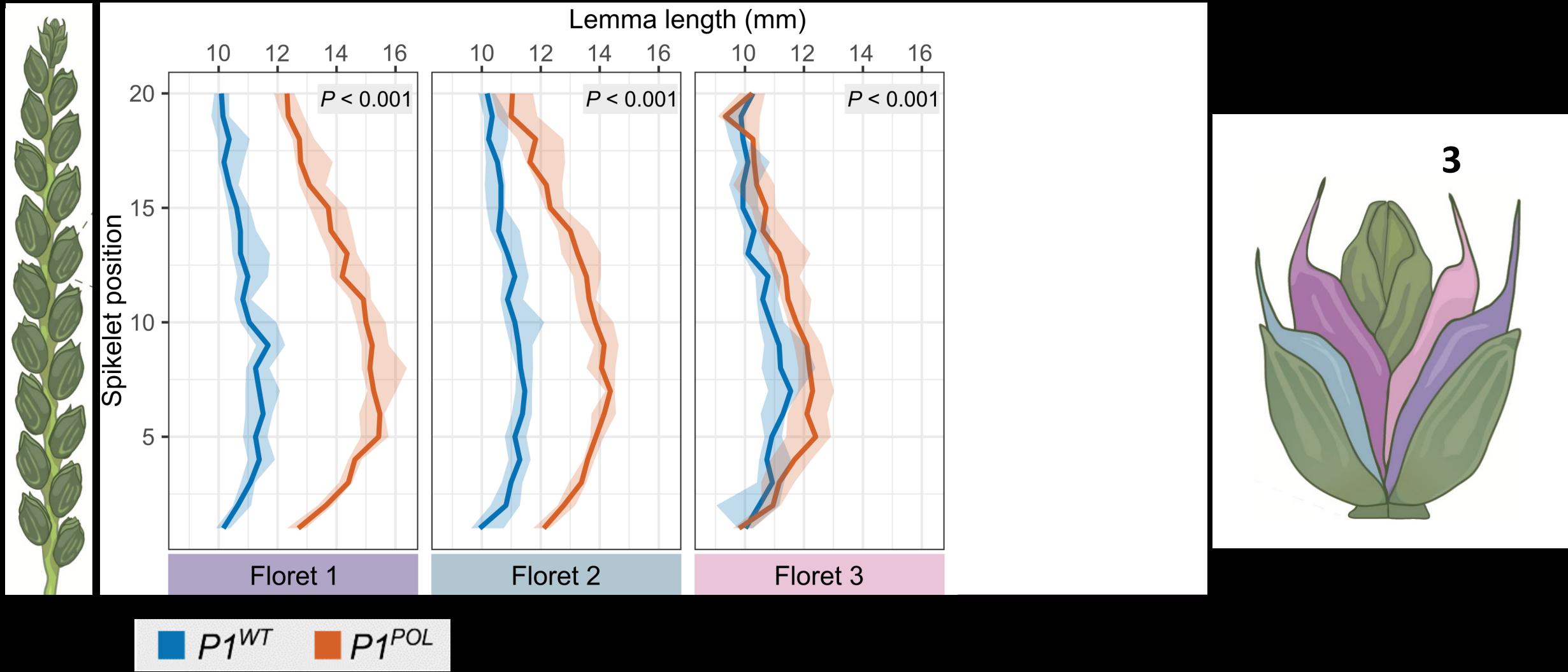
***VRT-A2b* affects outer and basal organs of spikelets**



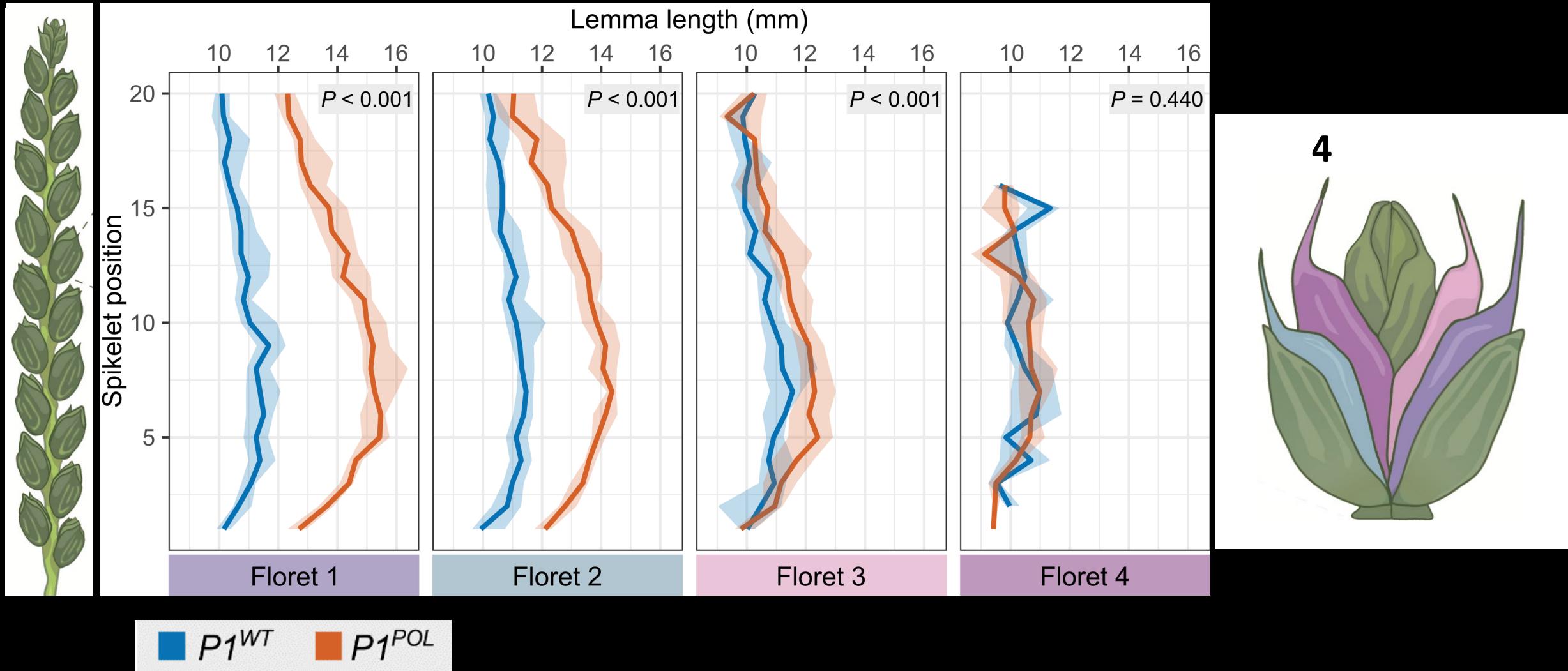
***VRT-A2b* affects outer and basal organs of spikelets**



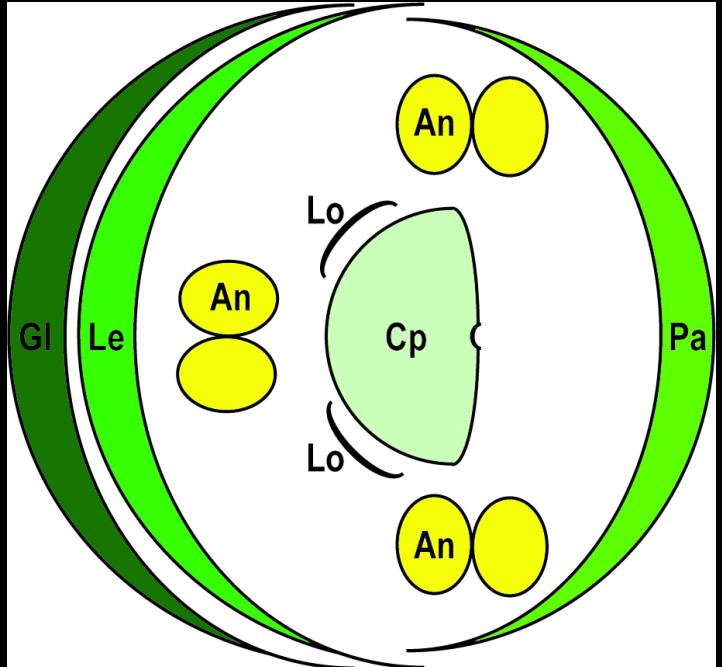
***VRT-A2b* affects outer and basal organs of spikelets**



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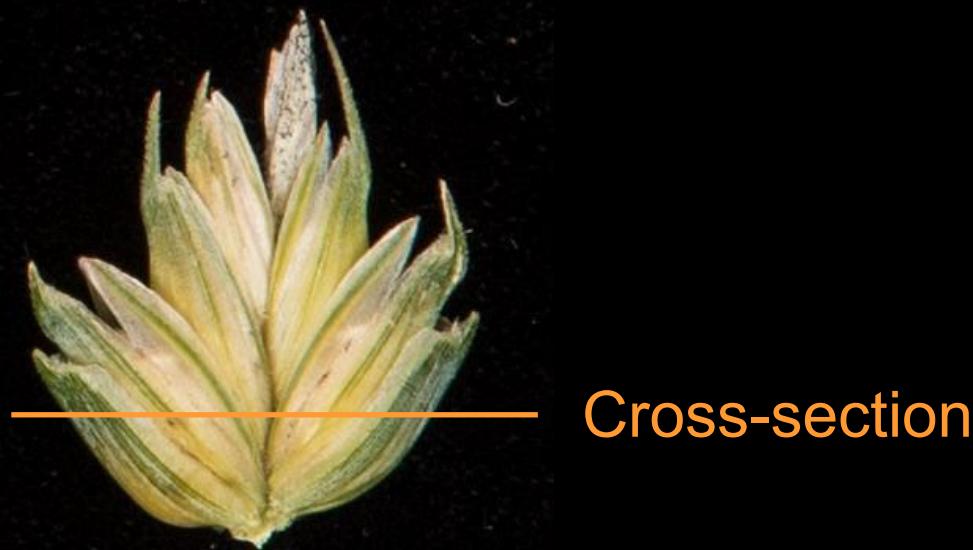
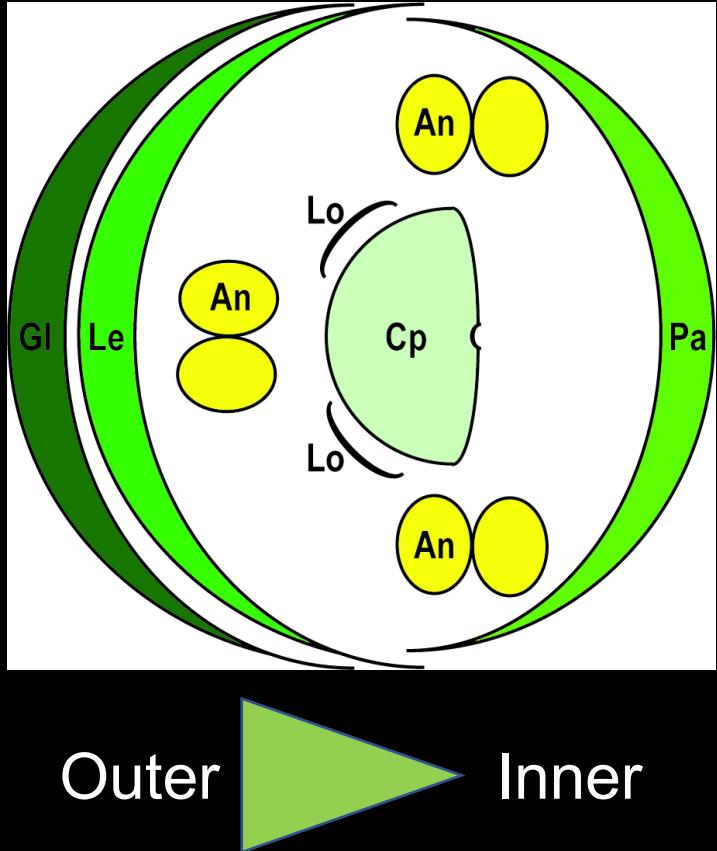


VRT-A2b affects outer and basal organs of spikelets



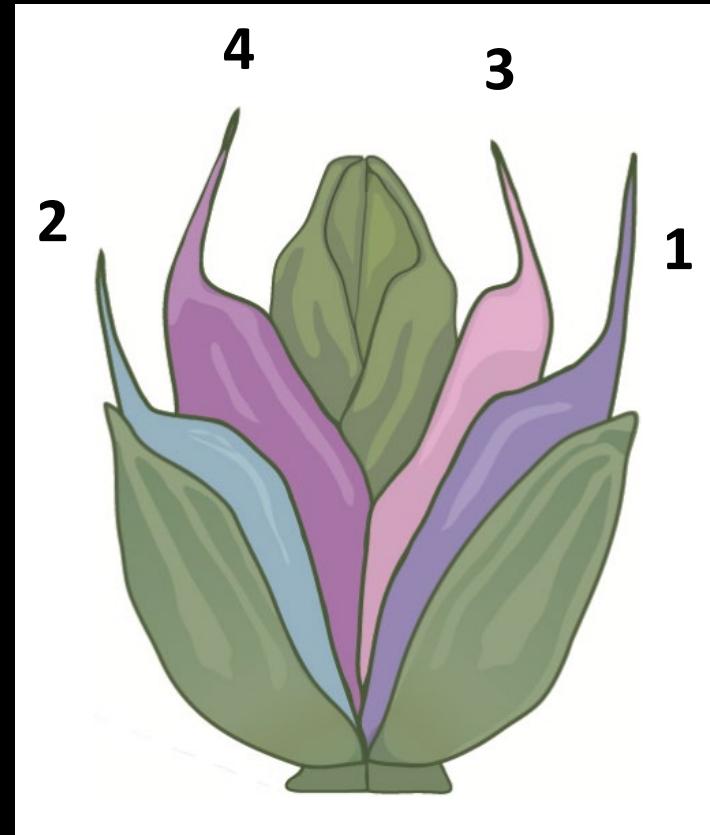
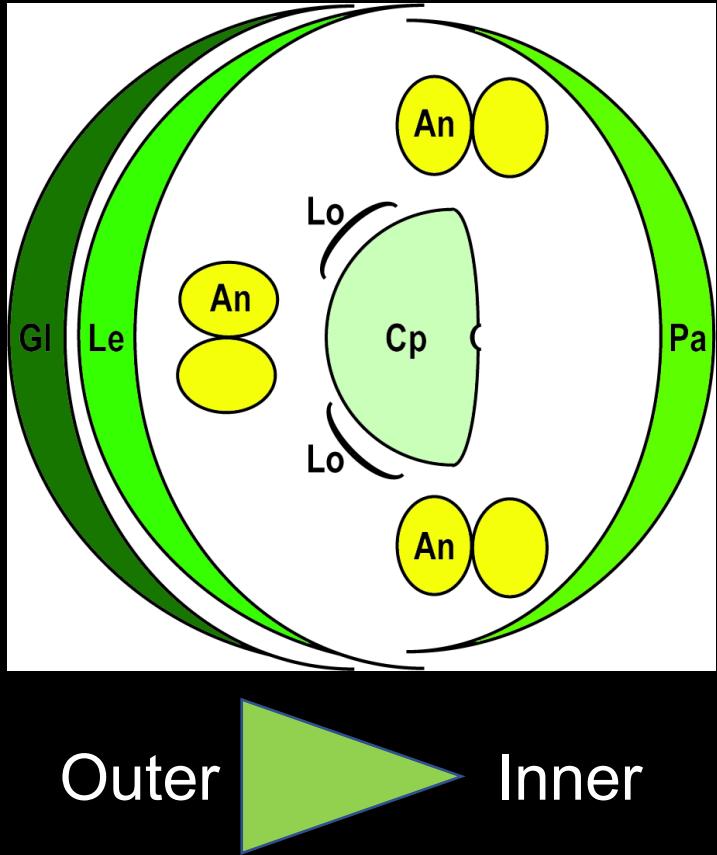
Cross-section

VRT-A2b affects outer and basal organs of spikelets

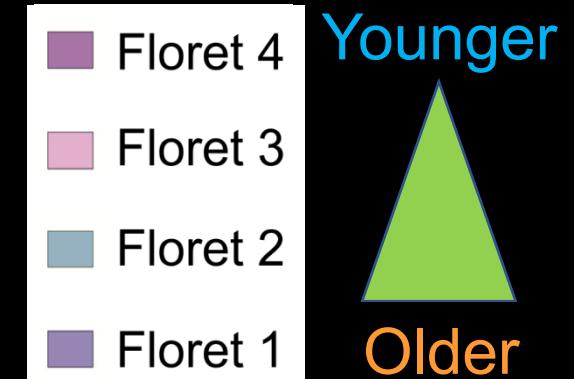
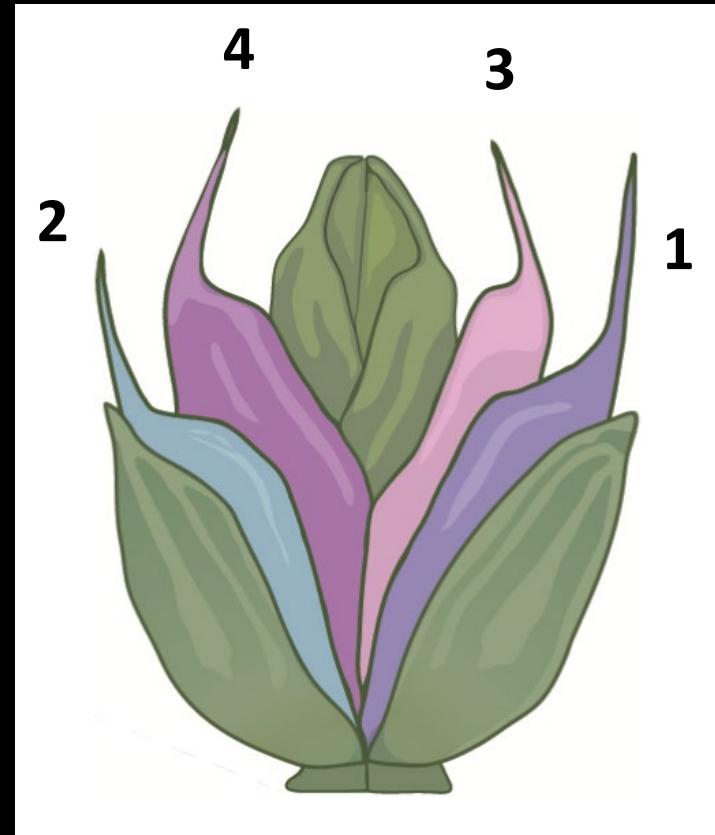
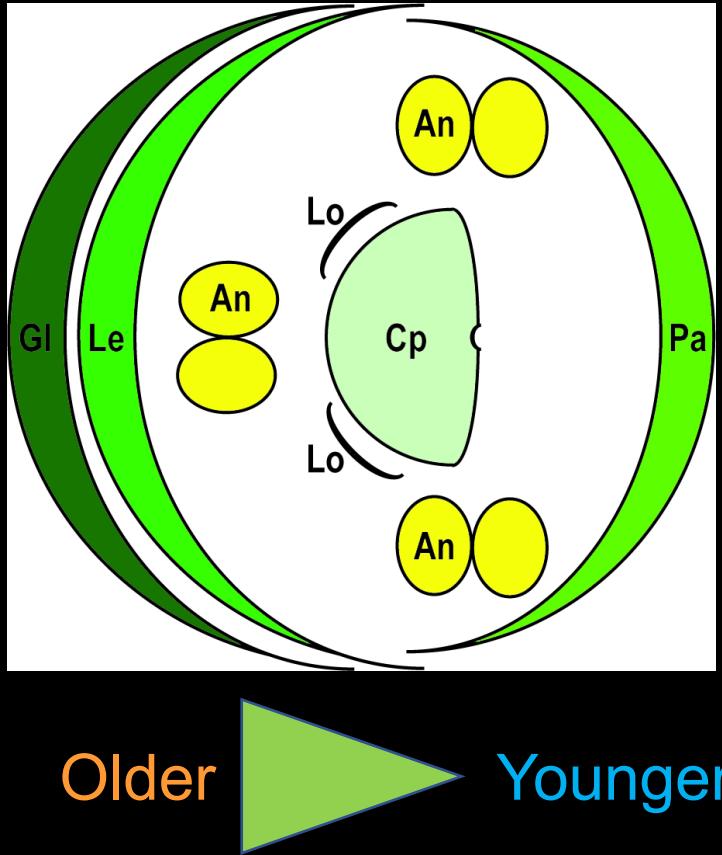


Cross-section

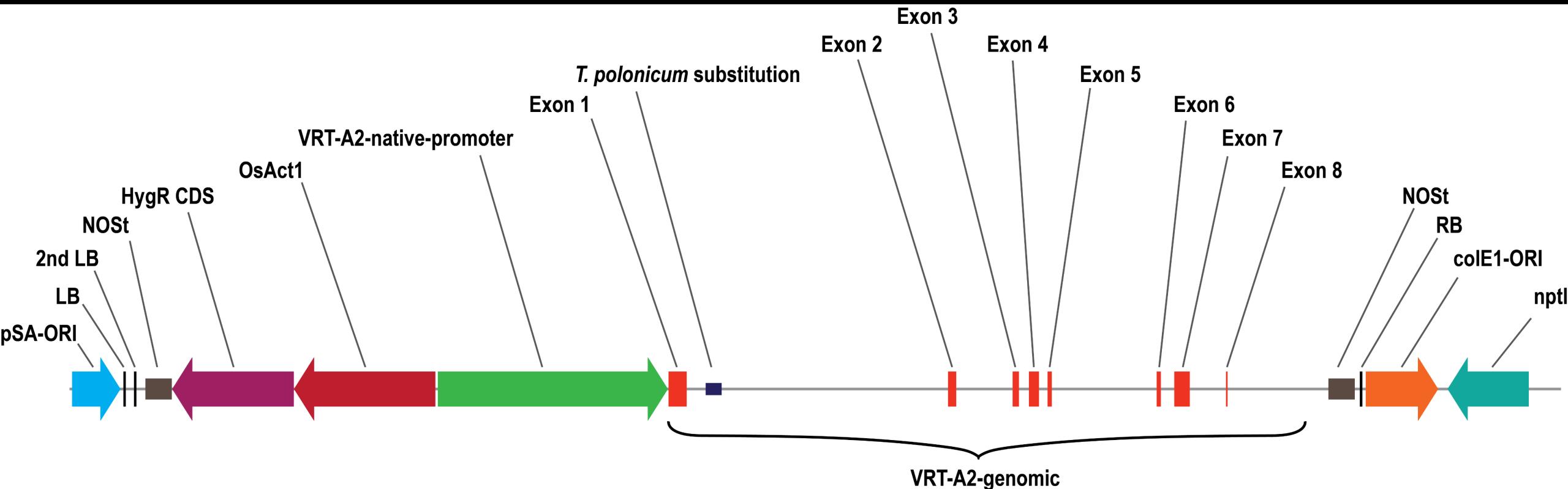
VRT-A2b affects outer and basal organs of spikelets



VRT-A2b affects outer and basal organs of spikelets



Complementation of “Fielder” with *VRT-A2b* allele



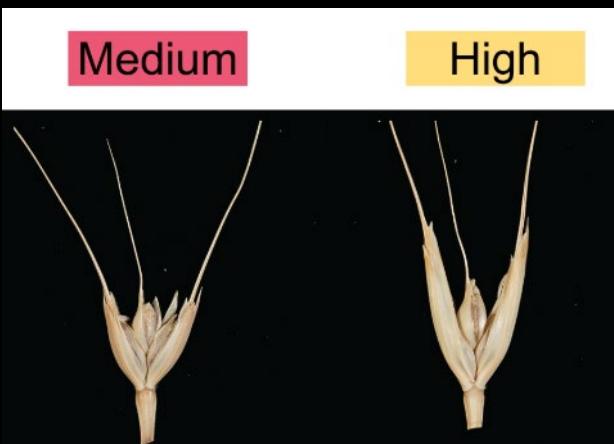
Sadiye Hayta



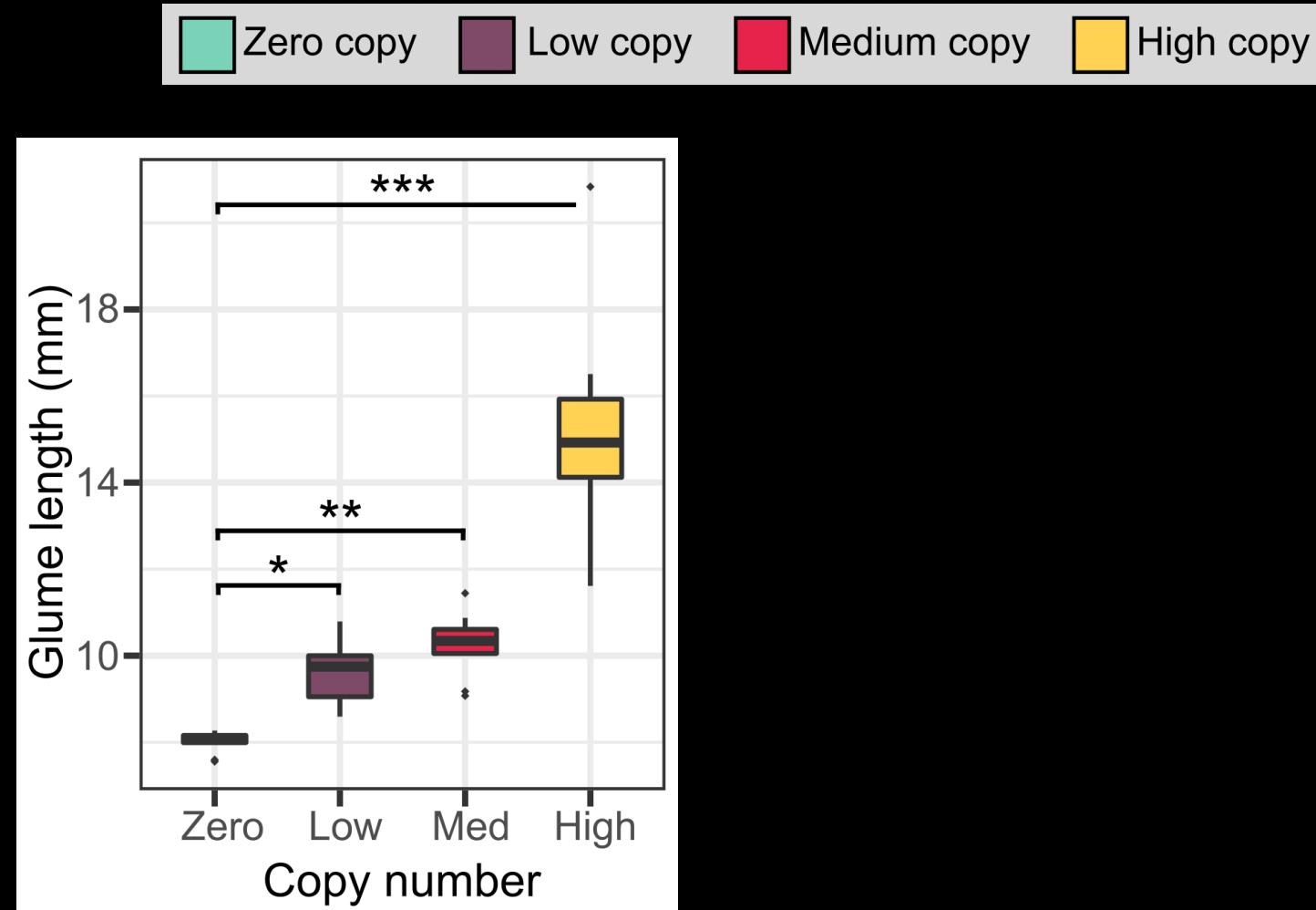
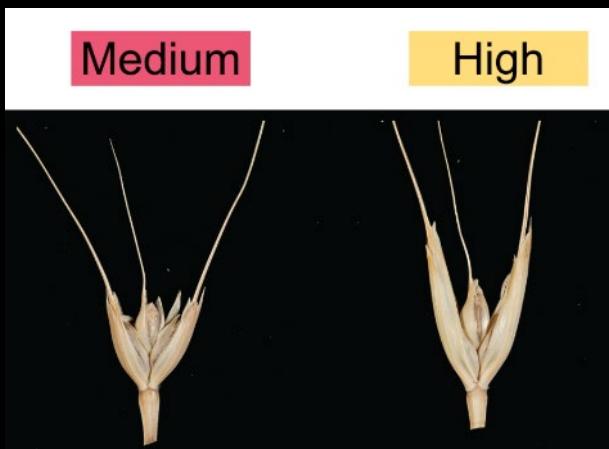
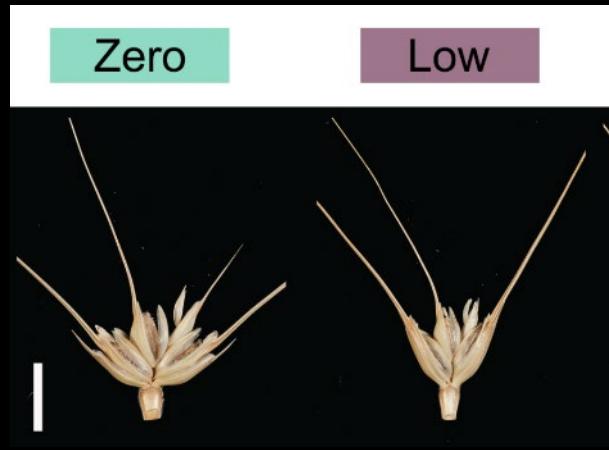
Mark Smedley

Hayta et al., 2019

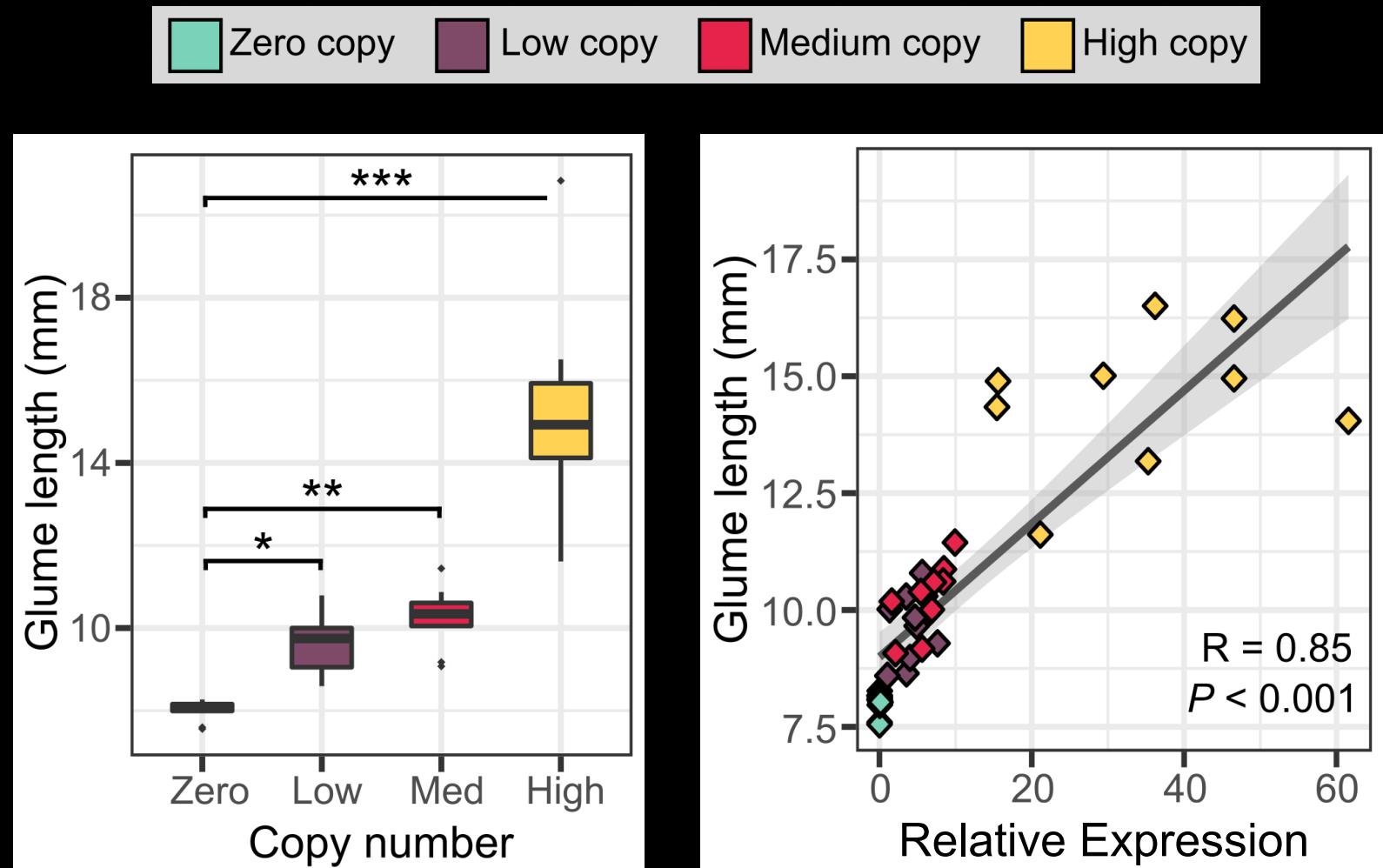
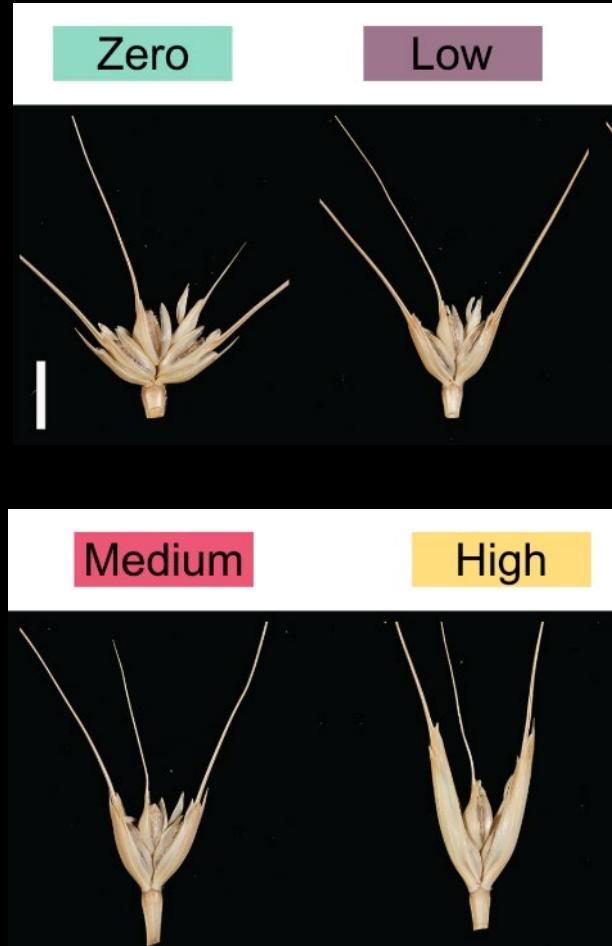
VRT-A2b acts in a dosage-dependent manner in T1 lines



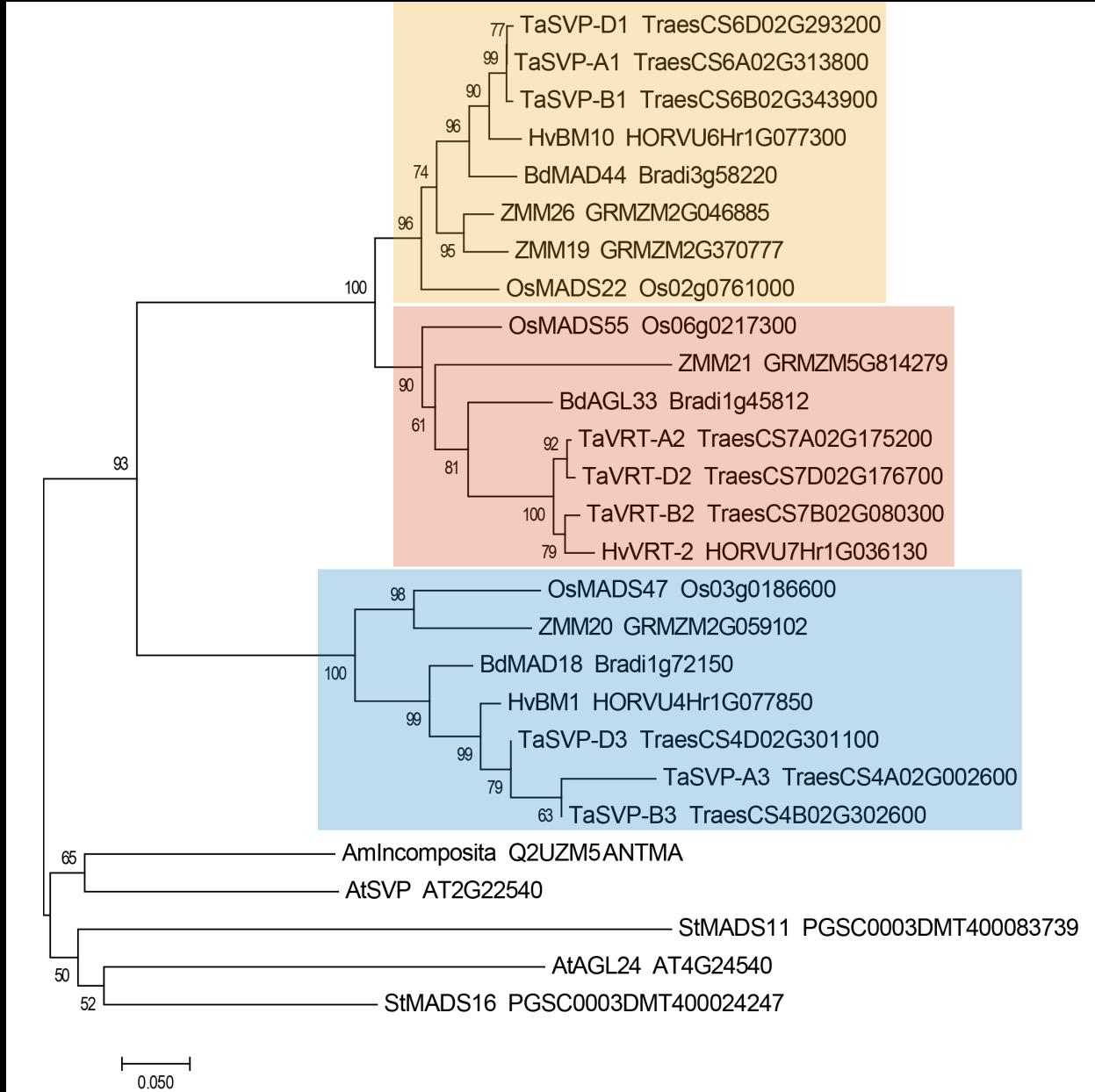
VRT-A2b acts in a dosage-dependent manner in T1 lines



VRT-A2b acts in a dosage-dependent manner in T1 lines

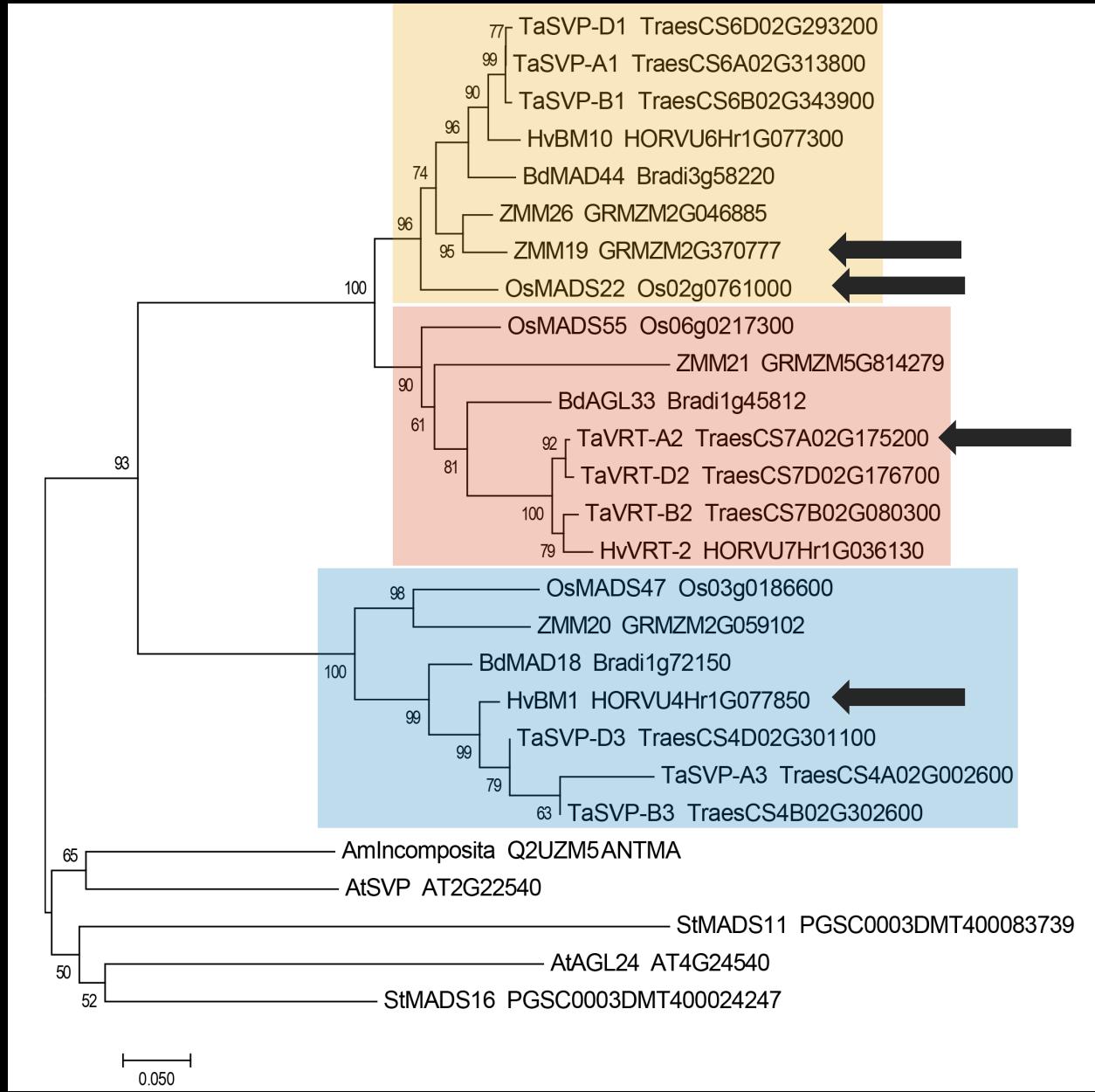


SVP family in grasses



Triplification of *SVP* genes in grasses

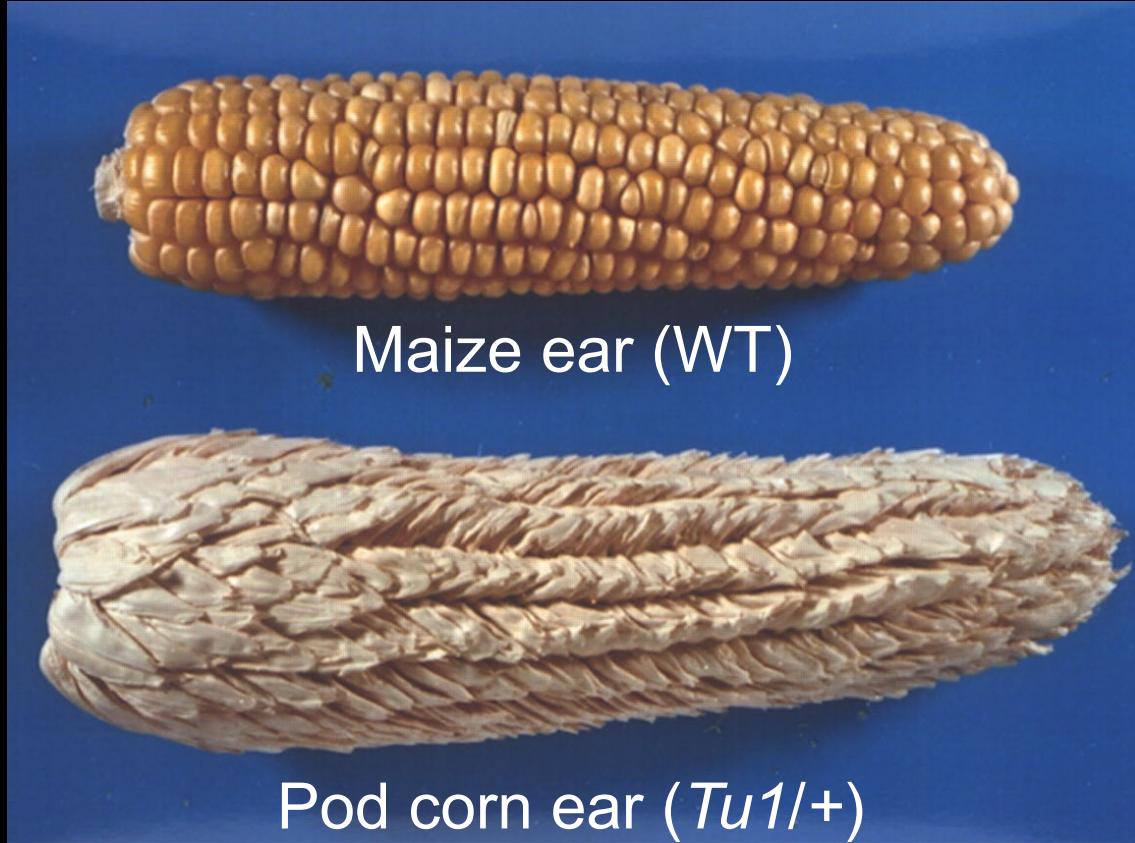
SVP family in grasses



SVP members shown to cause elongated glume or lemma.

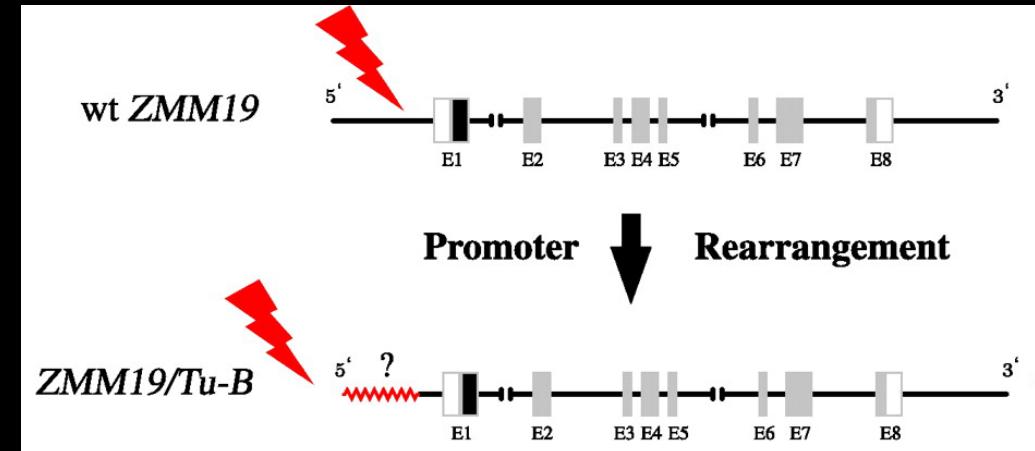
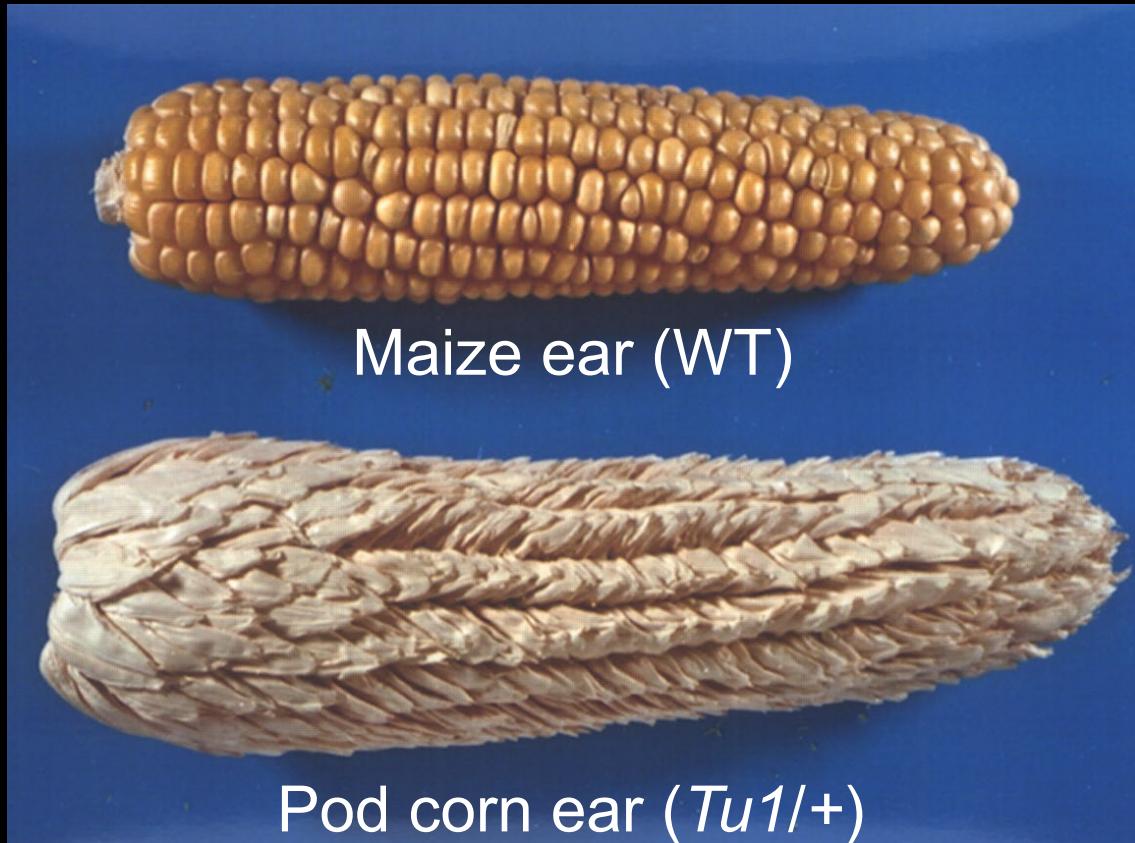
Sentoku et al., 2005; Trevaskis et al., 2007;
Wingen et al., 2012; Han et al., 2012

VRT-A2b reminiscent of “pod corn” allele in maize



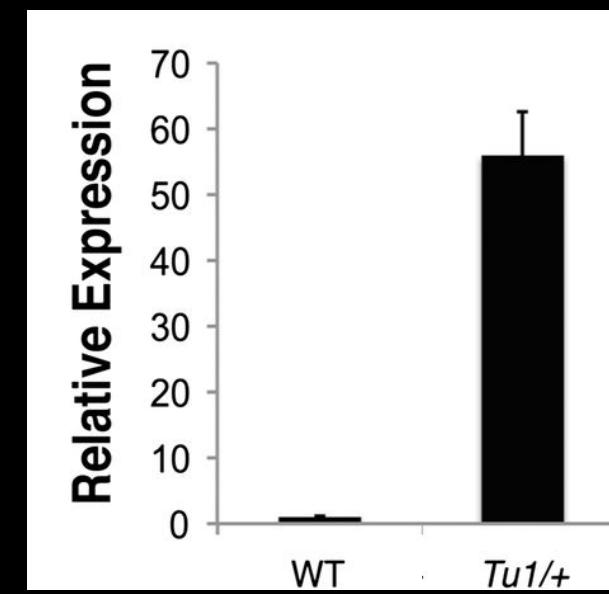
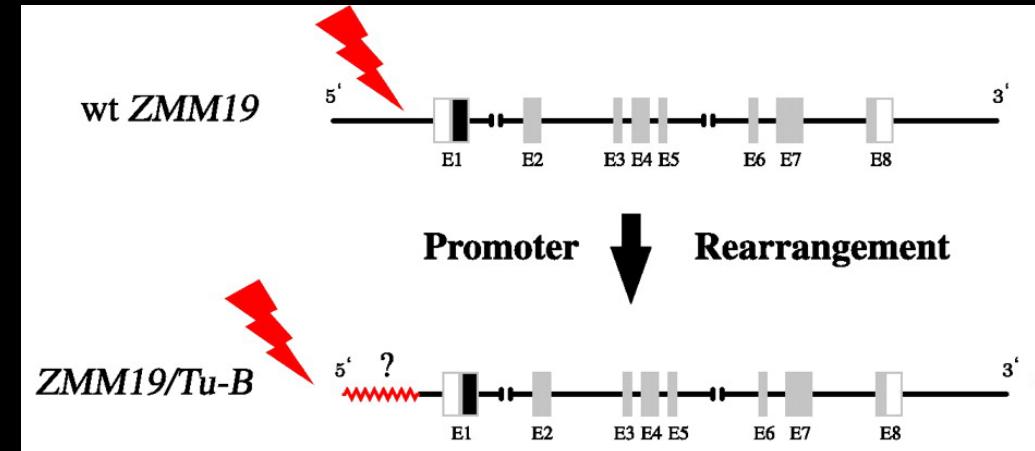
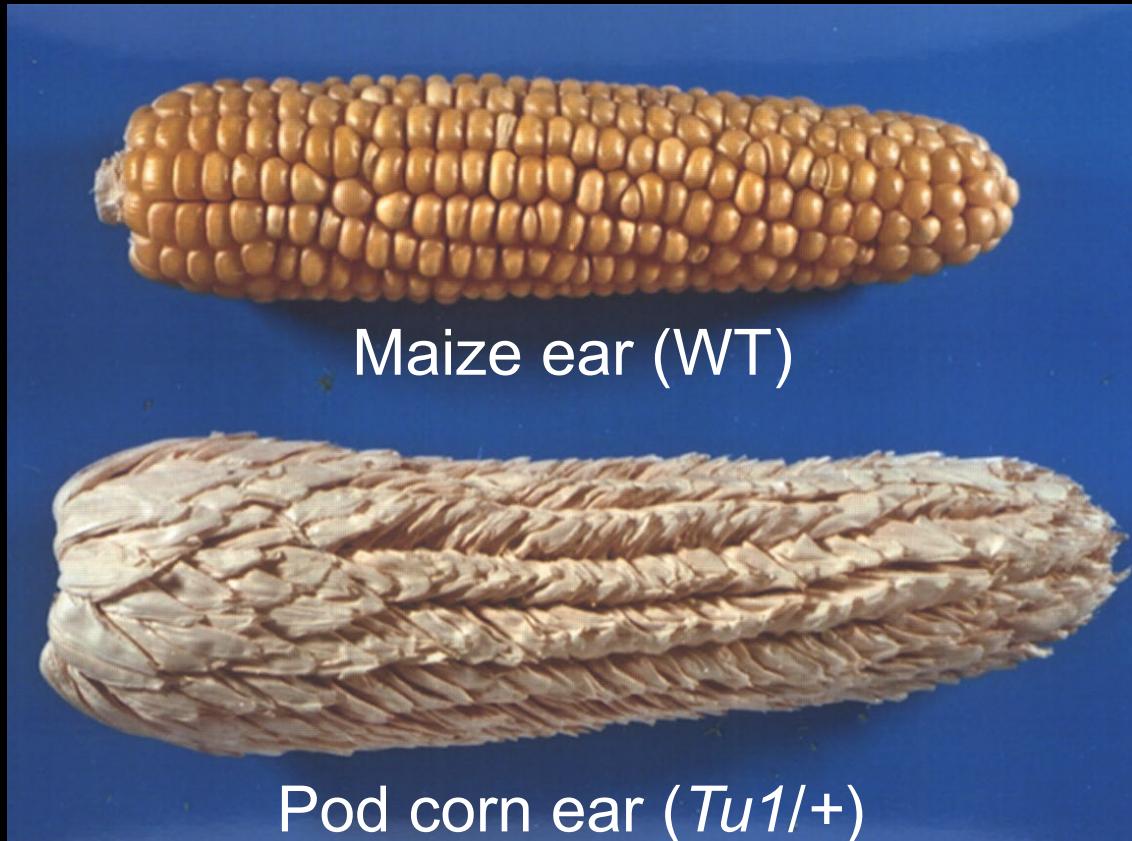
Wingen *et al.*, 2012; Han *et al.*, 2012

VRT-A2b reminiscent of “pod corn” allele in maize



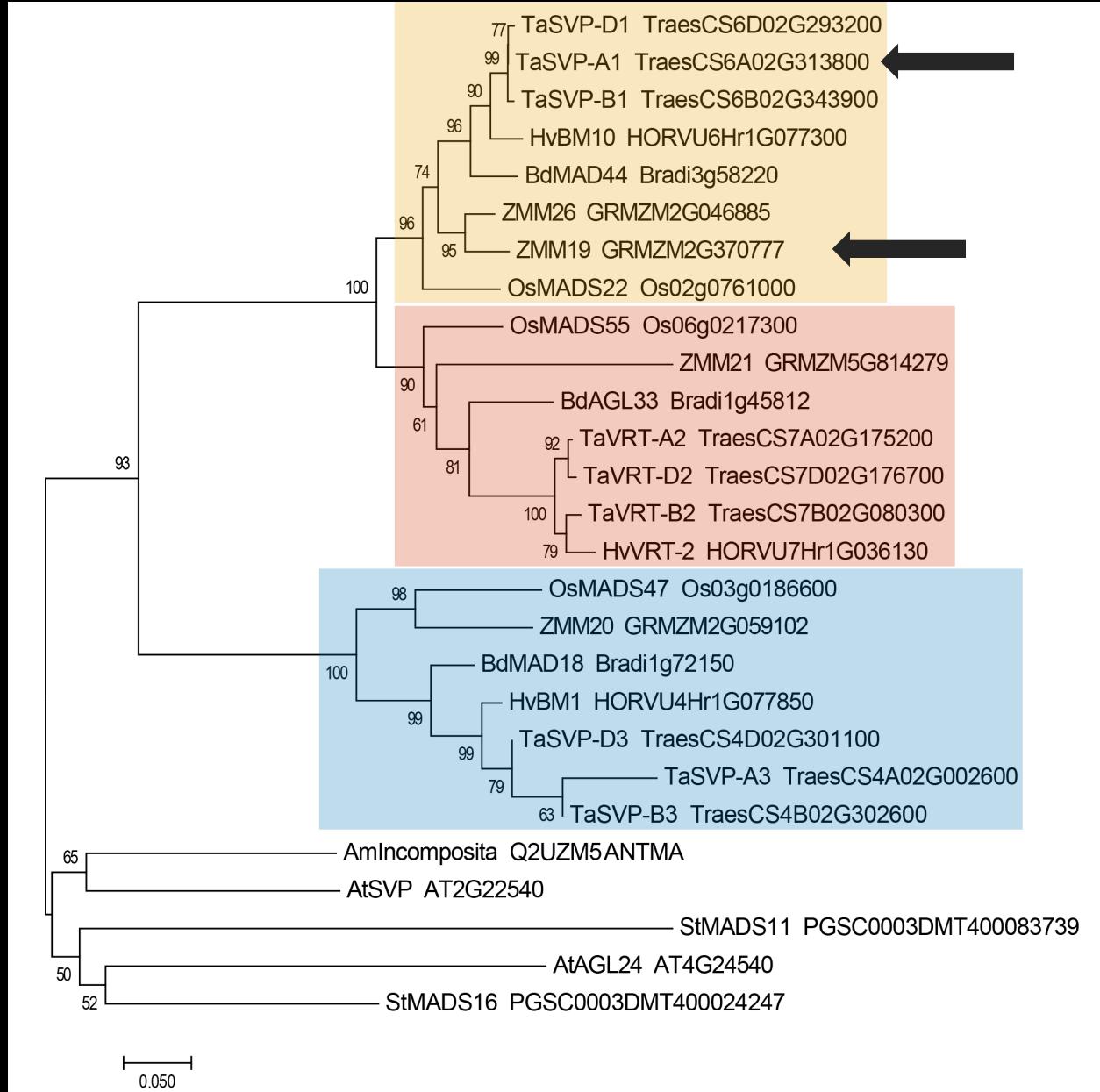
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SVP family in grasses



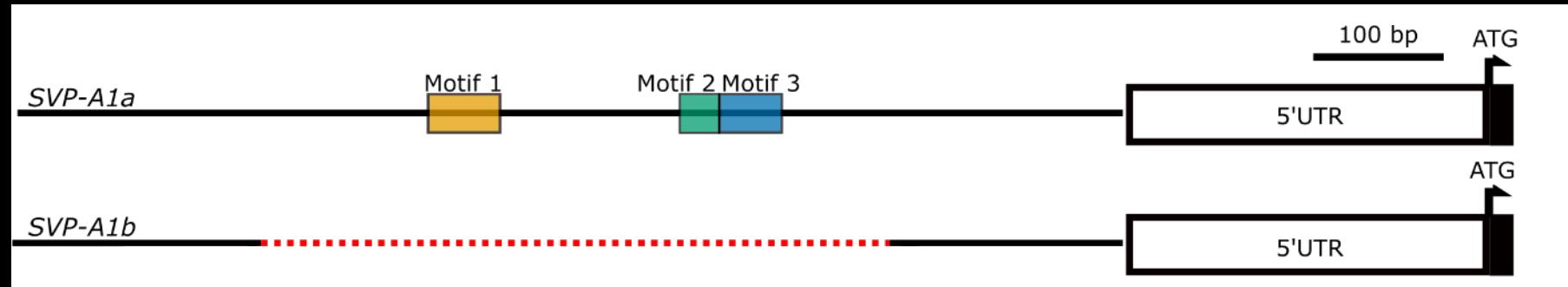
TaSVP-A1 linked to long glumes in *T. ispahanicum*



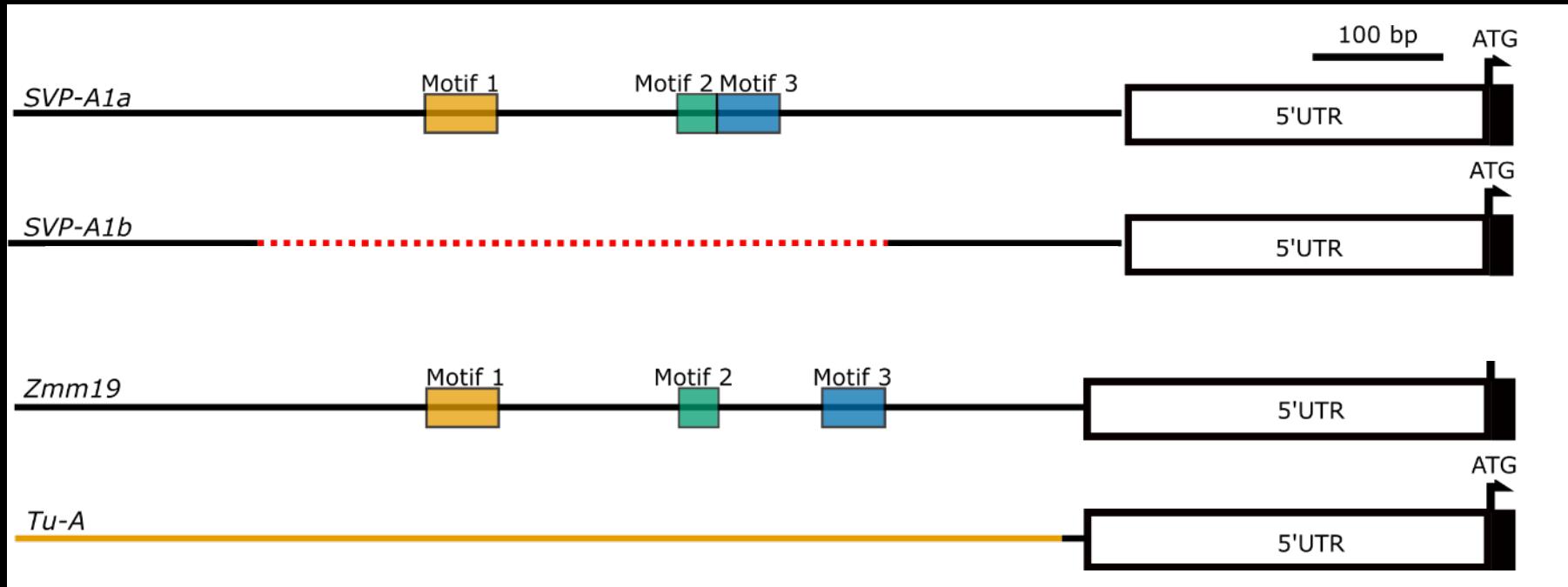
Yi (Andy) Chen



TaSVP-A1 linked to long glumes in *T. ispahanicum*



TaSVP-A1 linked to long glumes in *T. isspahanicum*



Transcription levels of SVPs essential for spike development

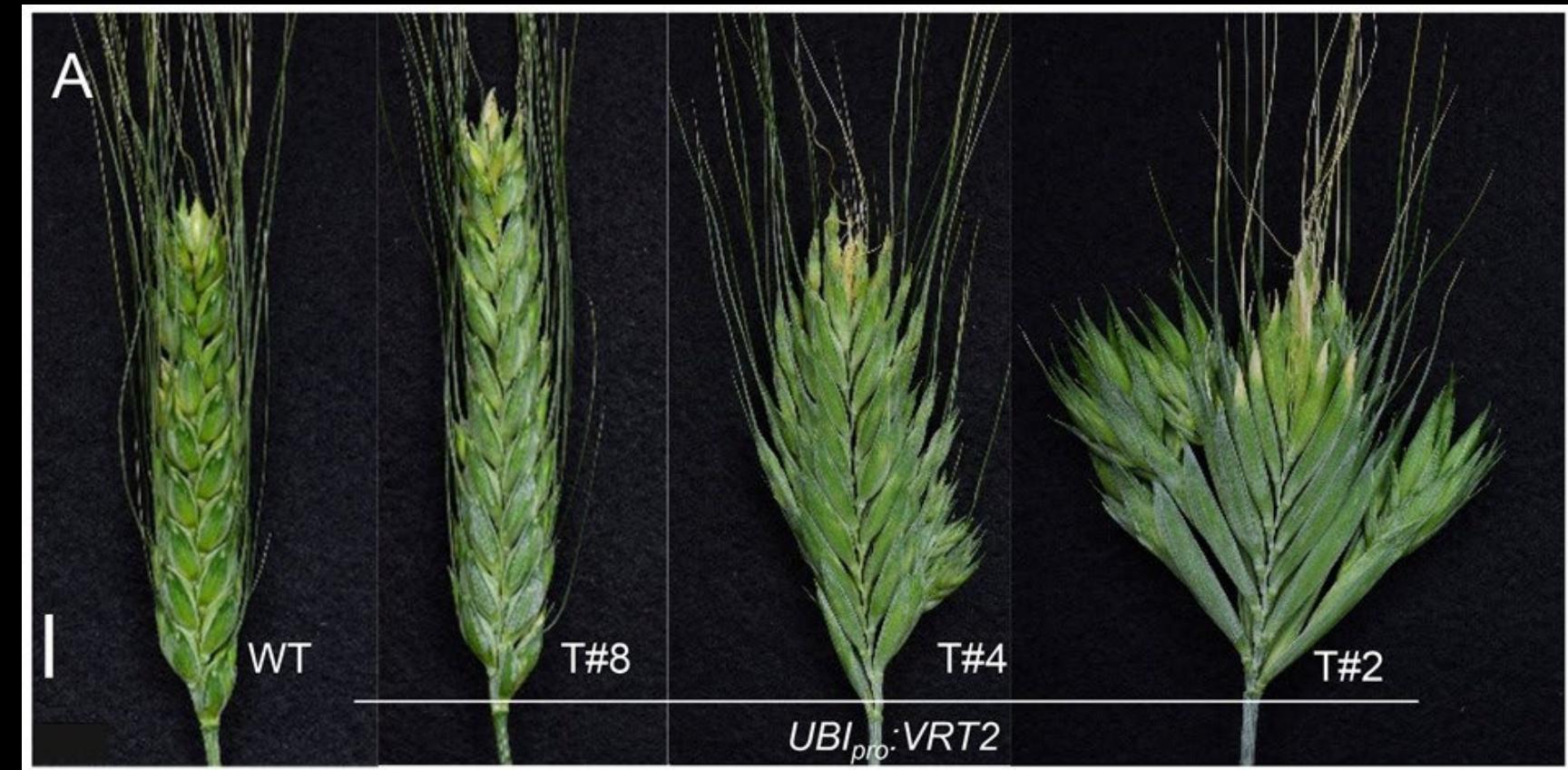


WT

UBI::HvSVP1

Trevaskis *et al.*, 2007

Transcription levels of SVPs essential for spike development



VRT2 transcription levels

WT

UBI::HvSVP1

Trevaskis *et al.*, 2007; Li *et al.*, 2021

Summary

- SVPs play a crucial role in grass spike development
- (Core) Functional regulation (motifs) conserved between distantly related grasses
- SVPs act in a dosage-dependent manner
- Modulating the expression pattern of SVPs holds potential to improve agronomic traits

Next steps

- Investigate motifs in intron 1 of *VRT-A2*
 - Is loss of one or both motifs necessary for “*polonicum*” phenotype?
 - Which proteins bind the intron 1 motifs?
- Elucidate genetic network surrounding *VRT2*
 - Upstream regulators
 - Downstream targets

Thank you for listening



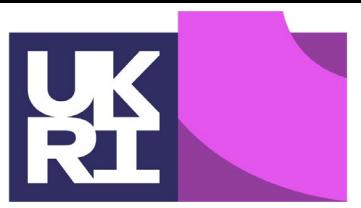
**James Simmonds
Jemima Brinton
Anna Backhaus
Andy Chen
Mark Smedley
Sadiye Hayta
Pamela Crane
Tobin Florio
Elaine Barclay
Cristobal Uauy**

Adamski *et al.*, 2021

<https://doi.org/10.1093/plcell/koab119>



@NikolaiAdamski



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Biological Sciences
Research Council



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IPK Genebank
USDA NSGC
CGN Wageningen**

