

The wheat stem rust resistance gene *Sr43* encodes an unusual protein kinase

Guotai Yu, PhD

 The Wulff Lab

[HOME](#) [PEOPLE](#) [RESEARCH](#) [PUBLICATIONS](#) [MEDIA](#) [TEACHING](#) [GRANTS](#) [CONTACTUS](#)



<https://wulfflab.org>

Stem rust in wheat

One of the major diseases affecting wheat worldwide.

Causal fungal pathogen: *Puccinia graminis* f. sp. *tritici* (Pgt).

In extreme epidemics, can cause severe lodging.



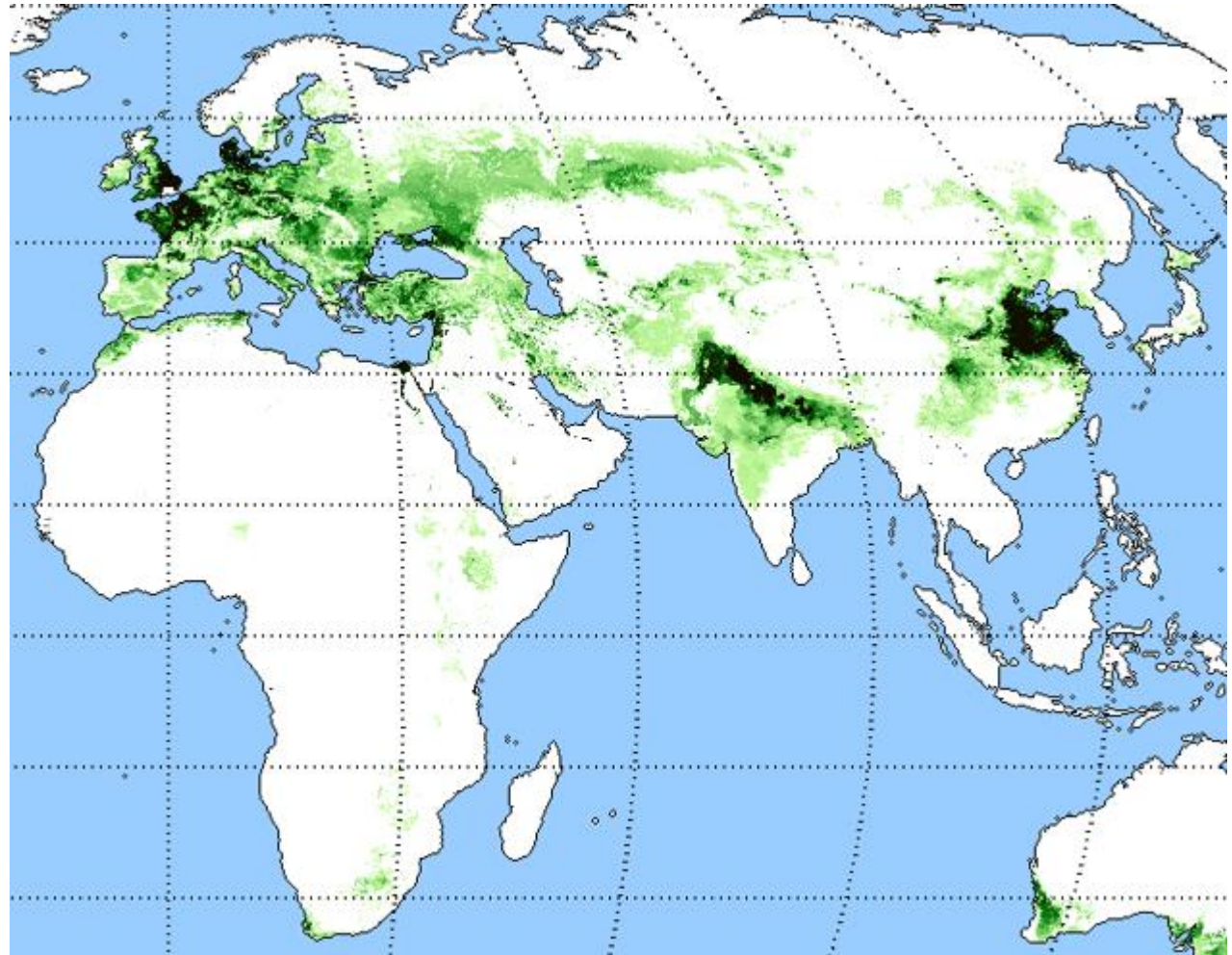
www.sciencedaily.com



Resistant cultivar

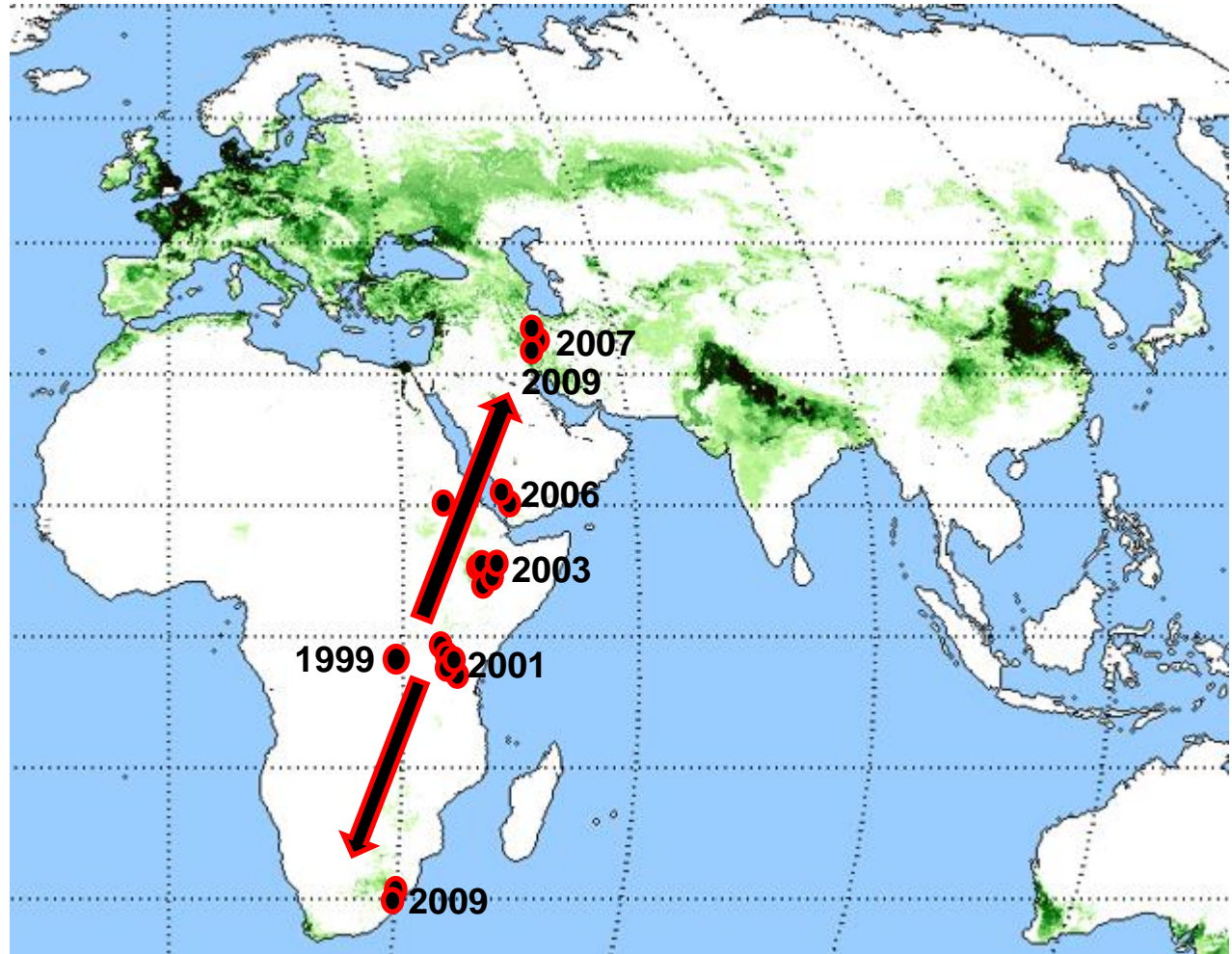
Susceptible cultivar

Wheat production map



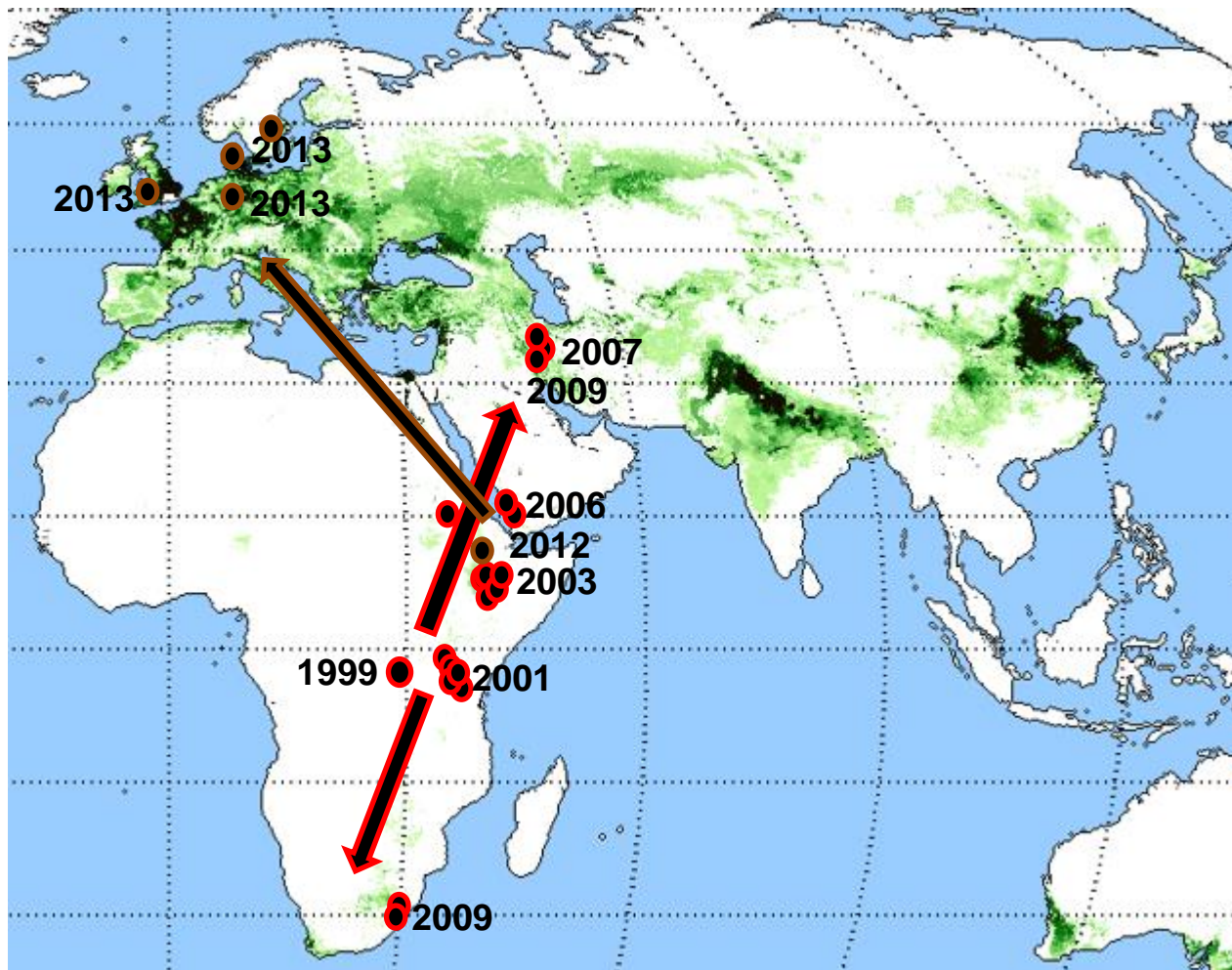
Stem rust in wheat

● Ug99 (TTKSK) sites



Stem rust in wheat

- Ug99 (TTKSK) sites
- TKTTF



Sr genes overcome by new races

Sr31 defeated by Ug99 lineage TTKSK (1999)

Sr24, defeated by Ug99 lineage TTKST (2006)

Sr36 defeated by Ug99 lineage TTTSK (2007)

SrTmp in Digalu defeated by race TKTTF (2013)

General Goal: multi-resistance gene stacking

Wheat



2007 David Monniaus

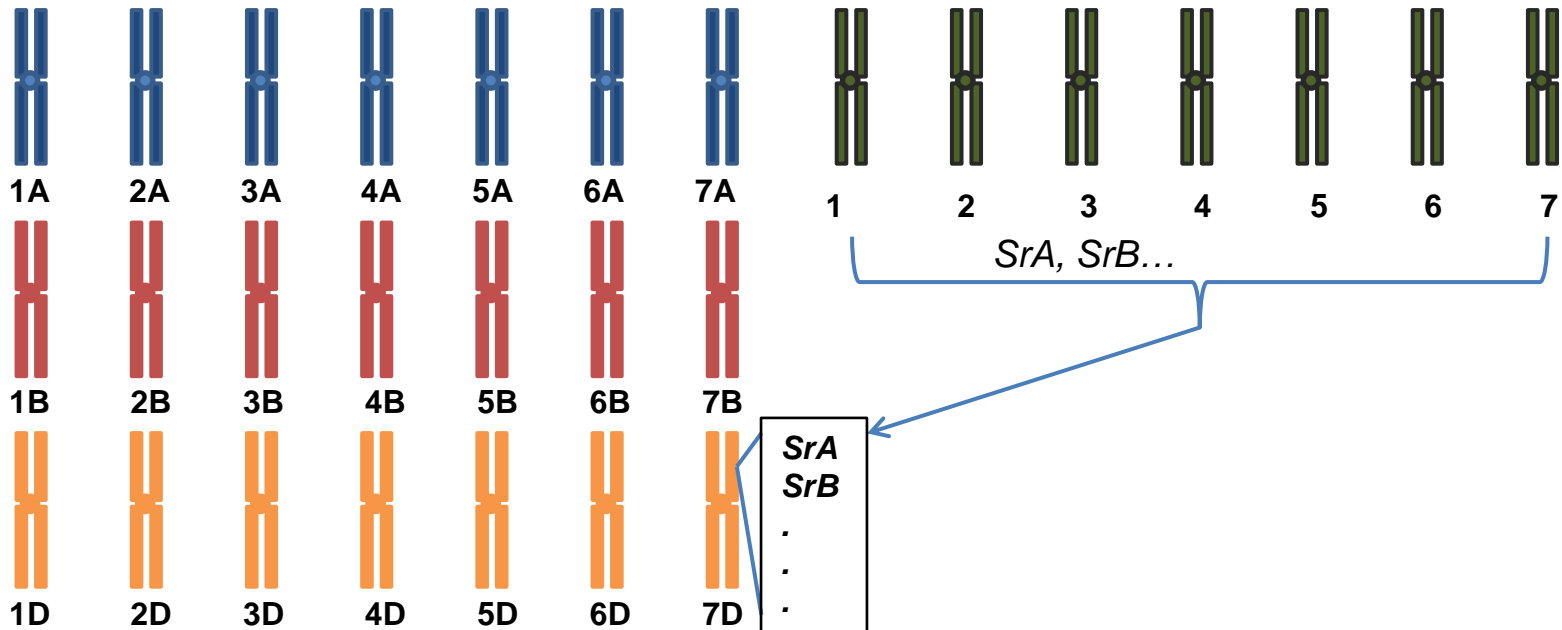
Wild relatives



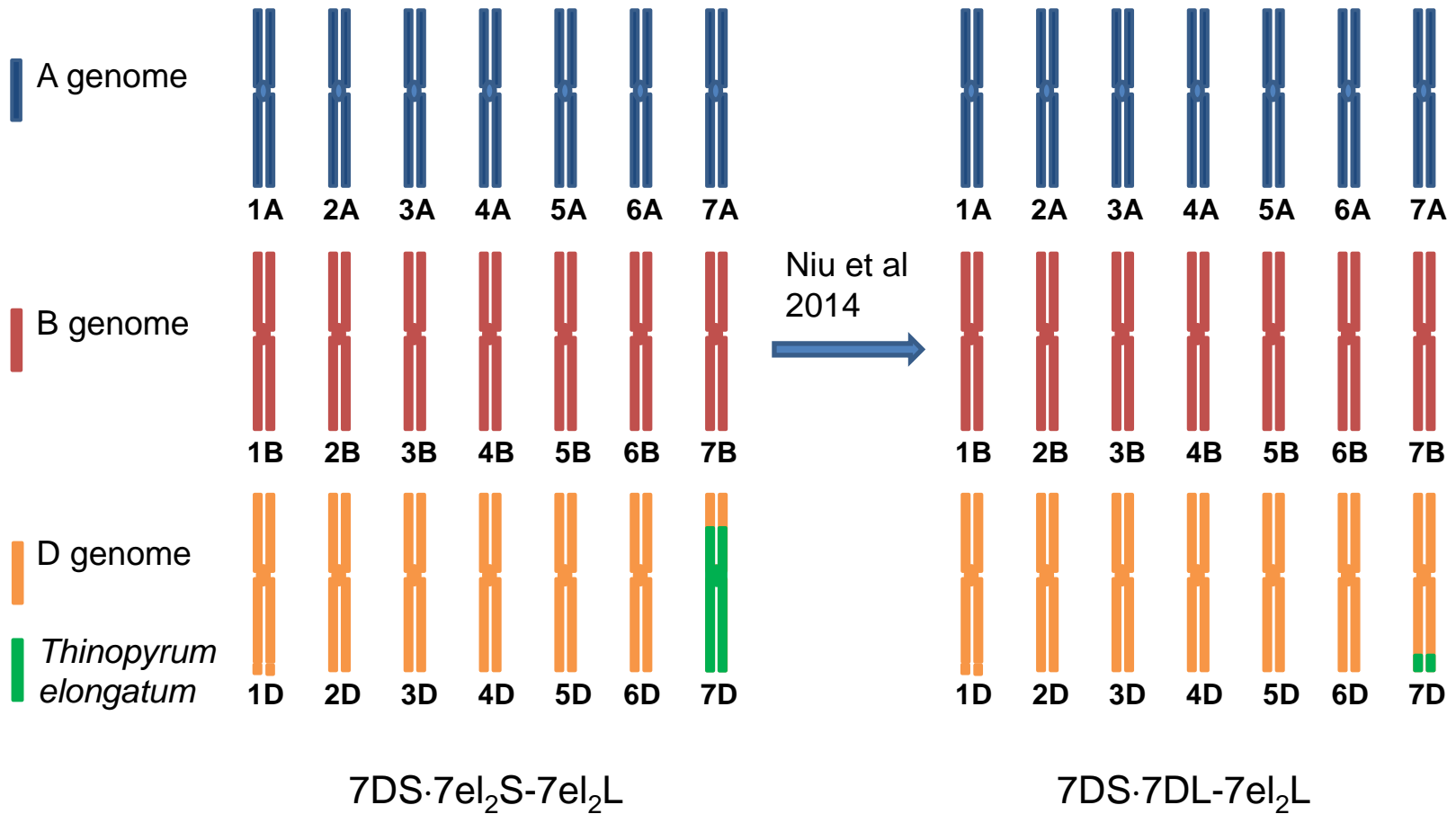
Ae. longissima

Ae. sharonensis

Ae. tauschii

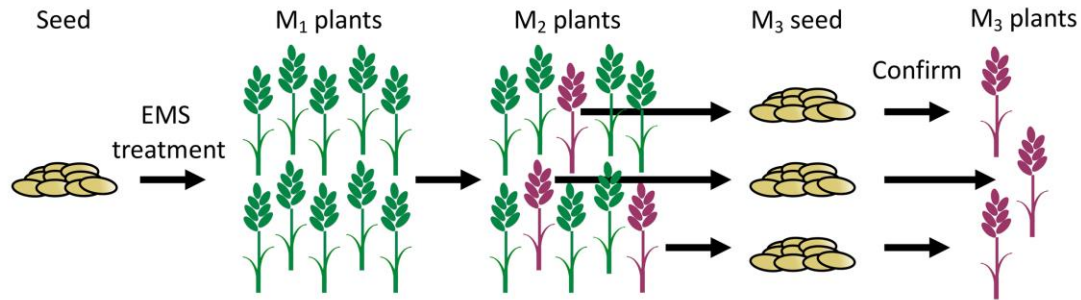


Sr43 introgression line

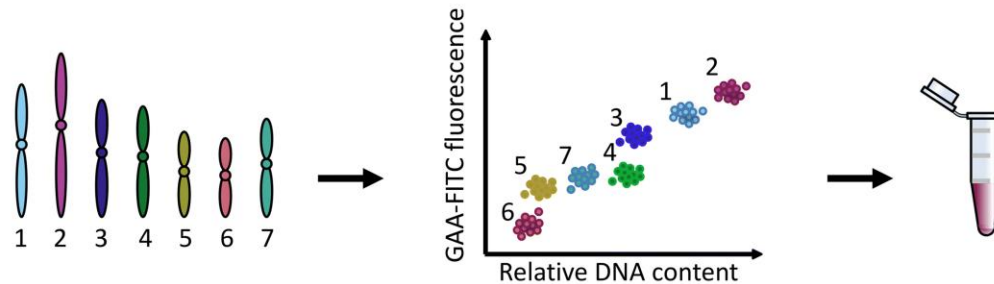


MutChromSeq method

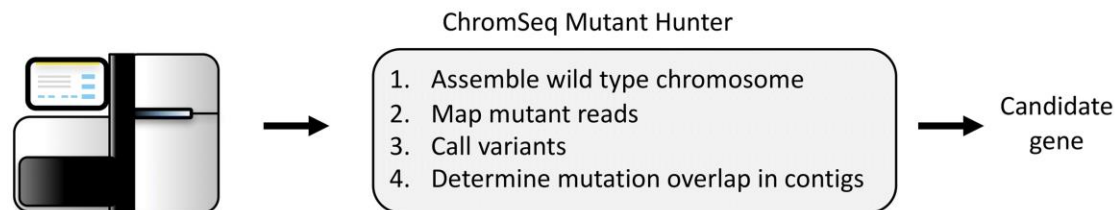
a Mutagenesis and screening: 18 to 24 months



b Chromosome flow sorting: 1 week → DNA amplification: 1 day



c Sequencing: 3 weeks → Bioinformatics: 4 days



Sánchez-Martín et al 2016

Sr43 mutant development

Mutagenized 2,700 *Sr43* introgression line seeds

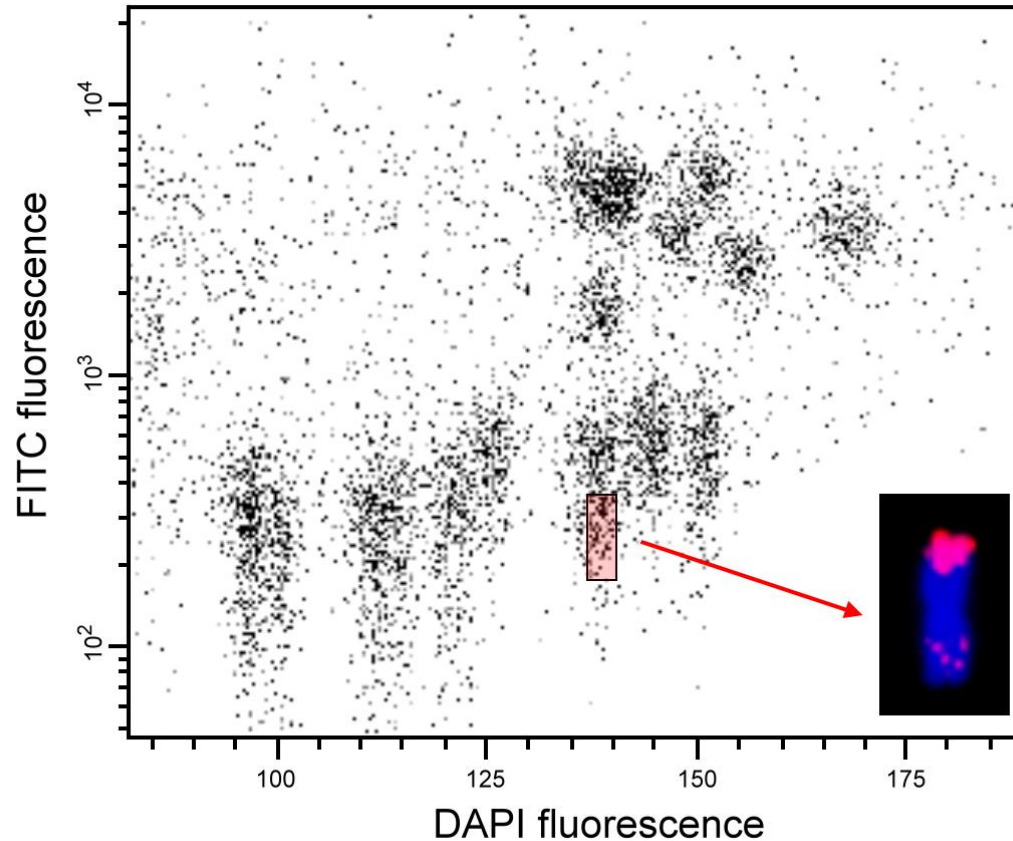
0.80% EMS

1,843 M₂ families with *Pgt* race TPMKC

11 susceptible mutants checked with GBS

8 mutants used for chromosome flow sorting

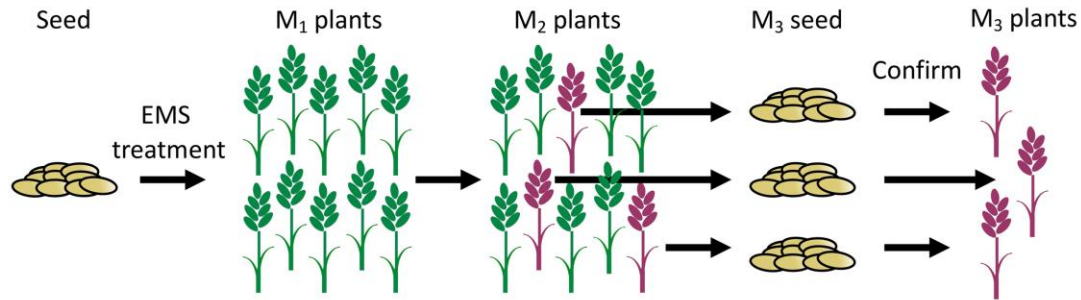
Chromosome flow sorting of *Sr43* mutants



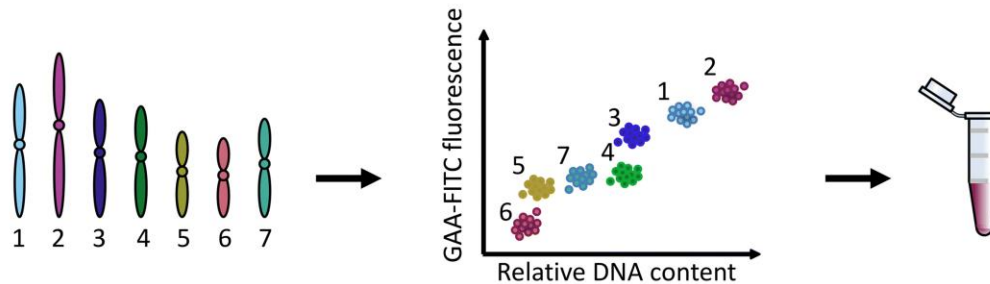
DAPI (x-axis) vs. FITC (y-axis) dot plot was obtained after the analysis of DAPI-stained chromosome suspensions labeled by FISHIS with FITC-conjugated probes for GAA and ACG microsatellites. The 7D/7eI2 translocation chromosomes were sorted from the sorting window shown as red rectangle at purities of 60–65%. Inset: 7D/7eI2 translocation chromosome after FISH with probes for pSc119.2 repeat (green), Afa family repeat (red) and 45S rDNA (yellow) that was used to identify chromosomes in the sorted fraction. Chromosomes were counterstained by DAPI (blue).

MutChromSeq method

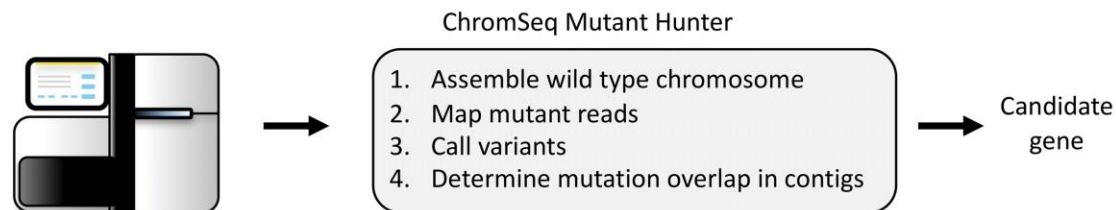
a Mutagenesis and screening: 18 to 24 months



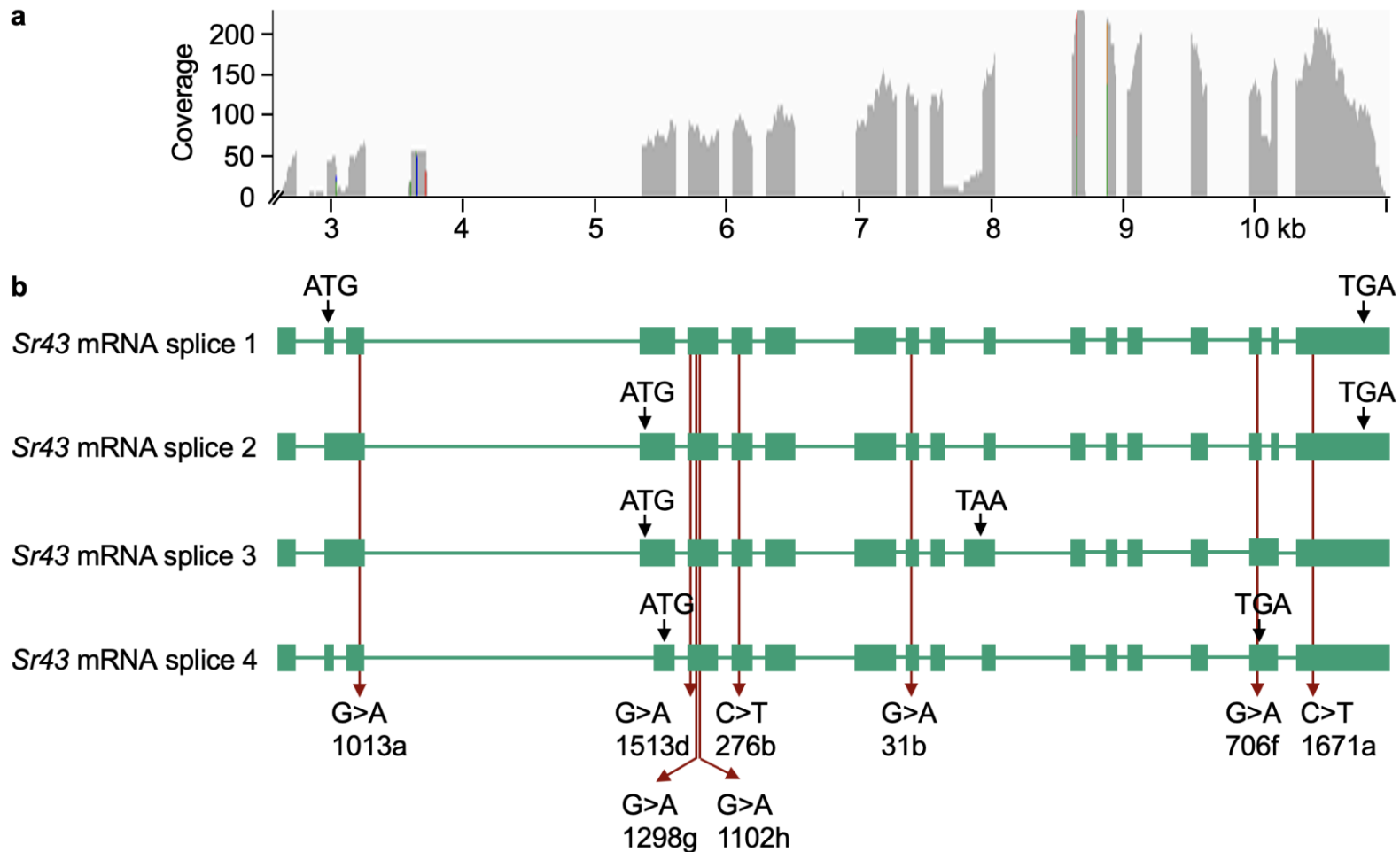
b Chromosome flow sorting: 1 week → DNA amplification: 1 day



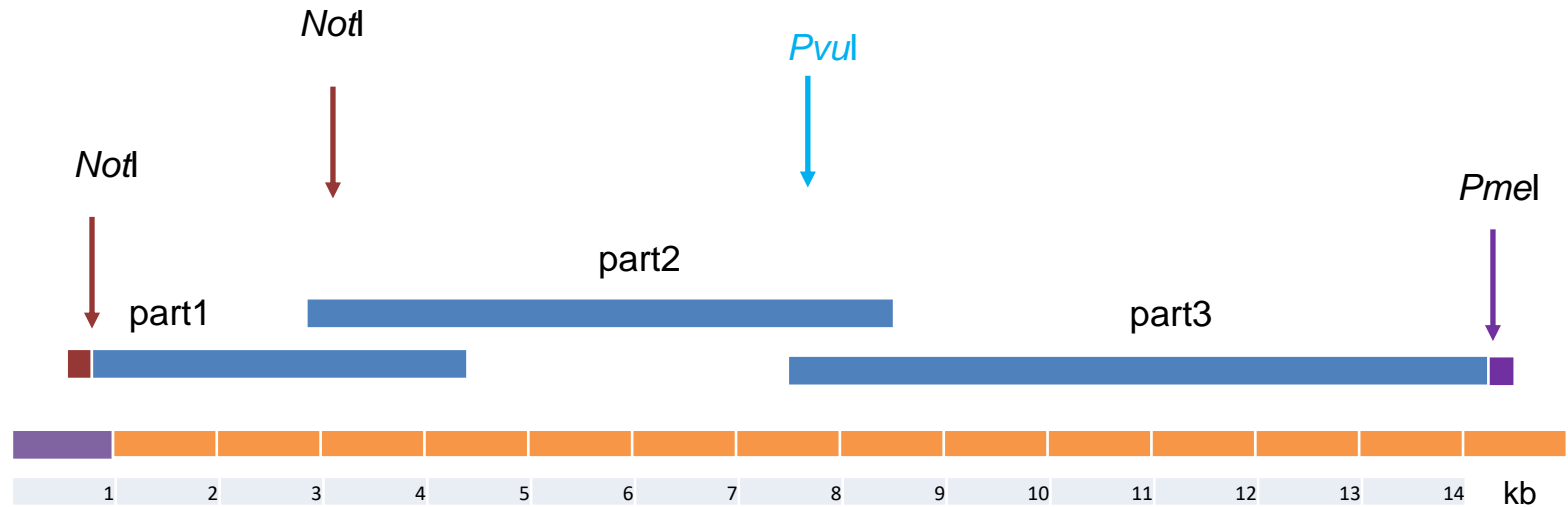
c Sequencing: 3 weeks → Bioinformatics: 4 days



Sr43 transcripts and alternative splicing

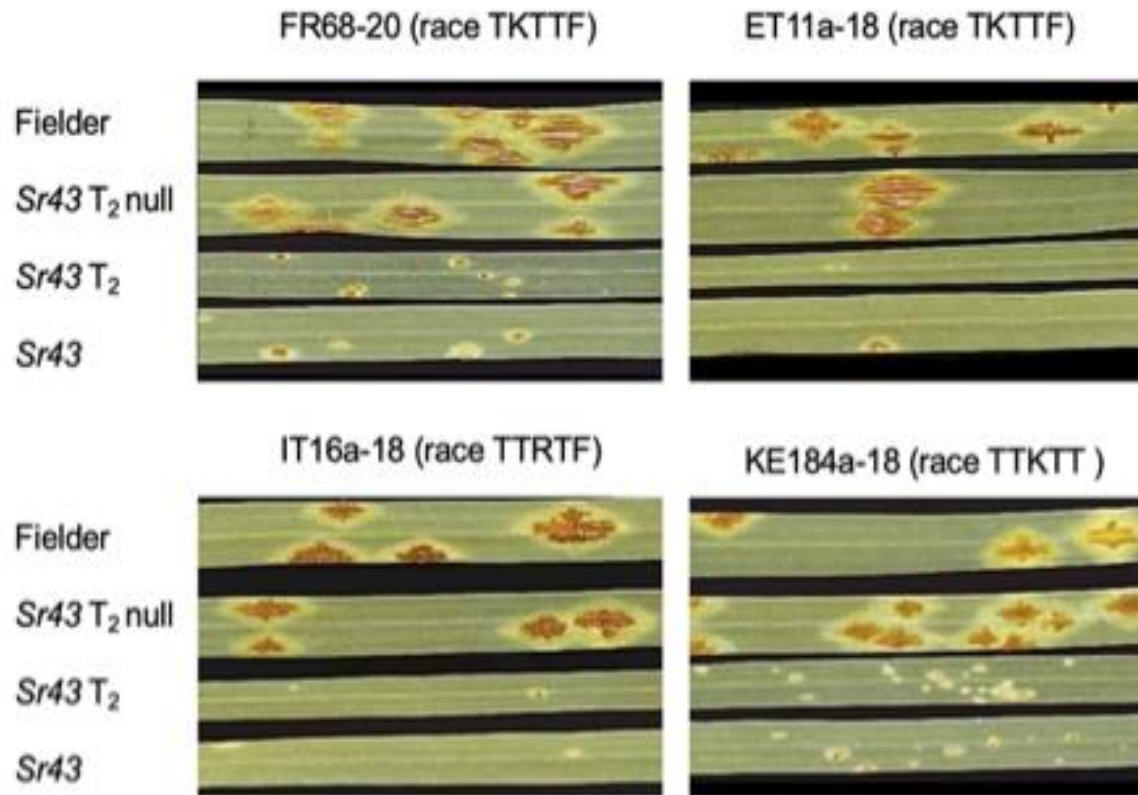


Sr43 construct for transformation



Ligated into vector pGGG and transformed into Fielder

Confirmation of *Sr43* function



Confirmation of *Sr43* race specificity and temperature sensitivity

a

14GEO189-1 21°C

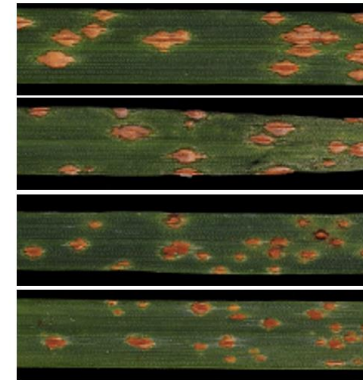
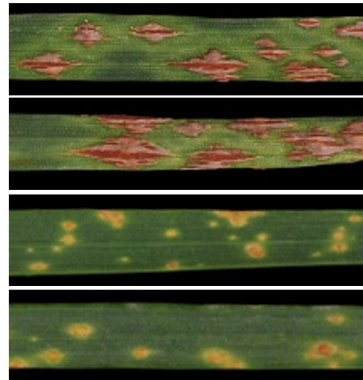
75ND717C 21°C

Fielder

Sr43-T₂-null

Sr43-T₂

Sr43



b

89MN399 21°C

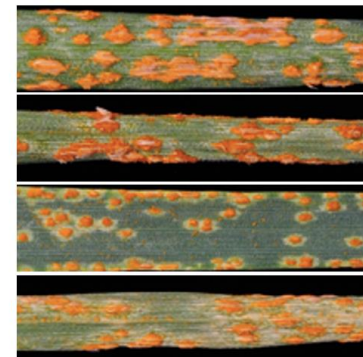
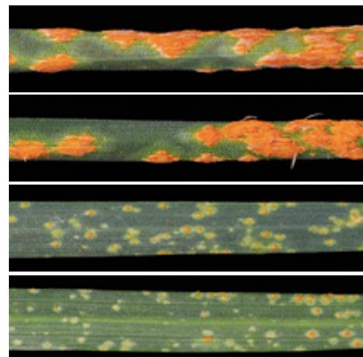
89MN399 26°C

Fielder

Sr43-T₂-null

Sr43-T₂

Sr43

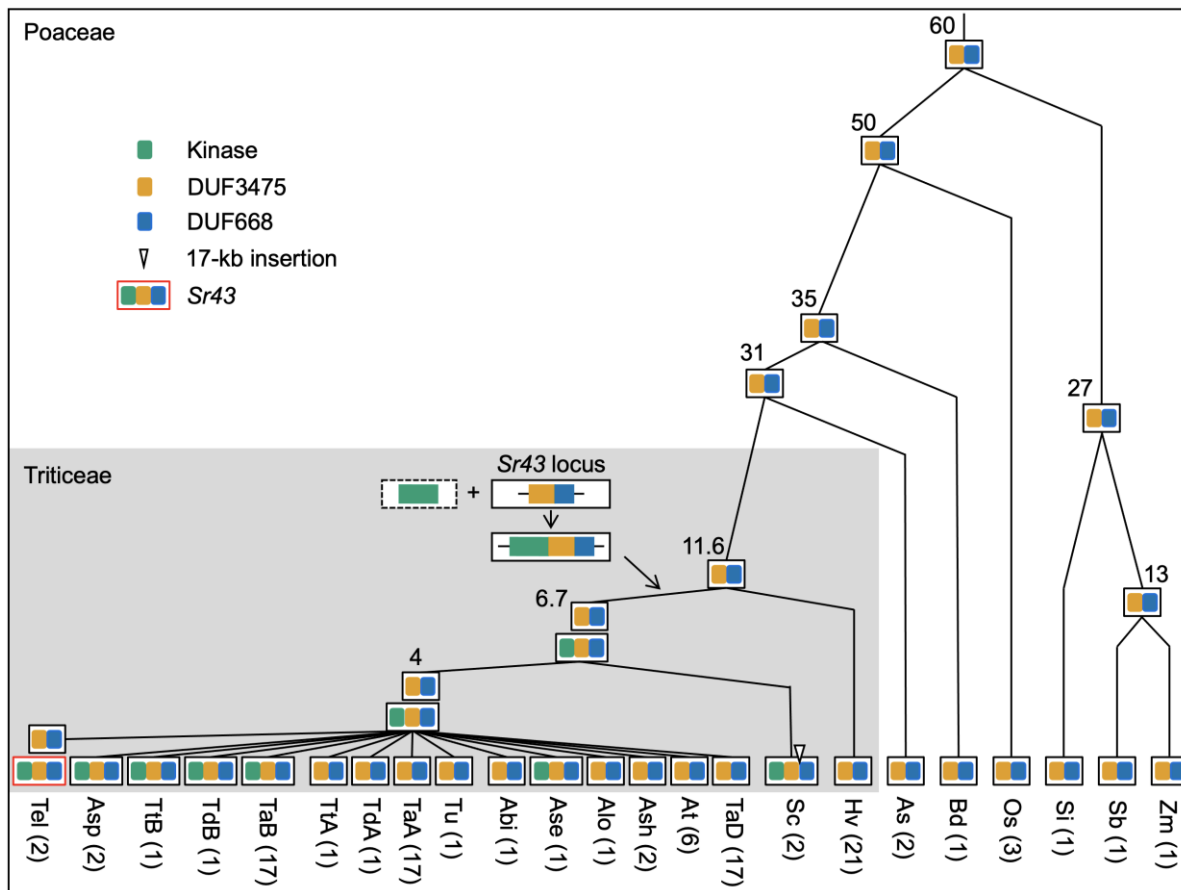


Sr43 has broad spectrum efficacy

Entry	Isolate	Race	Origin	Year	Use in study
1	74MN1409	TPMKC	USA	1974	Screening for mutants*
2	75ND717C	QTHJC	USA	1975	Phenotyping T2
3	69MN399	QTHJC	USA	1969	Phenotyping T2
4	ET11a-18	TKTTF	Ethiopia	2018	Phenotyping T2
5	KE184a/18	TTKTT	Kenya	2018	Phenotyping T2
6	IS#2079	TKTSC	Israel	2022	Phenotyping T1
7	IS#2127	TTTTF	Israel	2022	Phenotyping T1
8	IS#2135	TTTTTC	Israel	2022	Phenotyping T1
9	FR68-20	TKTTF	France	2020	Phenotyping T2
10	IT16a-18	TTRTF	Italy	2018	Phenotyping T2
11	UK-01	TKTTF	United Kingdom	2018	Phenotyping T1
12	14GEO189-1	TTRTF	Georgia	2014	Phenotyping T2

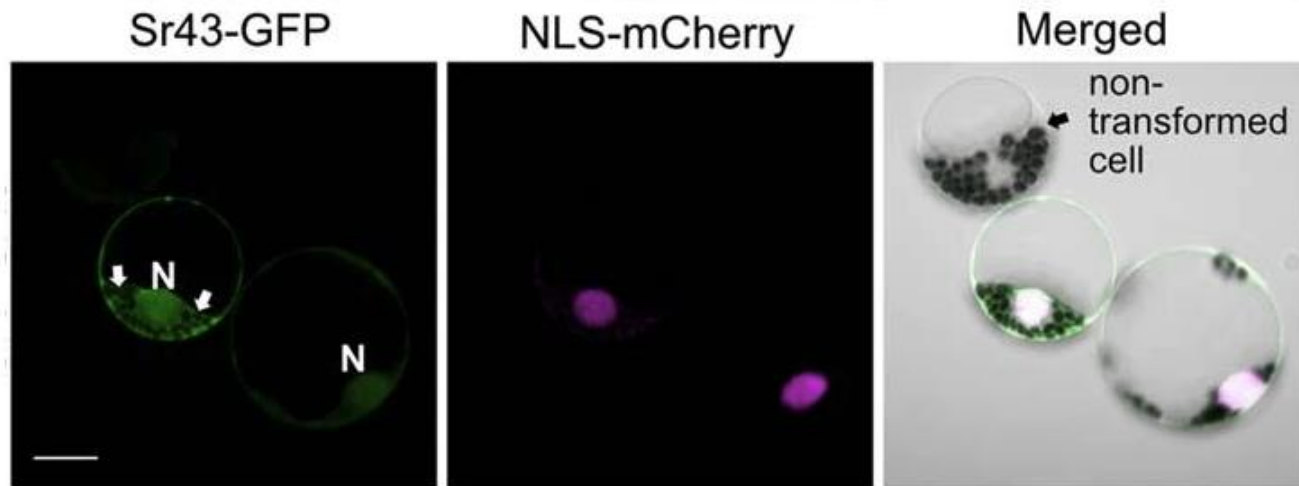
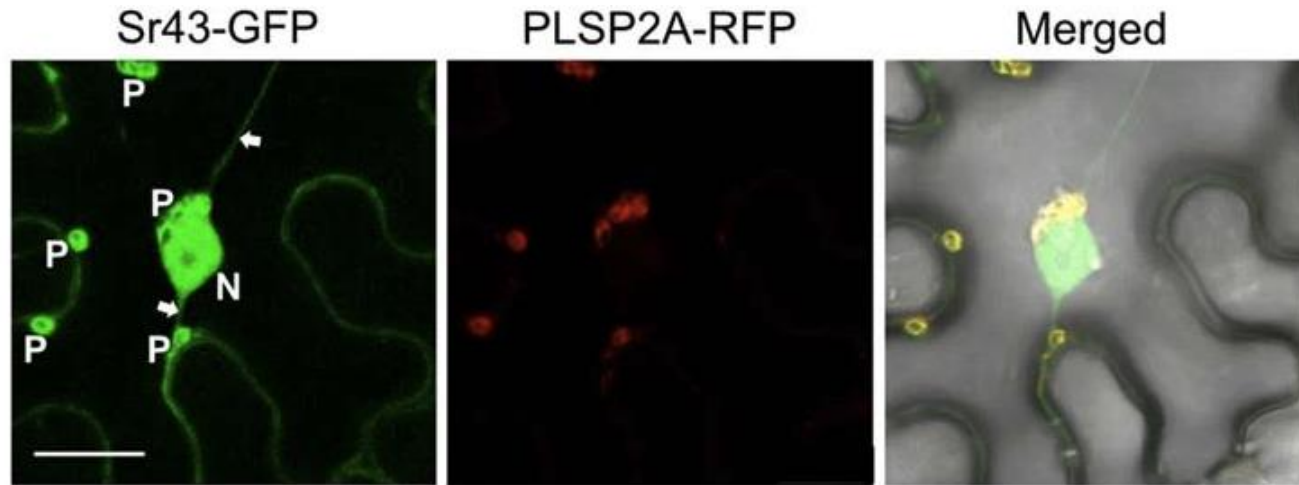
*Isolate 74MN1409 was not used to phenotype the transgenics because it is not virulent on wheat cv. Fielder.

Origin and distribution of *Sr43* orthologs



Species are indicated at the bottom and abbreviated as follows: Tel, *Thinopyrum elongatum*; Asp, *Ae. Speltoides*; TtB, *Triticum turgidum* spp. Durum B genome; TdB, *T. dicoccoides* B genome; Tab, *T. aestivum* B genome; TtA, *T. turgidum* ssp. Durum A genome; TdA, *T. dicoccoides* A genome; TaA, *T. aestivum* A genome; Tu, *T. urartu*; Abi, *Aegilops bicornis*; Ase, *Ae. searsii*; Alo, *Ae. longissima*; Ash, *Aegilops sharonensis*; At, *Ae. tauschii*; TaD, *T. aestivum* D genome; Sc, *Secale cereale* (Rye); Hv, *Hordeum vulgare*; As, *Avena sativa*; Bd, *Brachypodium distachyon*; Os, *Oryza sativa*; Si, *Setaria italica*; Sb, *Sorghum bicolor*; Zm, *Zea mays*. The number of genomes analyzed for each species is indicated in parentheses.

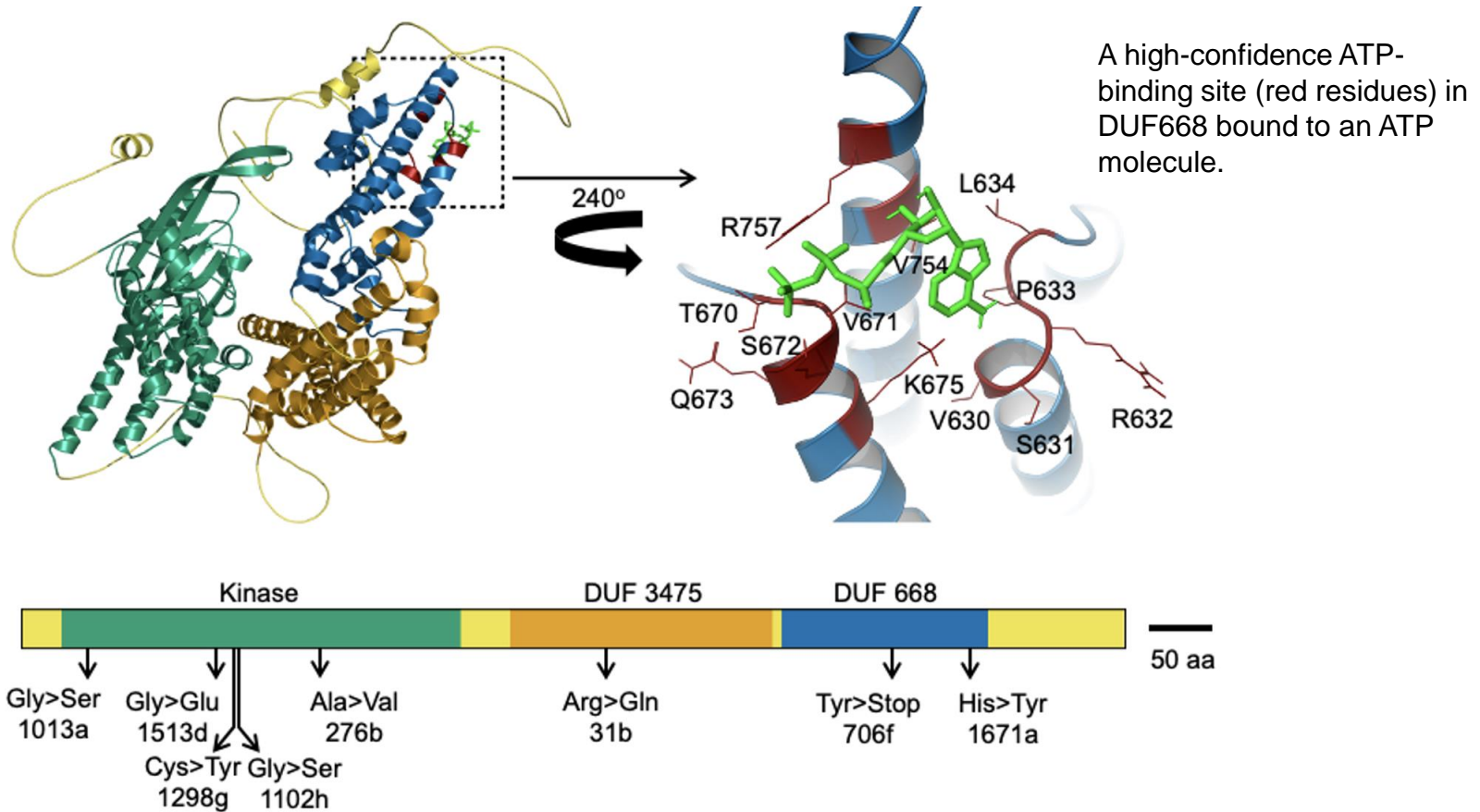
Sr43 localizes to the nucleus and plastids



PLSP2A-RFP: plastid marker

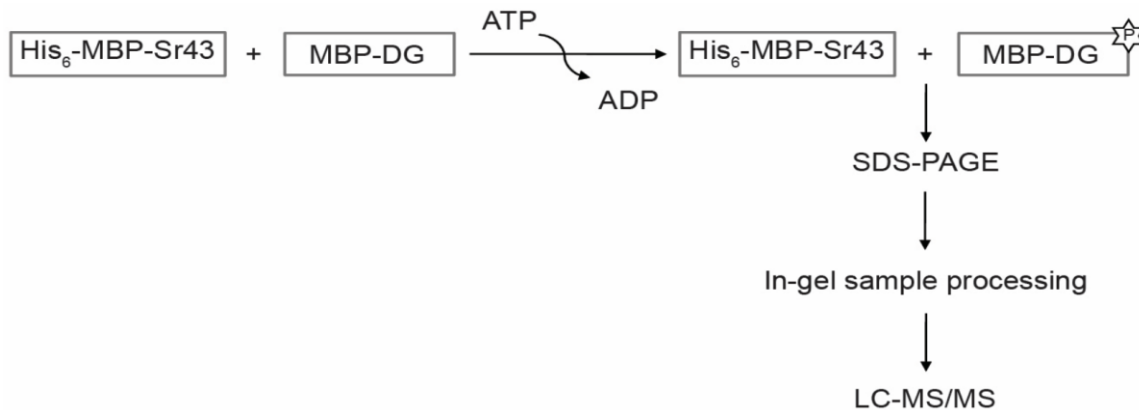
NLS-mCherry: nuclear marker

Sr43 encodes a protein kinase fused to two domains of unknown function



Activity of Sr43 encoded kinase

a



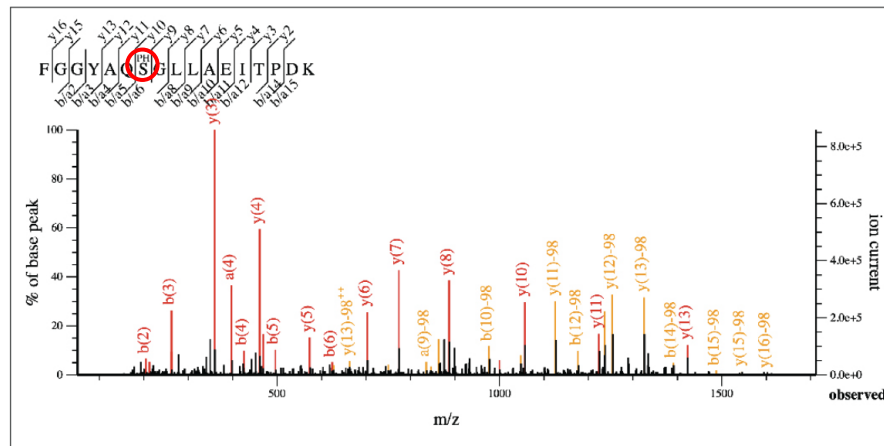
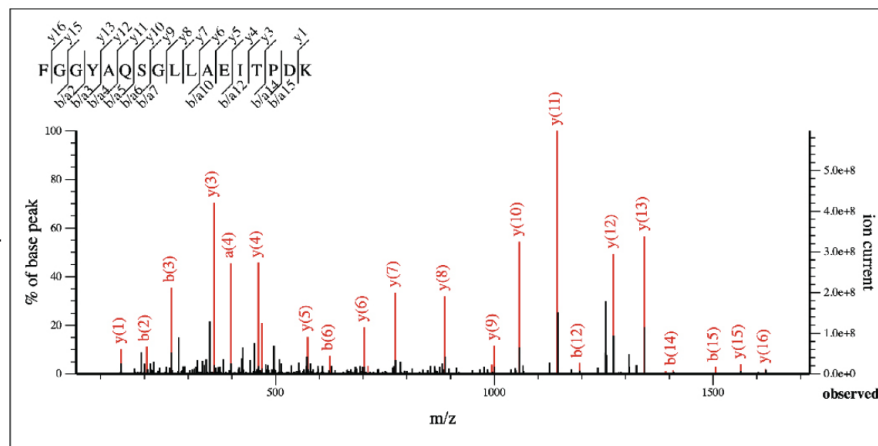
Experimental setups

MBP alone

MBP + His₆-MBP-Sr43

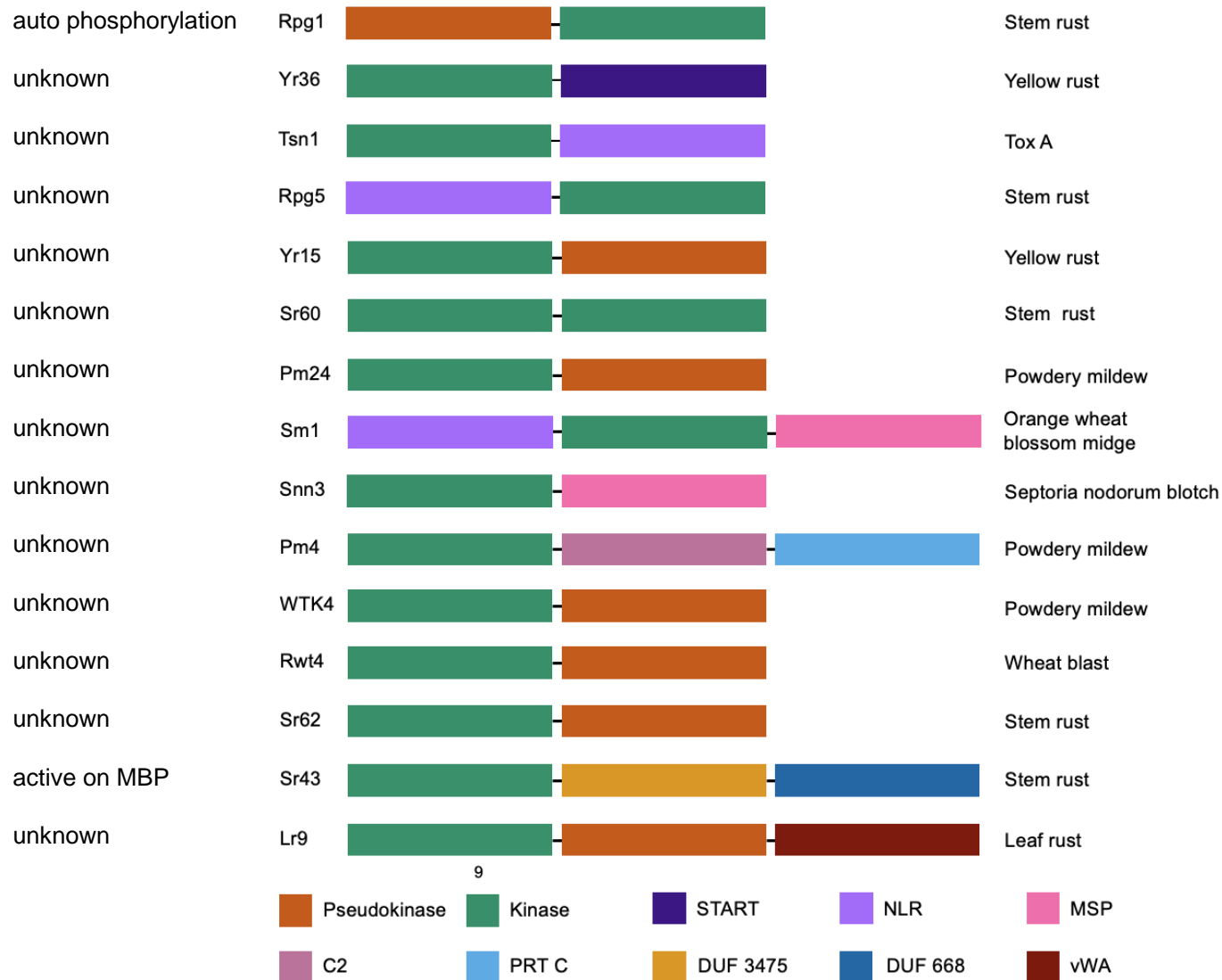
b

S74

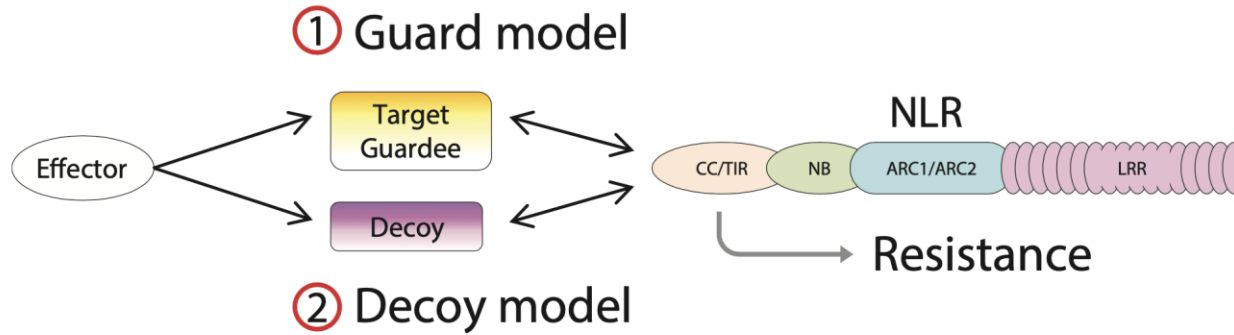


S74, one of the 10 amino-acids phosphorylated by Sr43 kinase.

Triticeae kinase fusion proteins with disease resistance function



NLR models



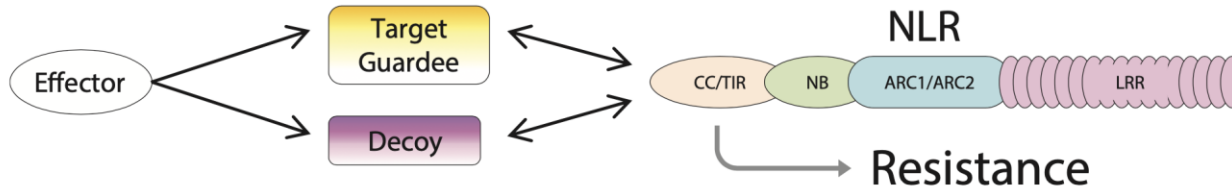
van der Biezen and Jones, 1988

van der Hoorn and Kamoun 2008

NLR models

① Guard model

van der Biezen and Jones, 1988

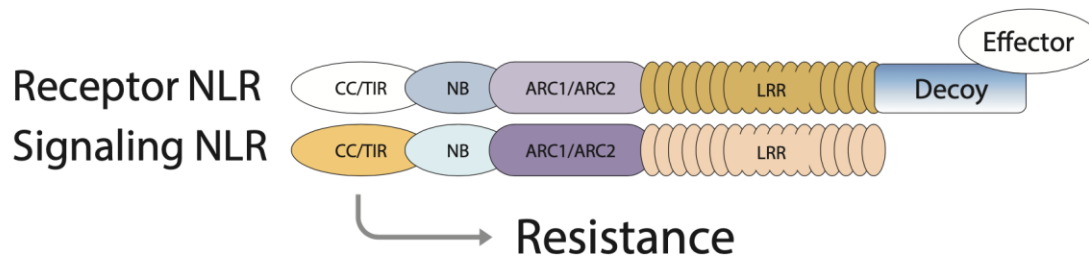


② Decoy model

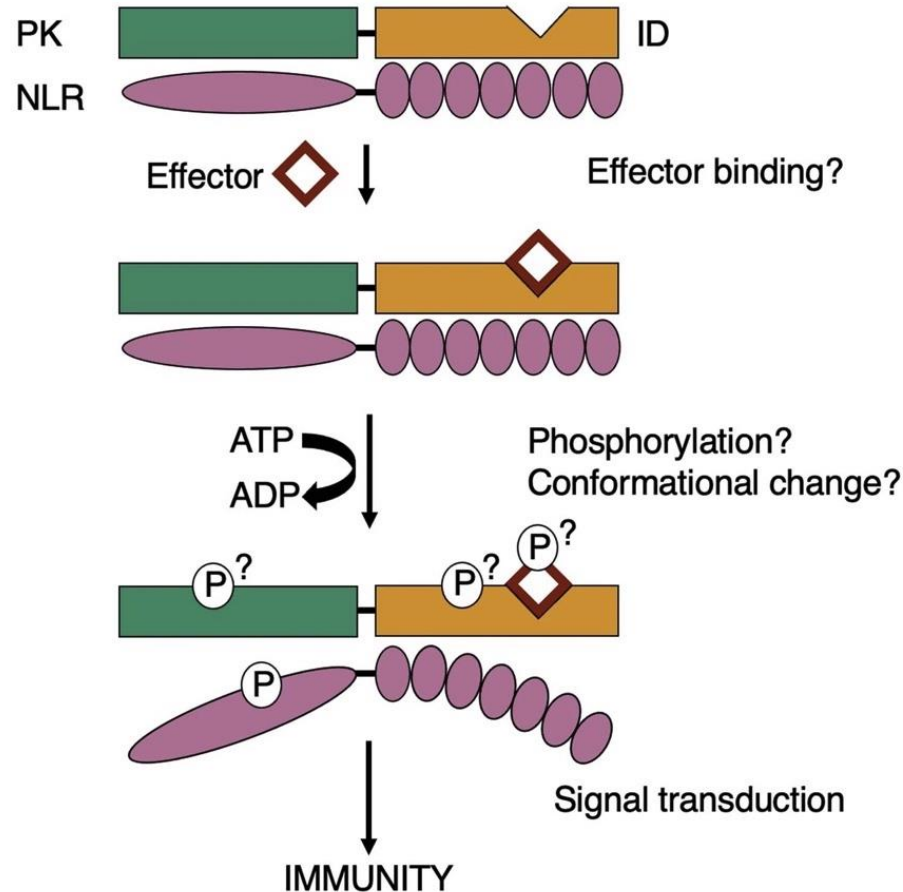
van der Hoorn and Kamoun 2008

③ Integrated decoy model

Cesari et al, 2014



Integrated decoy model for kinase fusion proteins



The protein kinase (PK) fusion protein contains an integrated decoy (ID) that traps the avirulence effector protein (red square). This triggers an autophosphorylation of either the protein kinase or decoy, the effector, or the nucleotide binding leucine-rich repeat (NLR) guard. This results in a conformational change which in turn triggers a signal cascade leading to downstream defense responses and immunity.

Summary

1. *Sr43* cloned
2. *Sr43* confers broad-spectrum stem rust resistance
3. *Sr43* encodes an unusual protein kinase
4. Kinase-fusion protein integrated decoy model

Acknowledgements

Thank you for your attention!

Brande Wulff-JIC, KAUST
Guotai Yu-JIC, KAUST
Catherine Gardener
Yajuan Yue
Ngonidzashe Kangara
Burkhard Steuernagel

Brian Steffenson-UMN
Oadi Matny
Ryan Johnson

Jaroslav Doležel-IEB
Kateřina Holušová
Jan Bartoš
István Molnár

Jesse Poland-KSU
Shuangye Wu

Heribert Hirt-KAUST
Naganand Rayapuram

Łukasz Jaremko-KAUST
Spyridon Gourdoupis

Ikram Blilou-KAUST
Fatimah R. Aljedaani

Wendy Harwood-JIC
Sadiye Hayta
Mark Smedley

Mehran Patpour-
Aarhus University

Matthew N. Rouse-
ARS-USDA

