Leveraging the IWGSC Gene Annotations with Additional Cultivars in Curio

Presented at the 2023 Plant and Animal Genome Conference Author: Shawn Quinn (CTO, Curio Genomics)



Leveraging the IWGSC Gene Annotations with Additional Cultivars in Curio

Presentation Overview

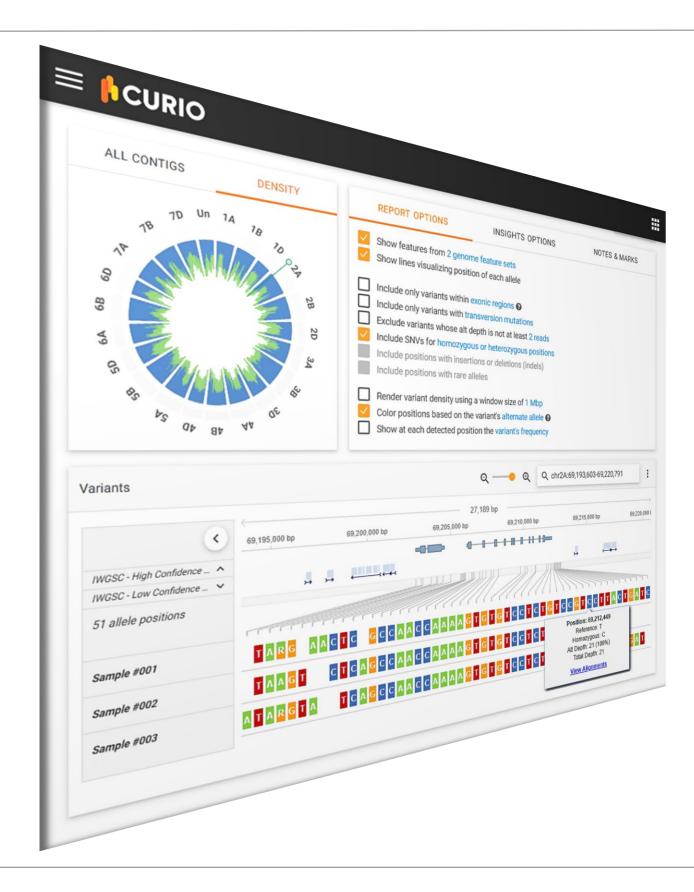
- What is Curio
- Leveraging the 10+ Wheat Genomes Project
 - Read Mapping/Alignment Impact
 - Impact on Variant Calling
 - Leveraging Projected IWGSC Structural and Functional Annotations
 - Sample Analysis Approach / Identifying Genes of Interest
- Looking Ahead
- Acknowledgements

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What is Curio?

- Modern big data management and genomic analysis platform, fully web-based, collaboration ready
- Supports <u>both</u> bioinformatics processing and scientific interpretive analysis
- Massively scalable data processing and interactive data visualizations using real-time databases and clustering technologies
- Designed for extensibility to continuously support new analysis methods, data types, etc.
- Includes robust crop research analysis solutions, including tetraploid and hexaploid wheat DNA-Seq and RNA-Seq analysis



10+ Wheat Genomes Project

- 9 assemblies that include pseudomolecules (including one for the "unknown" chromosome)
- 5 other assemblies that are scaffold-level only
- Gene projections based on the IWGSC annotations
- Gene/Transcript IDs structured like: "Traes<Cultivar><Contig>01<Feature Id>"
- Example: Traes<u>ARI2A01G145700</u>
- Translation table mapping to IWGSC RefSeq 1.1 gene/transcript IDs available at: https://galaxy-web.ipkgatersleben.de/libraries/folders/F1cd8e2f6b131e891





Nine Available Cultivars w/ Pseudo Molecules







	Wheat Cultivar	Region	Pedigree	Version	
	ArinaLrFor	Switzerland	Arina*3/Forno	3.0	
	Jagger	USA	KS-82-W-418/Stephens	1.1	
	Julius	Germany	Asketis/Drifter	1.0	
	Landmark	Canada	Unity/Waskada//Alsen/Superb	1.0	
	LongReach Lancer	Australia	VI184/Chara//Chara/3/Lang	1.0	
	Масе	Australia	Wyalkatchem/Stylet//Wyalkatchem	1.0	
	Norin 61	Japan	Fukuoka Komugi 18/Shinchunaga	1.1	
	Stanley	Canada	CDC Teal//EE8/Kenyon35//AC Barrie	1.2	
/	SY Mattis	France	Apache/Intense	1.0	

Ensembl Plants

CURIO

- Provides curated versions of the same 14 (9 + 5) cultivars from the 10+ Wheat Genomes Project
- Assemblies include individual hundreds of thousands of scaffolds not assigned to pseudomolecules, instead of a single "unknown" pseudomolecule
- Updated de-novo gene annotations, which were processed by the PGSB (Plant Genome and Systems Biology)
- Gene/Transcript IDs structured like: "Traes<Cultivar><Contig>03<Feature Id>"
- Example: Traes<u>ARI2A</u>03<u>G00626940</u>
- Translation table to IWGSC gene/transcript ids is not yet available



https://plants.ensembl.org/Triti cum_aestivum/Info/Strains

Projected and De-Novo Genes Annotations

Wheat Cultivar	Original Version ^(#1)	Updated Version (#2)
ArinaLrFor	1,231,314 features	2,213,22 features
Jagger	1,221,309 features	2,110,846 features
Julius	1,217,757 features	2,289,641 features
Landmark	1,216,606 features	2,166,879 features
LongReach Lancer	1,225,483 features	2,079,609 features
Масе	1,223,663 features	2,085,425 features
Norin 61	1,219,068 features	2,188,731 features
Stanley	1,220,222 features	2,128,177 features
SY Mattis	1,230,268 features	2,138,635 features
#1. Original version based on projection	s of the IWGSC High Confidence structural apportation	

#1: Original version based on projections of the IWGSC High Confidence structural annotations #2: Updated de-novo gene annotations, as of the "release 54" version of the plants.ensembl.org build



Wheat DNA-Seq: **Read Mapping and Navigation**

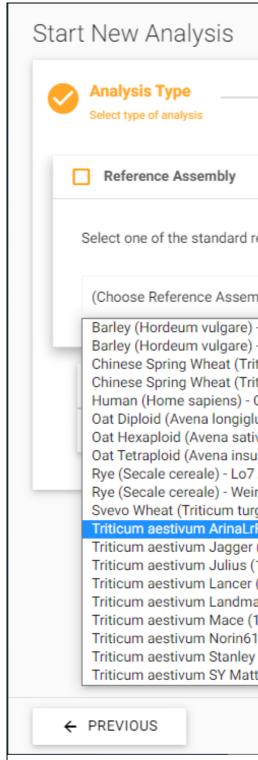
©2023, Curio Genomics

Multiple read mapping algorithms • with pre-built indexes that are deployed and ready on computational cluster

	nalysis Type 2 Select Inputs 3 Configuration Tune analysis options						
nich	type of data would you like to analyze?						
DNA-Seq							
) F	NA-Seq						
Q	Type here						
C	Bowtie2 Alignment Map the reads of one or more single or paired-end samples to a reference genome using the Bowtie2 algorithm BWA-MEM Alignment Map the reads of one or more single or paired-end samples to a reference genome using the BWA-MEM algorithm						
0	BWA-ALN Alignment Map the reads of one or more single or paired-end samples to a reference genome using the BWA-ALN algorithm						
O General Variant Detection Detect and call variants in one or more samples by comparing their aligned sequences to the reference							
0							

Wheat Read Mapping

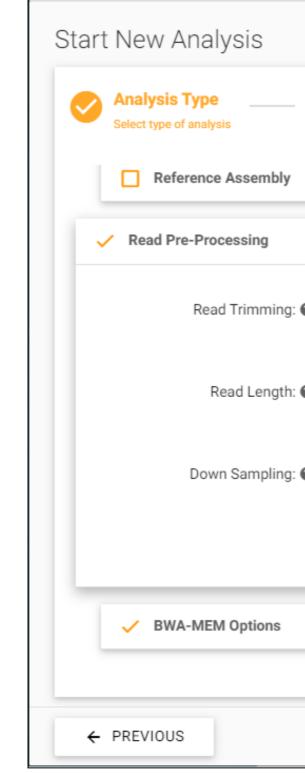
- Multiple read mapping algorithms with pre-built indexes that are deployed and ready on computational cluster
- Conveniently curated and instantly available assemblies from the "10+ Wheat Genomes" project and others
- Automatic support for custom reference assemblies



Wheat Read Mapping

	×
Choose files to analyze	
reference assemblies to align to, or use a custom reference. ?	
nbly) 🗸	
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START ANALYSIS	

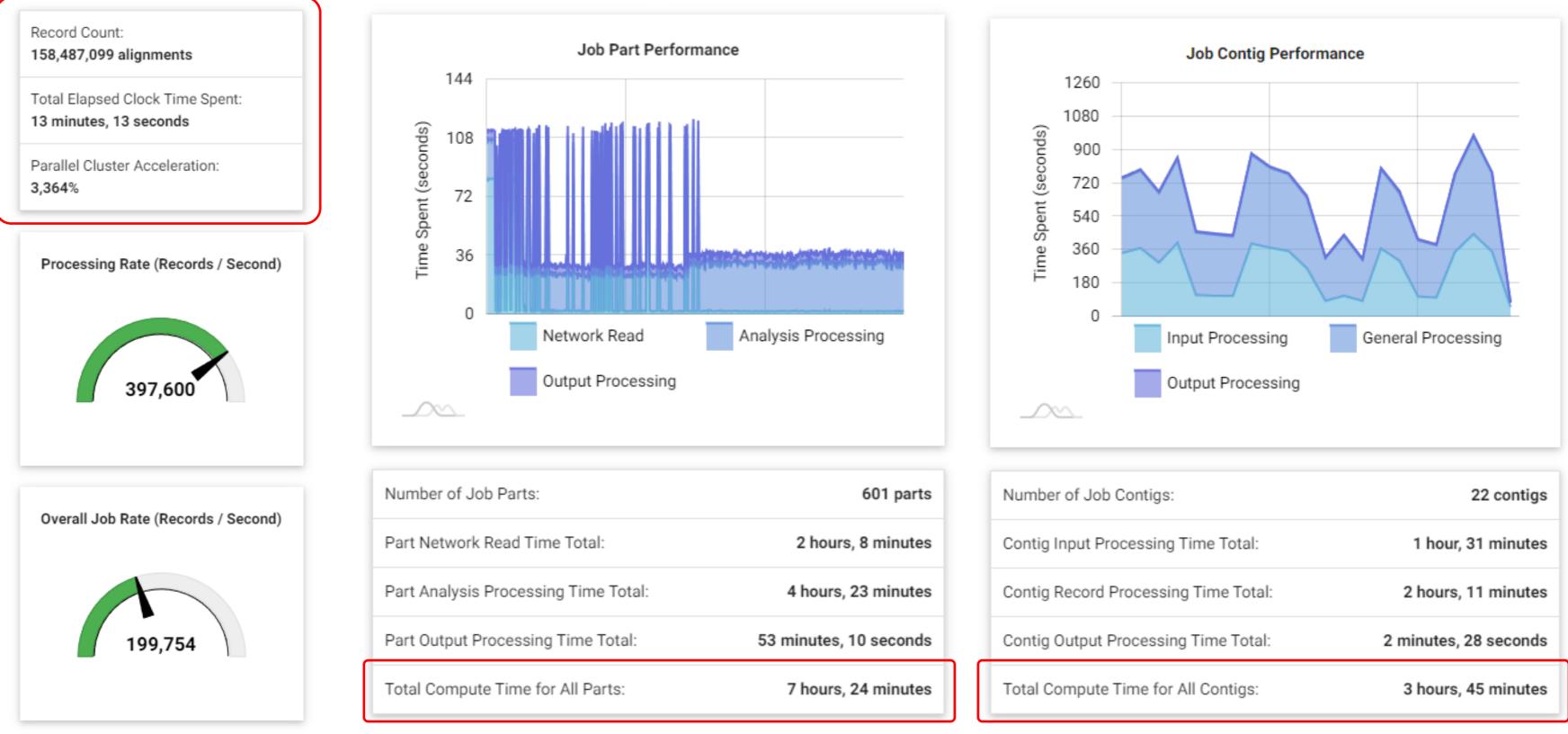
- Multiple read mapping algorithms with pre-built indexes that are deployed and ready on computational cluster
- Conveniently curated and instantly available assemblies from the "10+ Wheat Genomes" project and others
- Automatic support for custom reference assemblies
- Experiment with various alignment and read processing options without requiring any pipeline configuration



Wheat Read Mapping

			×
	~	Select Inputs 3 Configuration Choose files to analyze Tune analysis options	
0	•	Do not perform any read trimming	
	0	Trim the reads before aligning	
Ø	• 0	Process reads of any length Discard short reads	
Ø	0 0	Process all reads Process a sampling (%) of the reads Process a fixed number of reads	
	_		
		START ANALYSIS	•

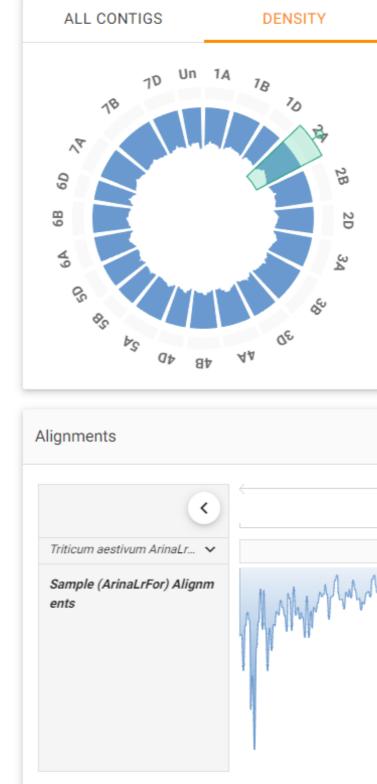
Parallel Read Mapping to ArinaLrFor Genome



er of Job Contigs:	22 contigs
Input Processing Time Total:	1 hour, 31 minutes
Record Processing Time Total:	2 hours, 11 minutes
Output Processing Time Total:	2 minutes, 28 seconds
Compute Time for All Contigs:	3 hours, 45 minutes

10+ Wheat Genomes: Alignment Visualization

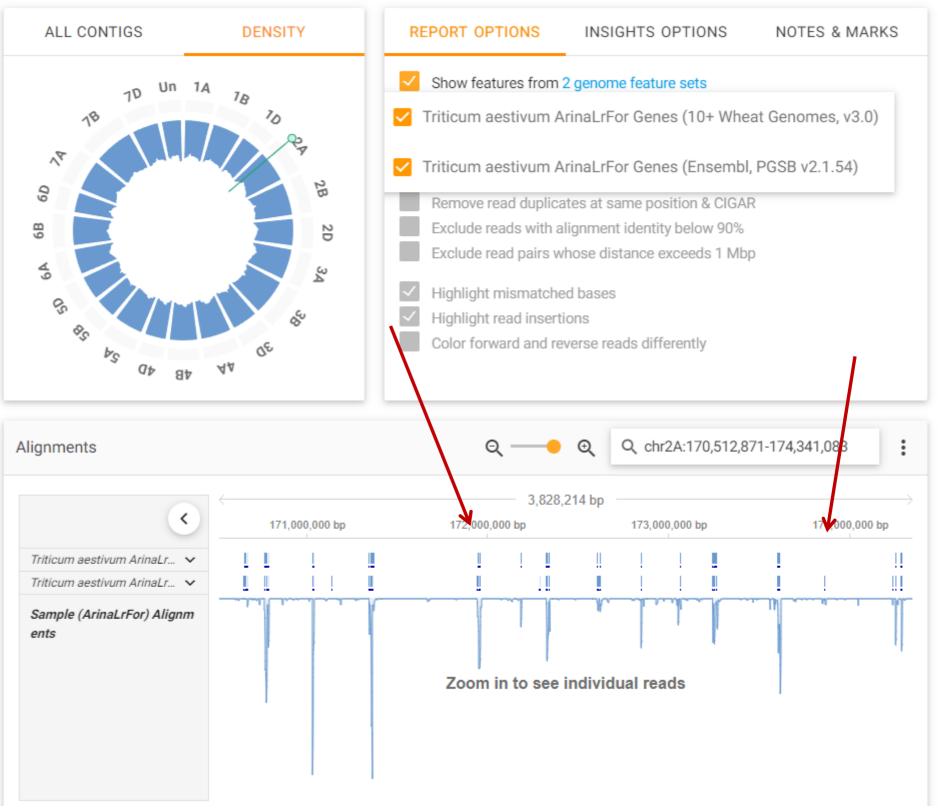
 Quickly browse and visualize aligned reads and depths from samples of any size, anywhere in the genome

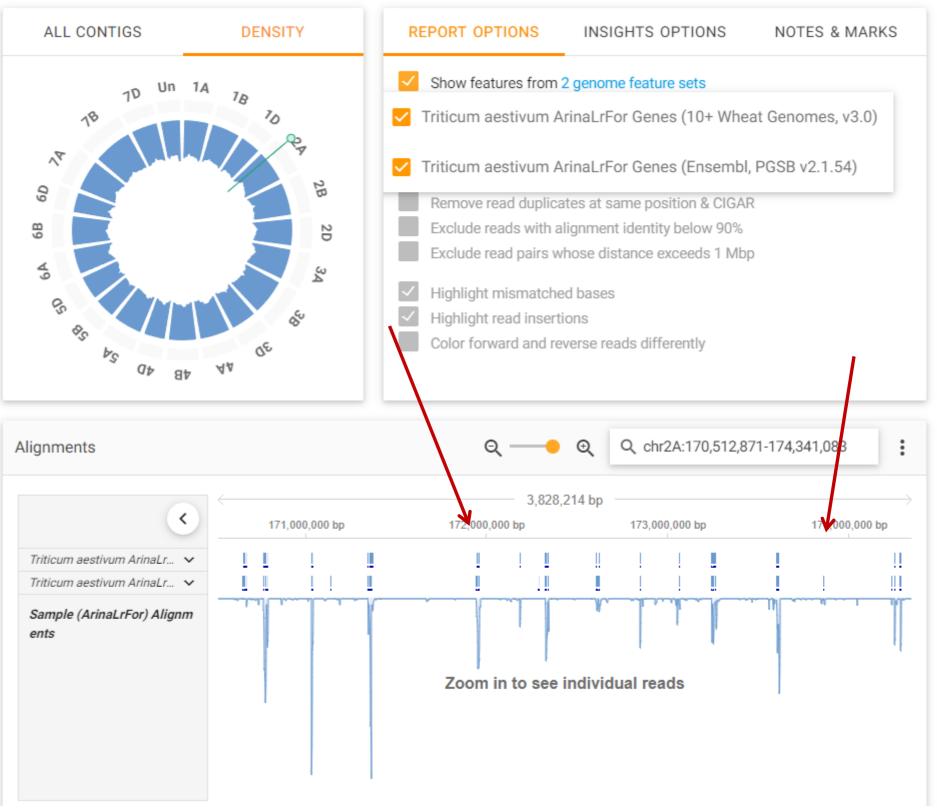


✓ Shov	 Show features from 1 genome feature set Show reference assembly bases 							
 Show coverage and highlight when differences exceed 20% Exclude reads and bases with quality under 99% 								
						Rem	ove read duplica	ates at s
_		-	nt identity below 90%					
Excl	ude read pairs w	hose di	stance exceeds 1 Mi	qq				
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	784,661	1,008 bp						
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10+ Wheat Genomes: Alignment Visualization

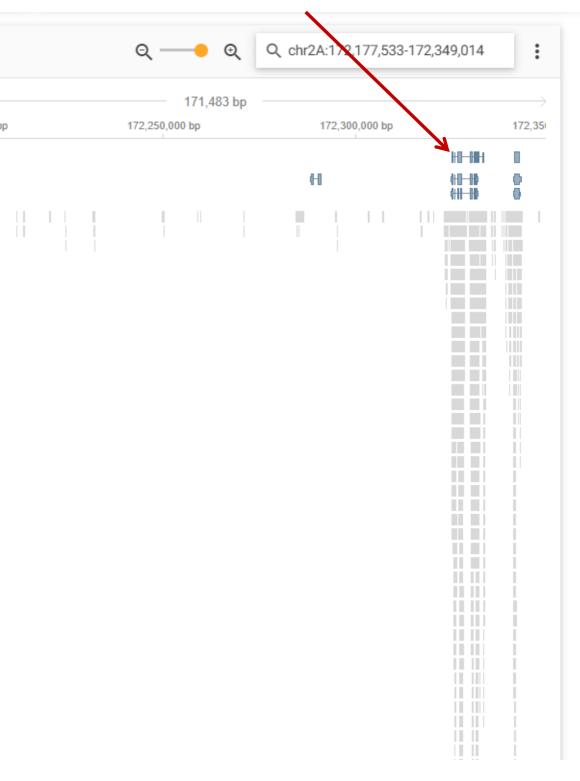
- Quickly browse and visualize aligned • reads and depths from samples of any size, anywhere in the genome
- Reference projected & de-novo gene • annotations from 10+ Wheat Genomes





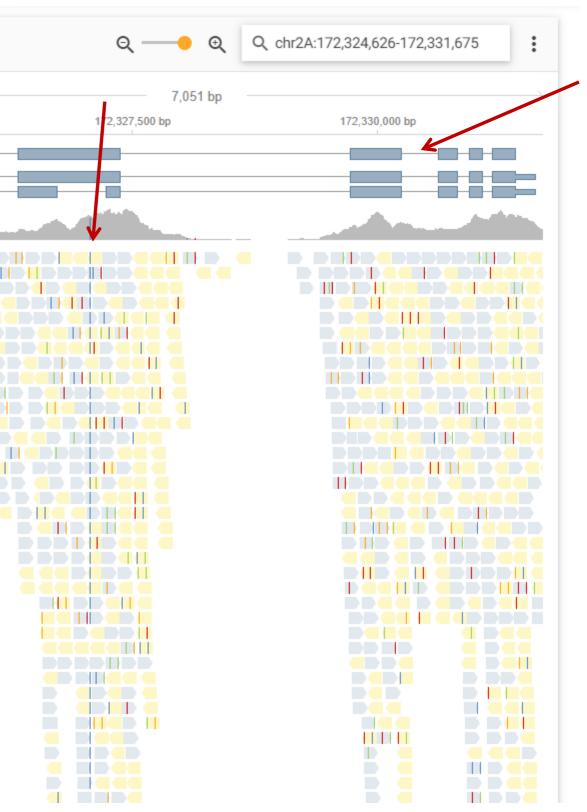
- Quickly browse and visualize aligned reads and depths from samples of any size, anywhere in the genome
- Reference projected & de-novo gene annotations from 10+ Wheat Genomes
- Quickly analyze areas of interest

Triticum aestivum ArinaLr Triticum aestivum ArinaLr Sample (ArinaLrFor) Alignm ents Showing 2,203 alignments. Excluded 87 of low quality. Removed 520 duplicates. Excluded 18 with low alignment identity. Excluded 2 with large pair distance.	<	172,200,000 bj	р
Sample (ArinaLrFor) Alignm ents Showing 2,203 alignments. Excluded 87 of low quality. Removed 520 duplicates. Excluded 18 with low alignment identity. Excluded 2 with large pair	Triticum aestivum ArinaLr 🔨		
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ccluded 87 of low quality. emoved 520 duplicates. ccluded 18 with low ignment identity. ccluded 2 with large pair			
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xcluded 18 with low lignment identity. xcluded 2 with large pair	xcluded 87 of low quality.		
lignment identity. Excluded 2 with large pair	emoved 520 duplicates.		



- Quickly browse and visualize aligned • reads and depths from samples of any size, anywhere in the genome
- Reference projected & de-novo gene • annotations from 10+ Wheat Genomes
- Quickly analyze areas of interest •
- Dynamically visualize impact of filtering • options
- Access gene transcript details •

Triticum aestivum Ari	naLr 🔨
Triticum aestivum Ari	naLr 🔨
Sample (ArinaLrFor ents) Alignm
Showing 1,200 alig	jnments.
Excluded 25 of low	/ quality.
Removed 295 dup	licates.
Excluded 9 with lo alignment identity.	
Excluded 1 with lar	rge pair

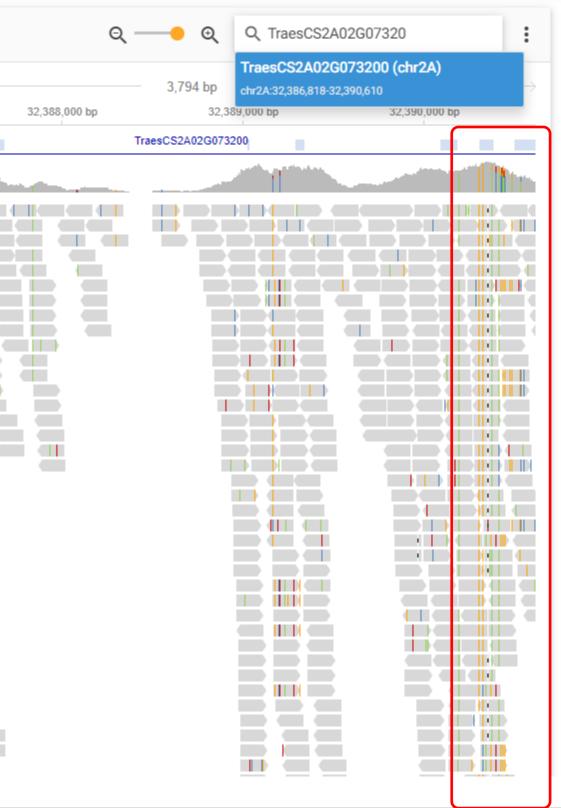


- Quickly browse and visualize aligned reads and depths from samples of any size, anywhere in the genome
- Reference projected & de-novo gene annotations from 10+ Wheat Genomes
- Quickly analyze areas of interest
- Dynamically visualize impact of filtering options
- Access gene transcript details
- Identified in ArinaLrFor as gene id "TraesARI2A01G085800"

i) General Details	
Feature Id:	TraesARI2A01G085800.1
Source:	PGSBv2.0
Туре:	mRNA
Position:	chr2A:35,779,864-35,783,656
Strand:	Negative (Reverse)
Name:	N/A
Coding Region:	chr2A:35,779,864-35,783,656
E Feature Attributes	
Associated Gene	
Transcript Exons	

- Quickly browse and visualize aligned reads and depths from samples of any size, anywhere in the genome
- Reference projected & de-novo gene annotations from 10+ Wheat Genomes
- Quickly analyze areas of interest
- Dynamically visualize impact of filtering options
- Access gene transcript details
- Identified in ArinaLrFor as gene id "TraesARI2A01G085800"
- IWGSC equivalent: TraesCS2A02G073200
- Landmark equivalent: TraesLDM2A01G082900

/	Alignments	
	IWGSC - High Confidence <	22,387,000 bp

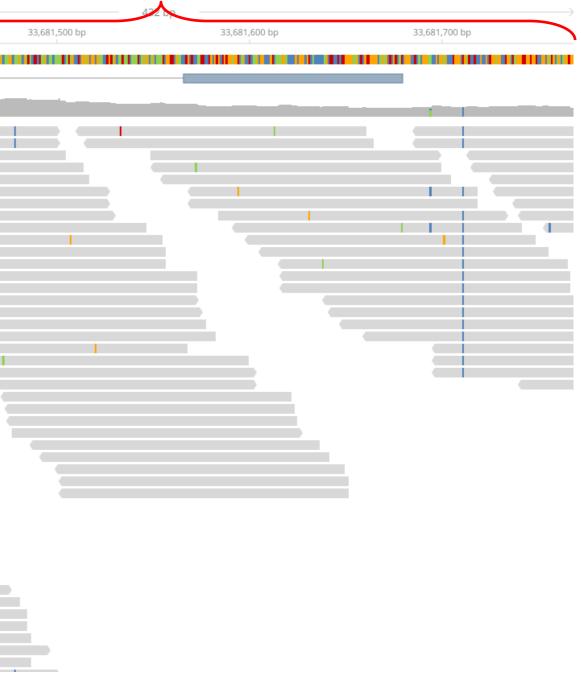


Comparing "10+ Wheat Genomes" Read Mapping

IWGSC "TraesCS2A02G073200" (Showing: chr2A:32,390,266-32,390,696)

32,390,300 bp	32,390,400 bp	32,390,500 bp	32,390,600 bp 32,3	33,681,400 bp

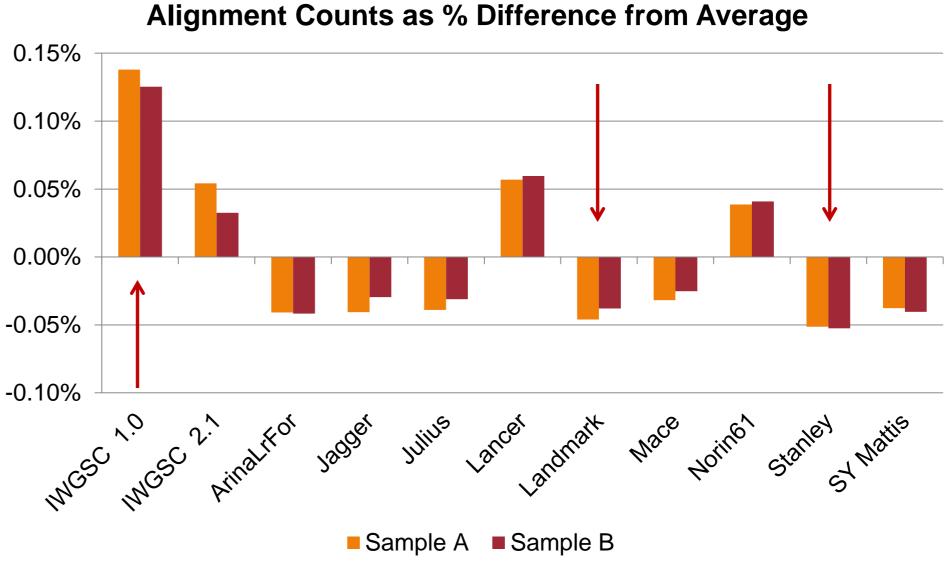
Landmark "TraesLDM2A01G082900" (Equivalent: chr2A:33,681,338-33,681,768)



Read Alignment Across "10+ Wheat Genomes"

	Sample A	Sample B
IWGSC 1.0	130,401,964	158,730,060
IWGSC 2.1	130,292,935	158,582,884
ArinaLrFor	130,169,124	158,465,361
Jagger	130,169,417	158,484,503
Julius	130,171,553	158,482,155
Lancer	130,296,404	158,625,929
Landmark	130,162,411	158,471,316
Mace	130,180,915	158,491,452
Norin61	130,272,621	158,596,217
Stanley	130,155,509	158,448,214
SY Mattis	130,173,317	158,467,435

129,567,060 157,465,504 Raw Reads:



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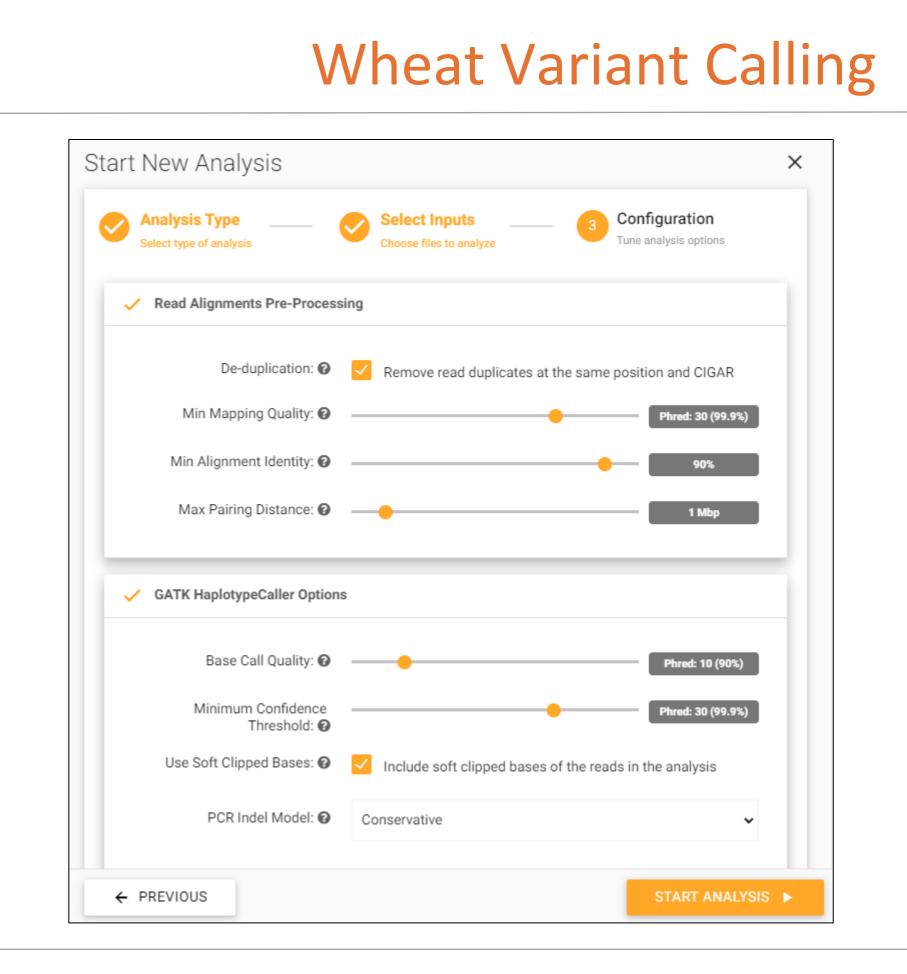
Variant Calling & Filtering Using 10+ Wheat Genomes Assemblies

- Various algorithms available to call variants, • including INDELS & reference alleles
- Alleles automatically called leveraging • associated 10+ Wheat Genomes reference assemblies

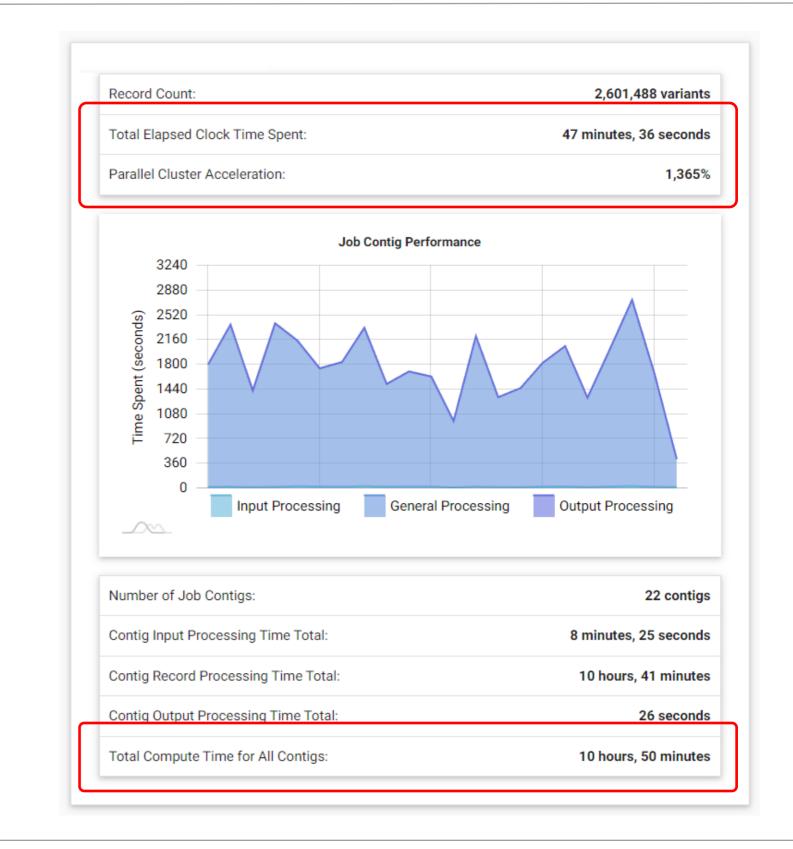
Start New Analysis	×						
Analysis Type 2 Select Inputs 3 Configuration Select type of analysis Choose files to analyze 3 Configuration							
Which type of data would you like to analyze?							
DNA-Seq							
O RNA-Seq							
Q Type here							
O Bowtie2 Alignment Map the reads of one or more single or paired-end samples to a reference genome using the Bowtie2 algorithm							
O BWA-MEM Alignment Map the reads of one or more single or paired-end samples to a reference genome using the BWA-MEM algorithm	Map the reads of one or more single or paired-end samples to a reference genome using the BWA-MEM						
O BWA-ALN Alignment Map the reads of one or more single or paired-end samples to a reference genome using the BWA-ALN							
algorithm							
O General Variant Detection Detect and call variants in one or more samples by comparing their aligned sequences to the reference							
 GATK Variant Detection (HaplotypeCaller) Call germline SNPs and indels via local re-assembly of haplotypes using GATK's HaplotypeCaller 							
O Analyze Coverage							
CONTINUE -	>						

Wheat Variant Calling

- Various algorithms available to call variants, • including INDELS & reference alleles
- Alleles automatically called leveraging • associated 10+ Wheat Genomes reference assemblies
- Experimentally adjust alignment pre-• processing options and tune for desired specificity & sensitivity
- **Convenient and highly-performant** • integration with the industry-standard GATK Haplotype Caller



- Various algorithms available to call variants, including INDELS & reference alleles
- Alleles automatically called leveraging associated 10+ Wheat Genomes reference assemblies
- Experimentally adjust alignment preprocessing options and tune for desired specificity & sensitivity
- Convenient and highly-performant integration with the industry-standard GATK Haplotype Caller
- Parallel processing technology efficiently processes even massive individual samples or large batches of multiple samples

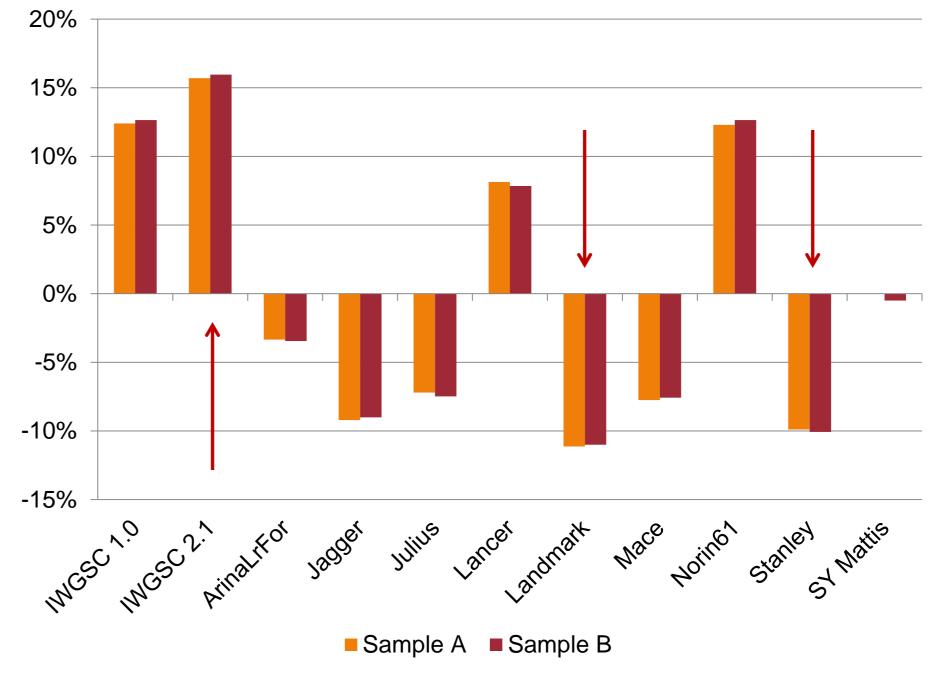


Wheat Variant Calling

Variant Calling Across 10+ Wheat Genomes

	Sample A	Sample B
IWGSC 1.0	2,705,646	3,035,134
IWGSC 2.1	2,785,165	3,124,243
ArinaLrFor	2,326,655	2,601,488
Jagger	2,185,424	2,451,528
Julius	2,233,777	2,492,644
Lancer	2,603,096	2,905,634
Landmark	2,139,171	2,397,919
Mace	2,220,472	2,490,270
Norin61	2,703,332	3,035,071
Stanley	2,168,964	2,422,944
SY Mattis	2,406,907	2,680,842

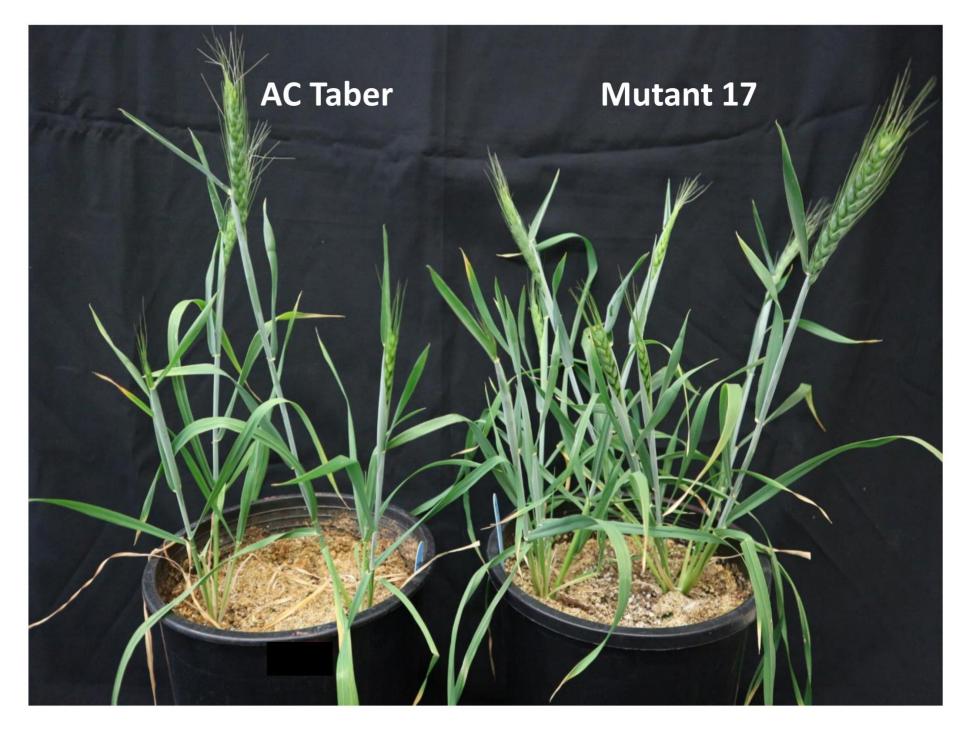




Variant Counts as % Difference from Average

Identifying Key Mutations

- AAFC project used mutagenesis to try and identify genotype of more drought tolerant wheat lines
- Under drought stress mutant shows: •
 - Higher biomass
 - Stress adaptation features
 - Comparable spike length
 - Increased number of spikelets
 - Higher seed weight
- <u>Abscisic acid</u> pathways direct drought response signaling



(Presented at PAG 2022, Neha Vaid)

Source: "Genomics-Driven Development of Drought Tolerant Wheat Cultivars",

Leveraging IWGSC Functional Annotations

- 78 genes (85 transcripts) reference <u>abscisic acid</u> (ABA) within their IWGSC functional descriptions
- Gene names are based on the IWGSC
 1.0 naming convention, which is easy to translate to the IWGSC 1.1 convention
- Translation table from the "10+ Wheat Genome" project can then be used to identify equivalent genes
- Example of translating the IWGSC gene with name "TraesCS2A01G317000"

Genome
IWGSC 1.0
IWGSC 1.1
ArinaLrFor
Jagger
Julius
Landmark
LongReach Lancer
Mace
Norin 61
Stanley
SY Mattis

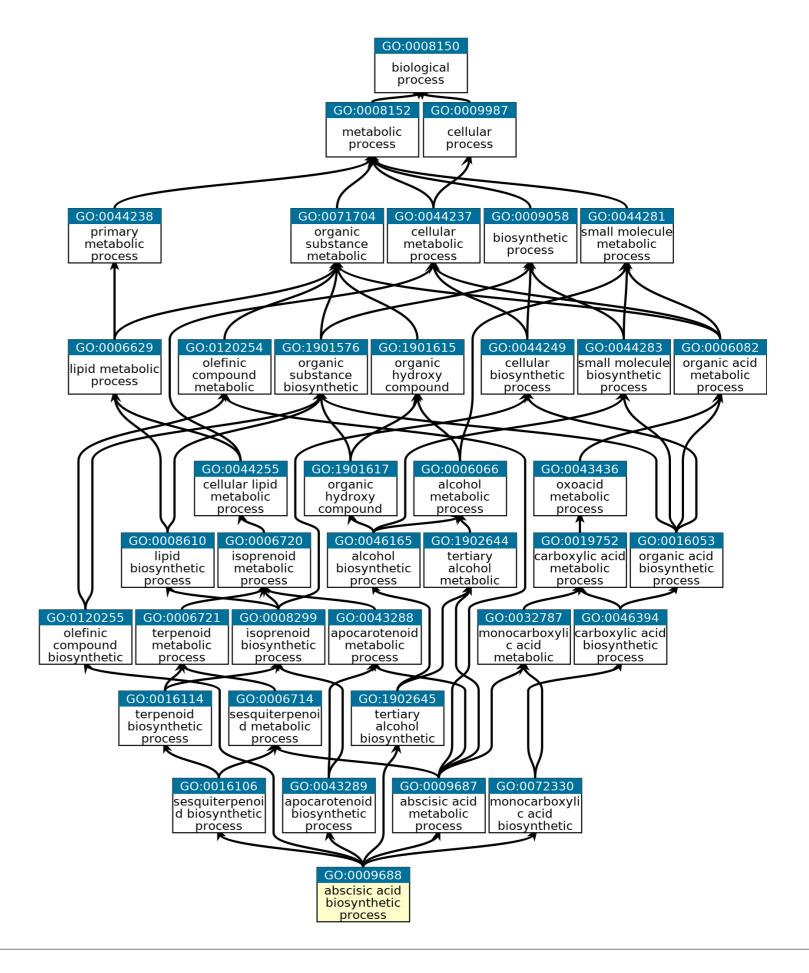
Gene

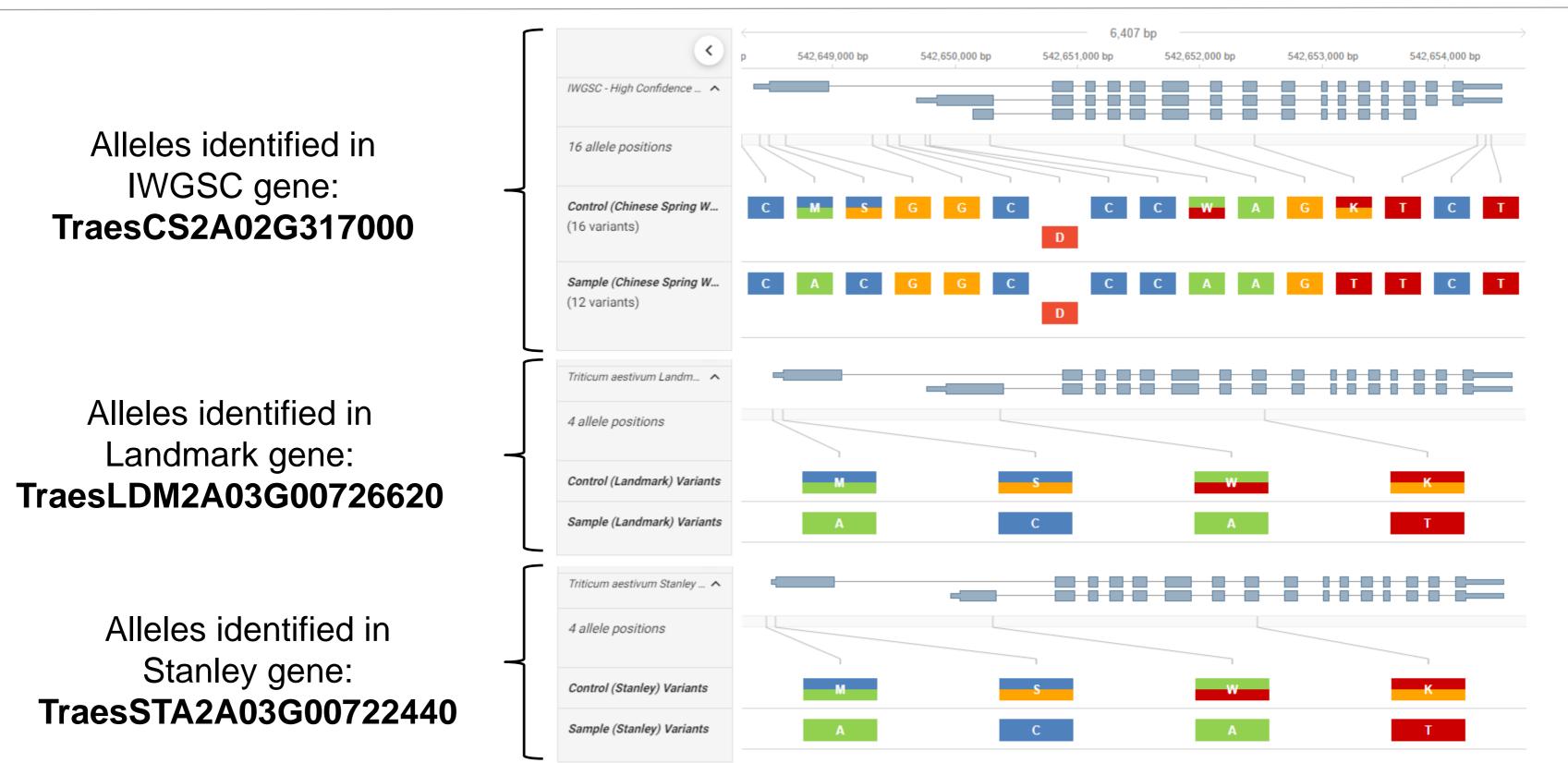
TraesCS2A01G317000 TraesCS2A02G317000 TraesARI2A01G344100 TraesJAG2A01G345500 TraesJUL2A01G344300 TraesLDM2A01G344000 TraesLAC2A01G344600 TraesMAC2A01G334600 TraesNOR2A01G347400 TraesSTA2A01G333700 TraesSYM2A01G334400

Gene Ontology for Example Gene

- IWGSC annotations show <u>TraesCS2A02G317000</u> associated with 'GO:0009688'
- Ancestor tree for this gene ontology shown
- GO:0009688, biological process:
 abscisic acid biosynthetic process
- GO:0009688 definition:
 - The chemical reactions and pathways resulting in the formation of abscisic acid, 5-(1-hydroxy-2,6,6,trimethyl-4-oxocyclohex-2-en-1-y1)-3methylpenta-2, 4-dienoic acid.

Source: https://www.ebi.ac.uk/QuickGO/GTerm?id=GO:0009688

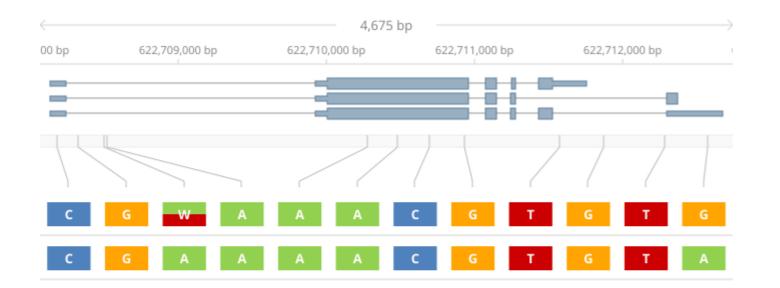




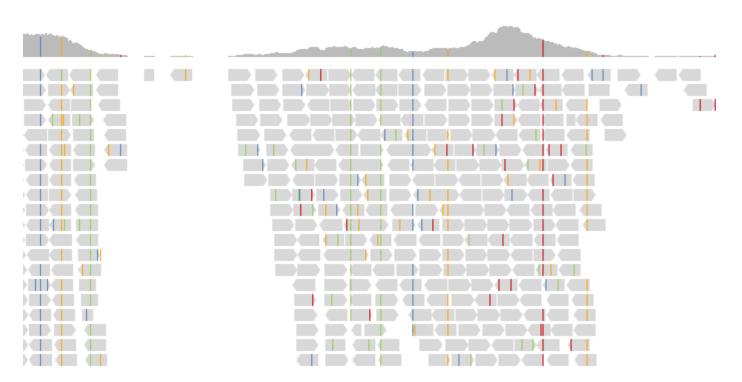


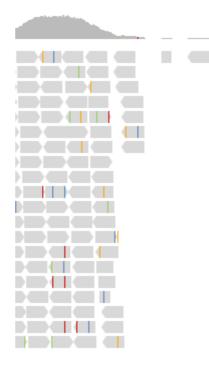
Alleles and Mapped Reads Using IWGSC (Showing Gene: TraesCS3A02G371900)

Same Samples Mapped to Landmark (Equivalent Gene: TraesLDM3A03G01467370)

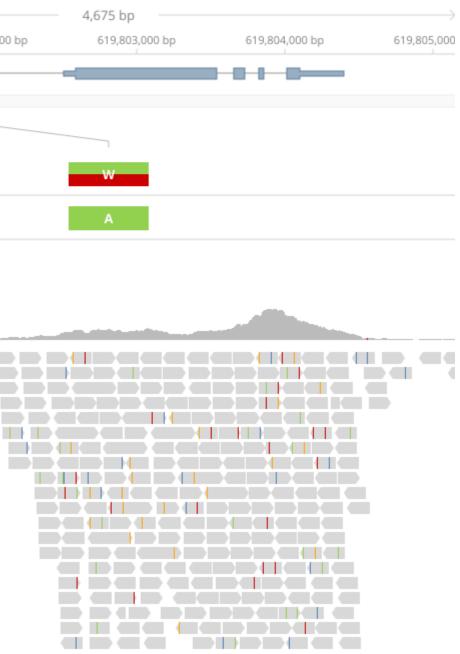


619,801,000 bp 619,802,000 bp

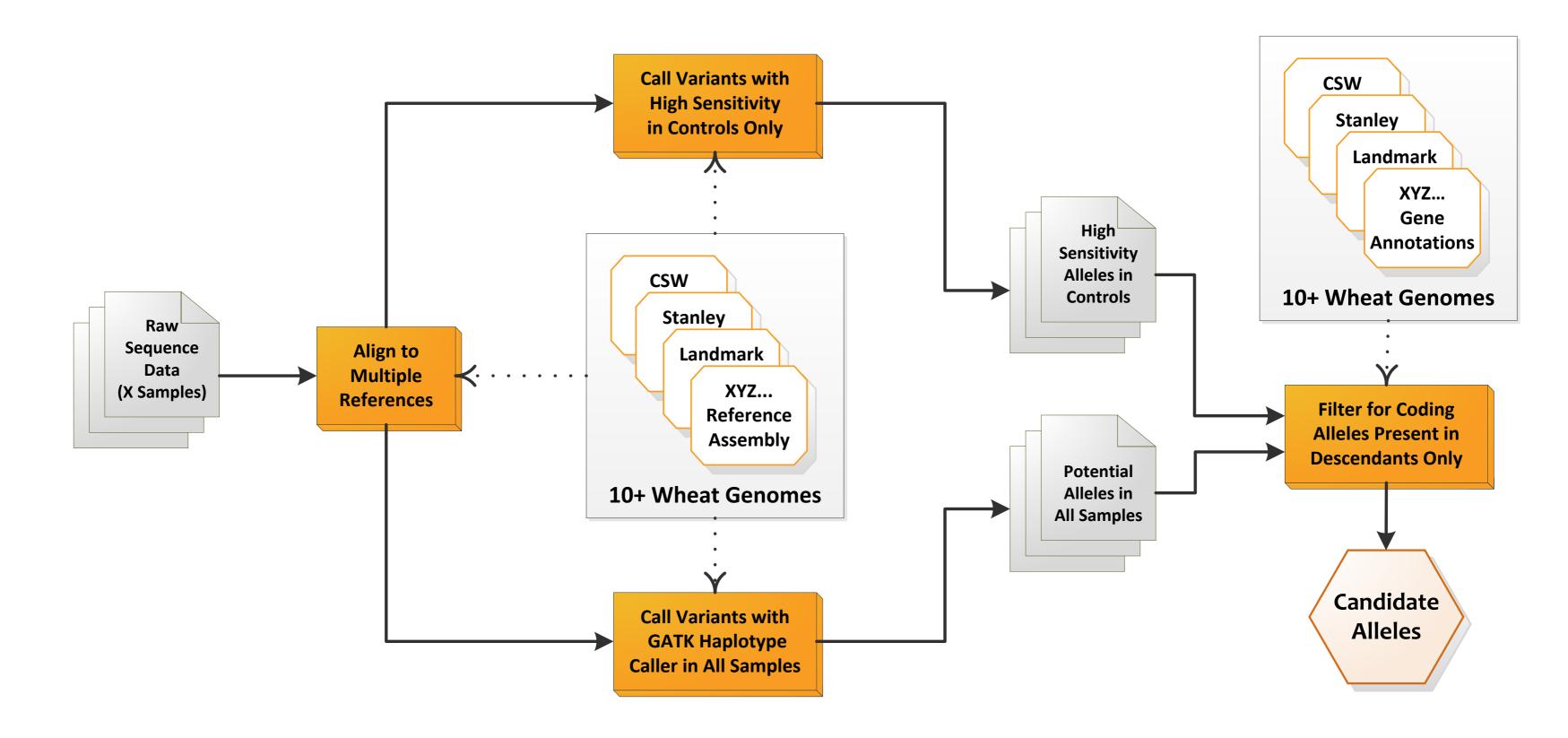


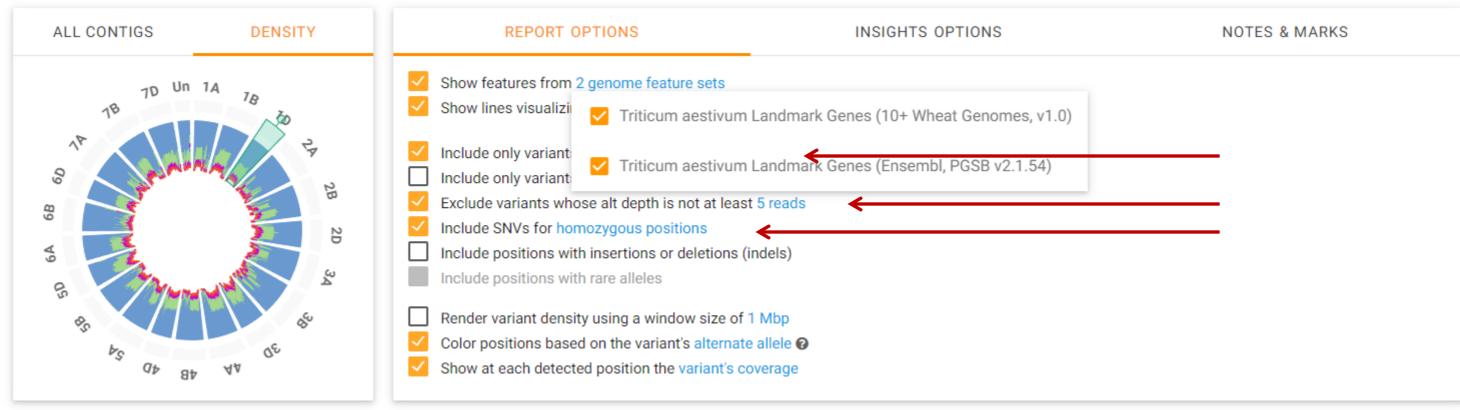


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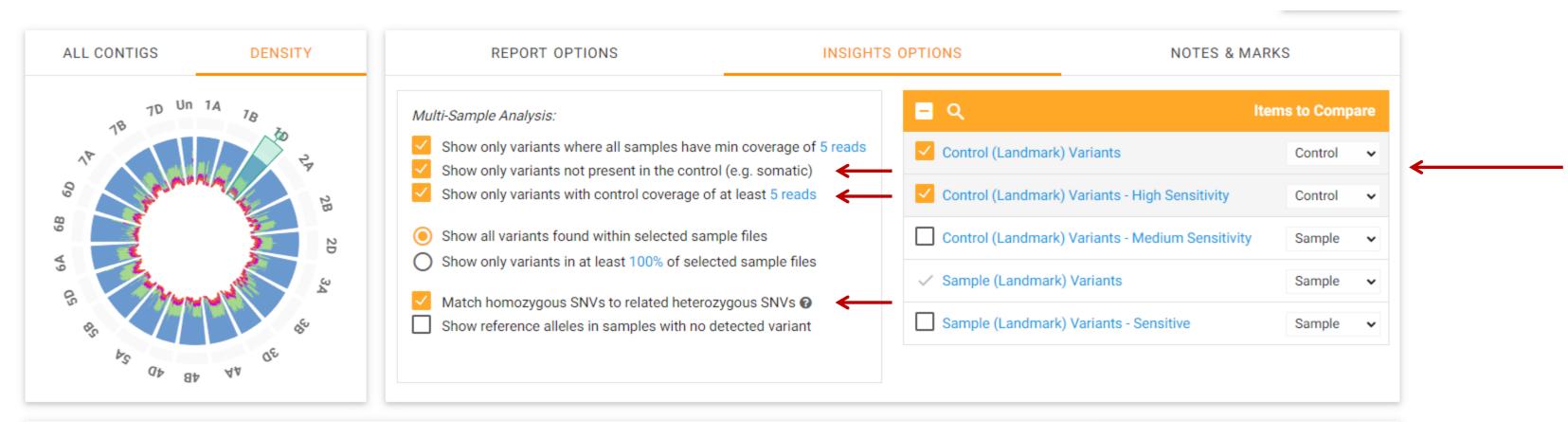
Candidate Alleles Analysis Approach

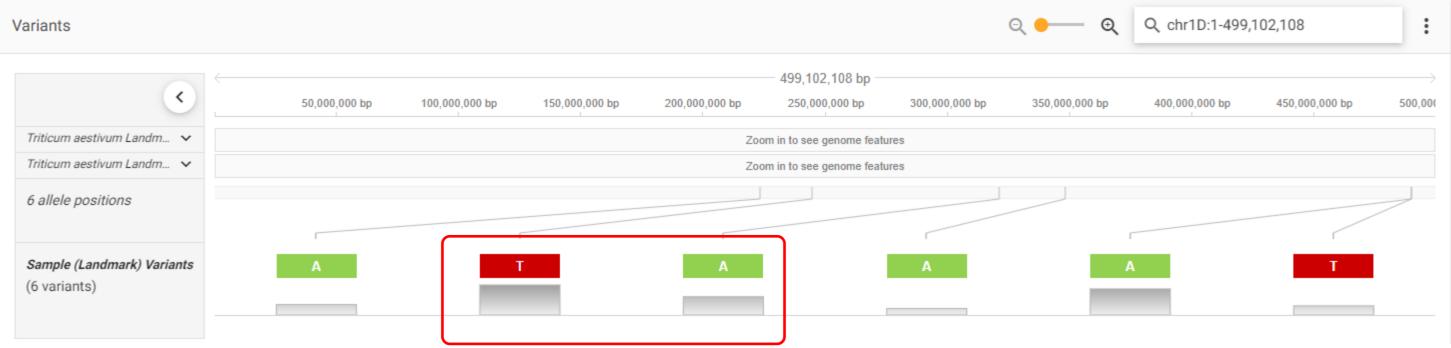




Candidate Allele Filtering





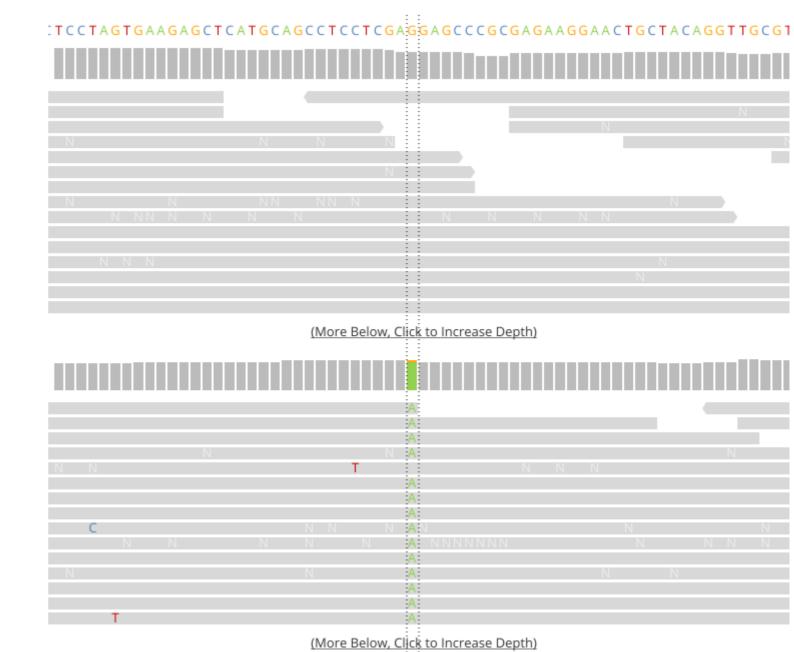


Leveraging the IWGSC Gene Annotations with Additional Cultivars in Curio / Presented at PAG 2023

Candidate Allele Filtering



Gene: TraesLDM1D01G192000 (chr1D:244,524,468-244,524,532)





Leveraging the IWGSC Gene Annotations with Additional Cultivars in Curio / Presented at PAG 2023

Candidate Allele Filtering

Gene: TraesLDM1D01G256200 (chr1D:321,109,370-321,109,434)

Candidate Allele Filtering (SNV Counts)

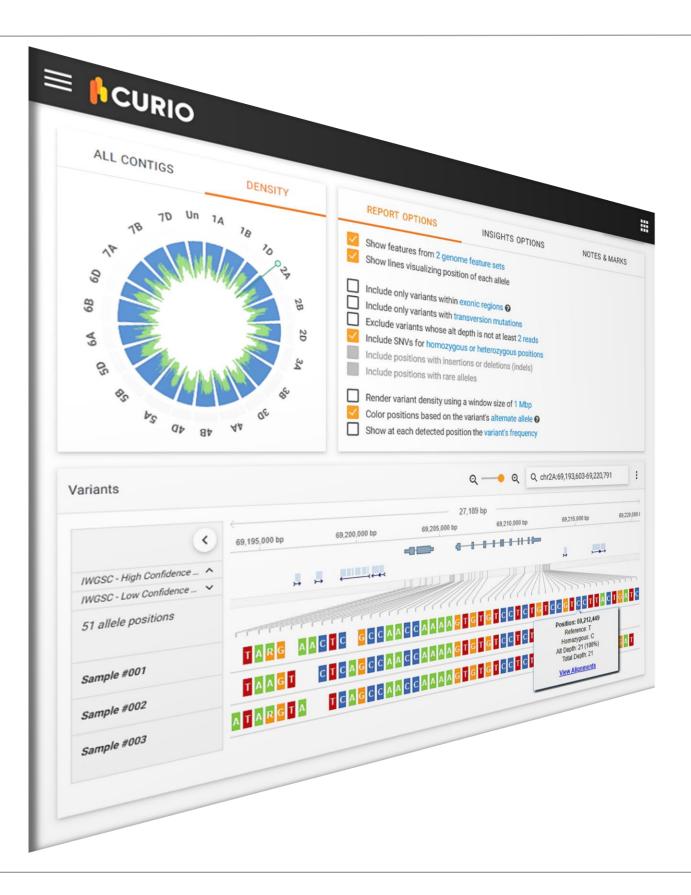
	<u>IWGSC 2.1</u>	IWGSC 1.0	<u>Stanley</u>	Landmark
Detected Potential Polymorphisms (SNVs)	2,820,397	2,751,824	2,190,860	2,168,141
Only Those Unique to Mutant Line	167,601	166,288	159,369	154,670
Require Minimum Number of Reads to Demonstrate Allele	42,061	41,398	37,565	35,812
Limit to Exonic Coding Regions (10+ Wheat Genomes)	11,161	11,305	10,979	10,448
Remove Compatible Heterozygous Matches	3,238	3,400	3,481	3,413
Candidate Homozygous Polymorphisms Only	28	35	39	41



In Conclusion

Looking Ahead

- Continued incorporation of new public reference assemblies, gene annotations, and variant call sets
- Enabling pangenome analysis, graph construction, alignment, variant calling , and visualization of sample data
- Enabling haplotype-based analysis approaches, both genome wide and gene based
- Incorporation of other data types, such as SNP array data
- Additional crop-research-specific analysis types and interpretive visualizations



Acknowledgements

Special thanks to collaborators with the Agriculture and Agri-Food Canada (AAFC) project:

- Marcus Samuel, PhD (Professor), University of Calgary •
- Raju Soolanayakanahally, PhD (Research Scientist), AAFC, Saskatoon •
- Sateesh Kagale (Research Officer), National Research Council, Saskatoon •
- Neha Vaid, PhD (Research Associate), University of Calgary / AAFC, Saskatoon •

And the collective efforts of the:

- International Wheat Genome Sequencing Consortium (IWGSC)
- 10+ Wheat Genomes Project •

