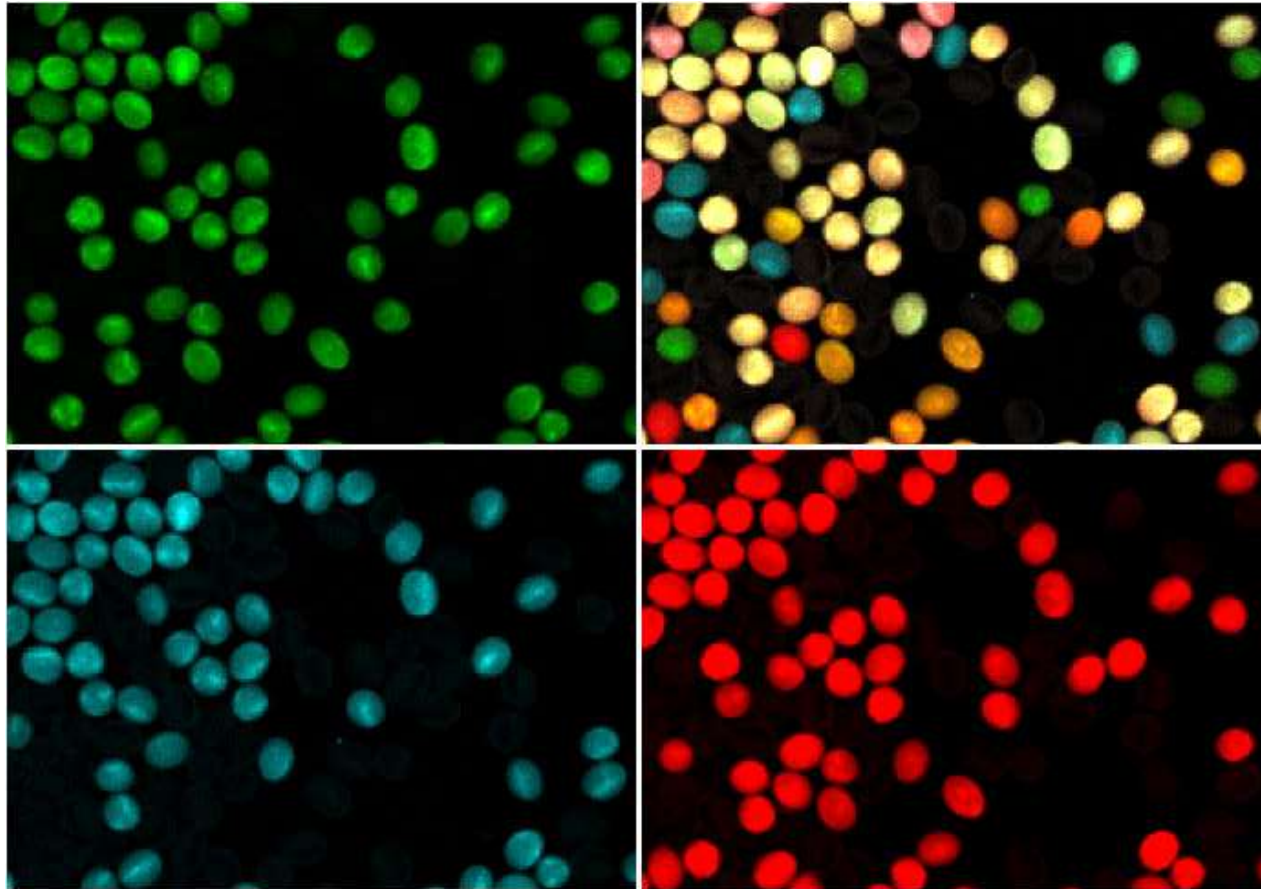
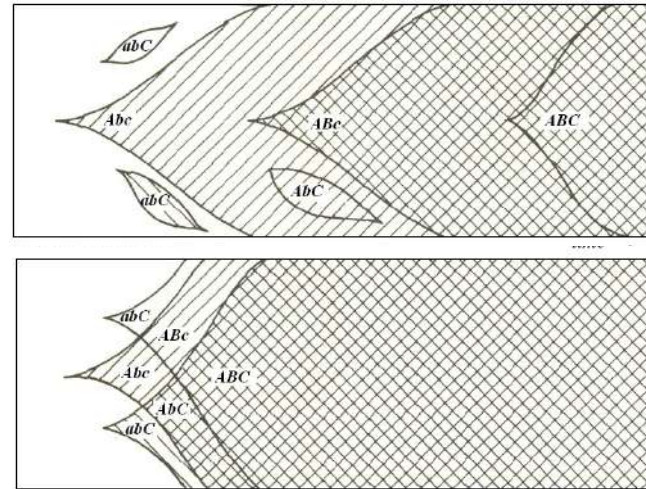
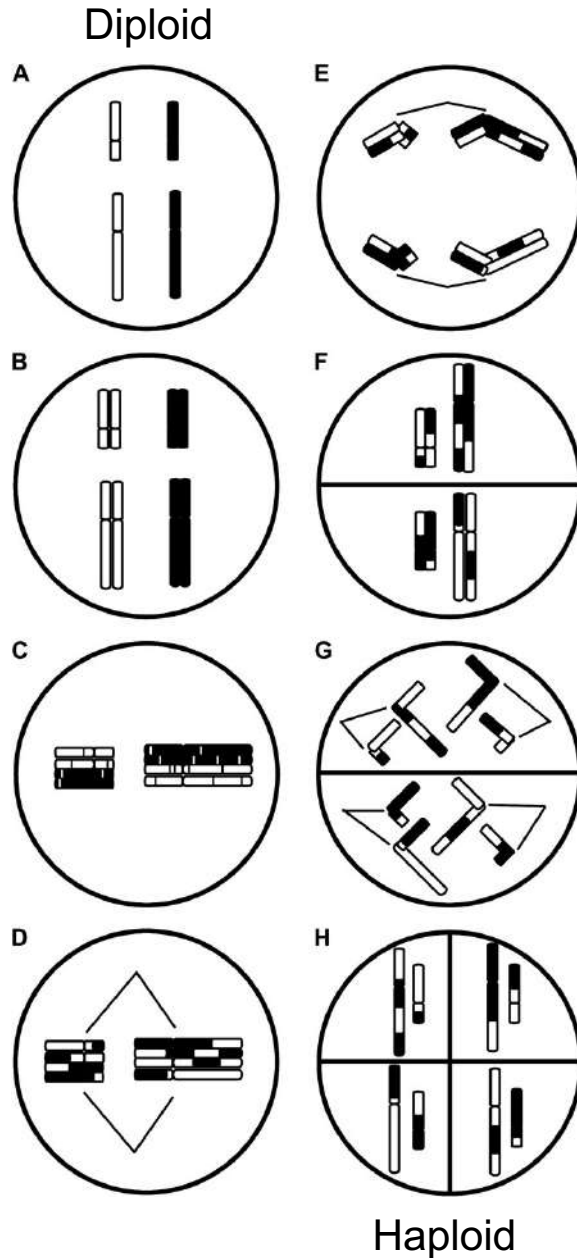


Recombination and chromatin landscapes in the wheat genome



Ian Henderson, University of Cambridge
IWGSC webinar, Jan 2022

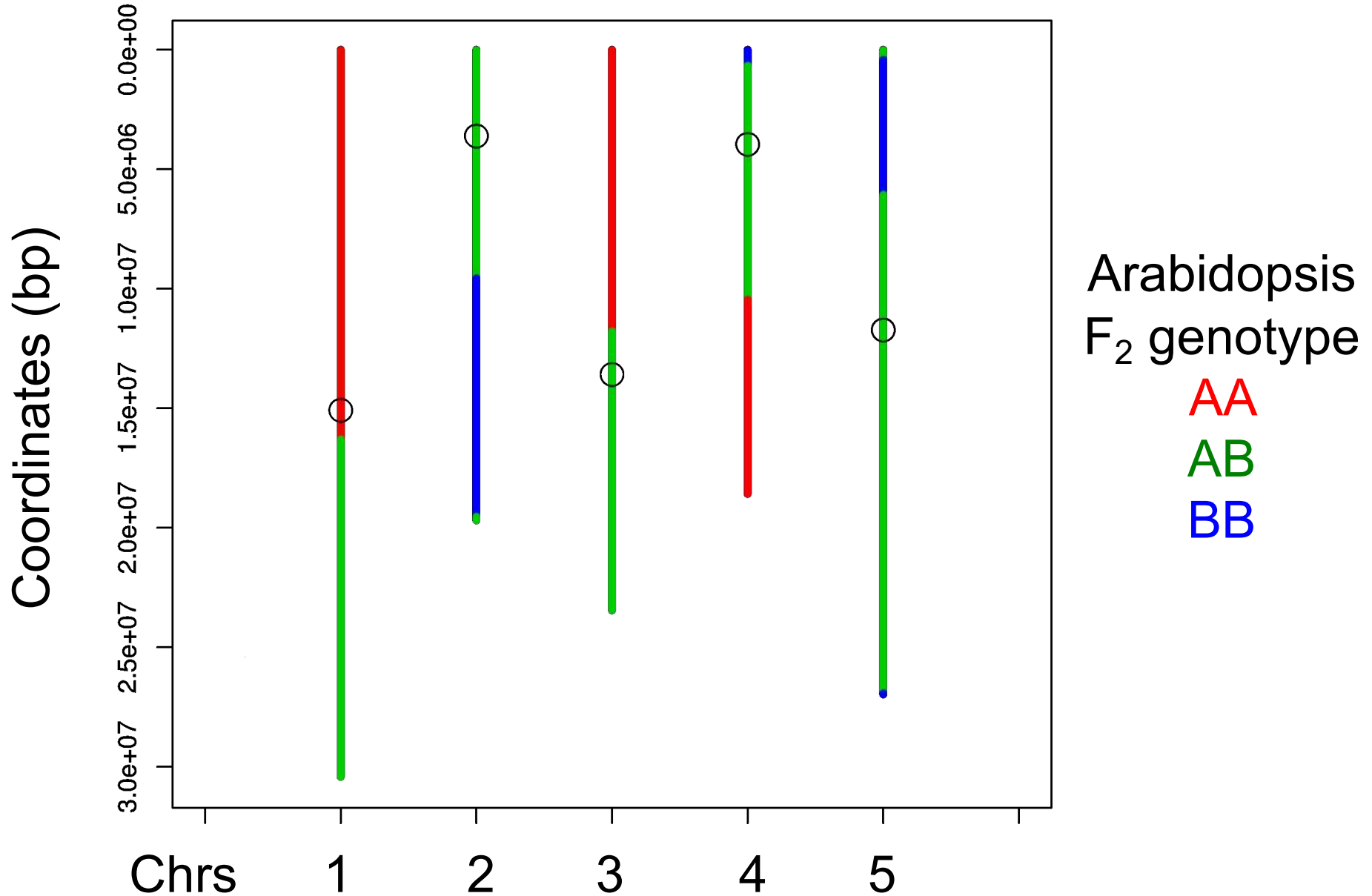
Meiosis, recombination and evolution














Fisher (1930) *The Genetical Theory of Natural Selection*

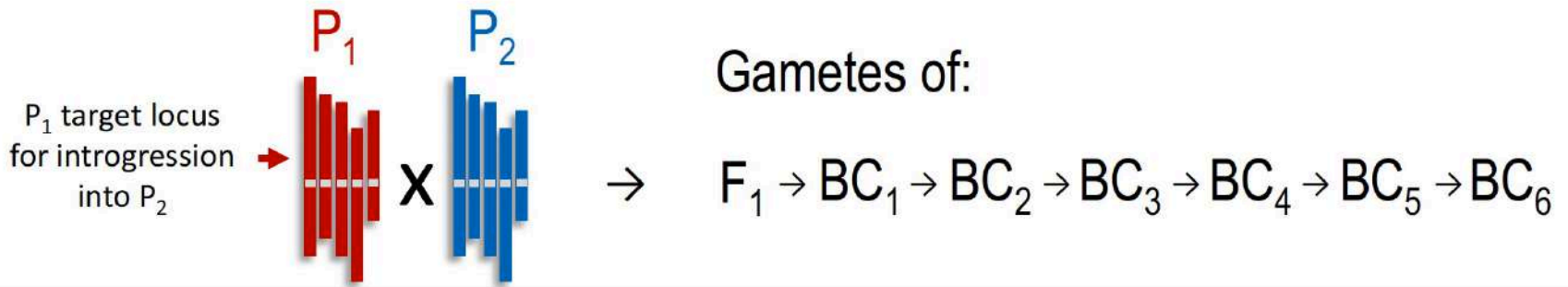
Muller (1932) *American Naturalist*

Meiosis generates genetic diversity

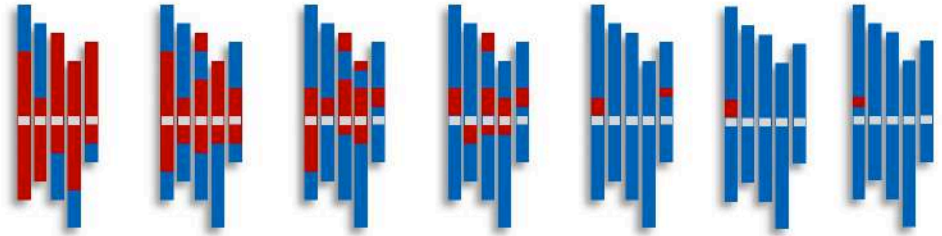


Addressing Research Bottlenecks to Crop Productivity

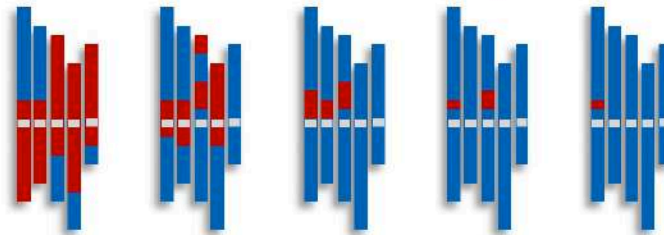
Matthew Reynolds ^{1,*} Owen K. Atkin ^{2,*,@} Malcolm Bennett ^{3,*} Mark Cooper ⁴ Ian C. Dodd,⁵
M. John Foulkes,³ Claus Frohberg,⁶ Graeme Hammer ⁴ Ian R. Henderson,⁷ Bingru Huang,^{8,*}
Viktor Korzun ⁹ Susan R. McCouch ^{10,*} Carlos D. Messina ^{11,*} Barry J. Pogson ^{2,@}
Gustavo A. Slafer ^{12,13,*} Nicolas L. Taylor ^{14,@} and Peter E. Wittich^{15,*}



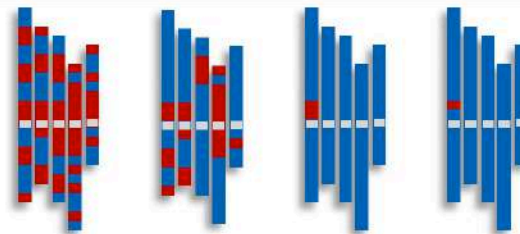
Wild type recombination



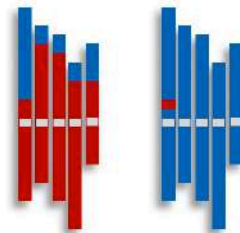
Unlocking centromere proximal crossover cold spots



Genome-wide hyper-recombination

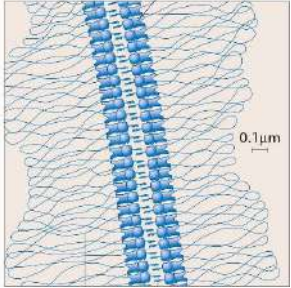


Targeted recombination



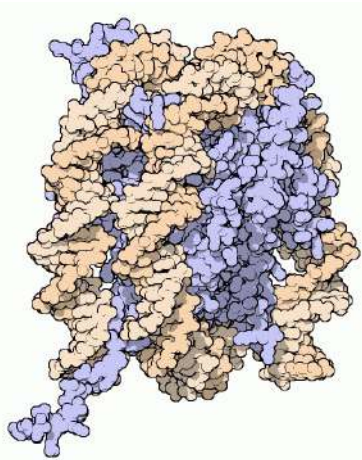
Meiotic recombination pathways

Chromatin, axis, SC

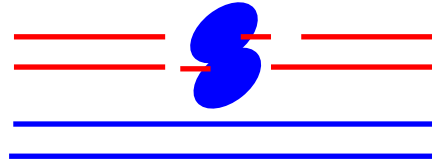


ASY1
 ASY3
 REC8
 ZYP1a
 ZYP1b

H2A.Z
 H3K4^{me3}
 5-mC
 H3K9^{me2}

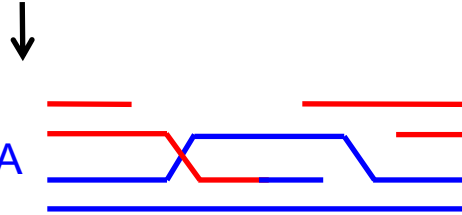


SPO11-1/SPO11-2/
MTOPIVIB

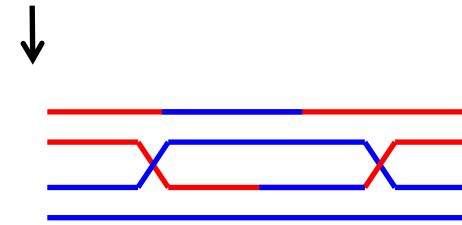


~200 DNA breaks

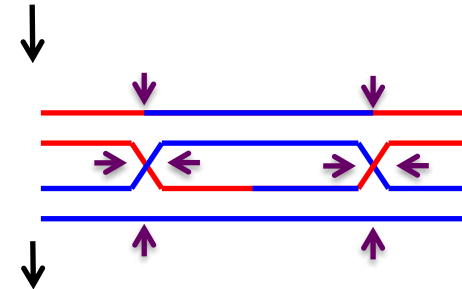
DMC1, RAD51, RPA



HEI10, ZIP4,
MSH4, MSH5,
MER3, SHOC1,
PTD



MLH1, MLH3



Crossover

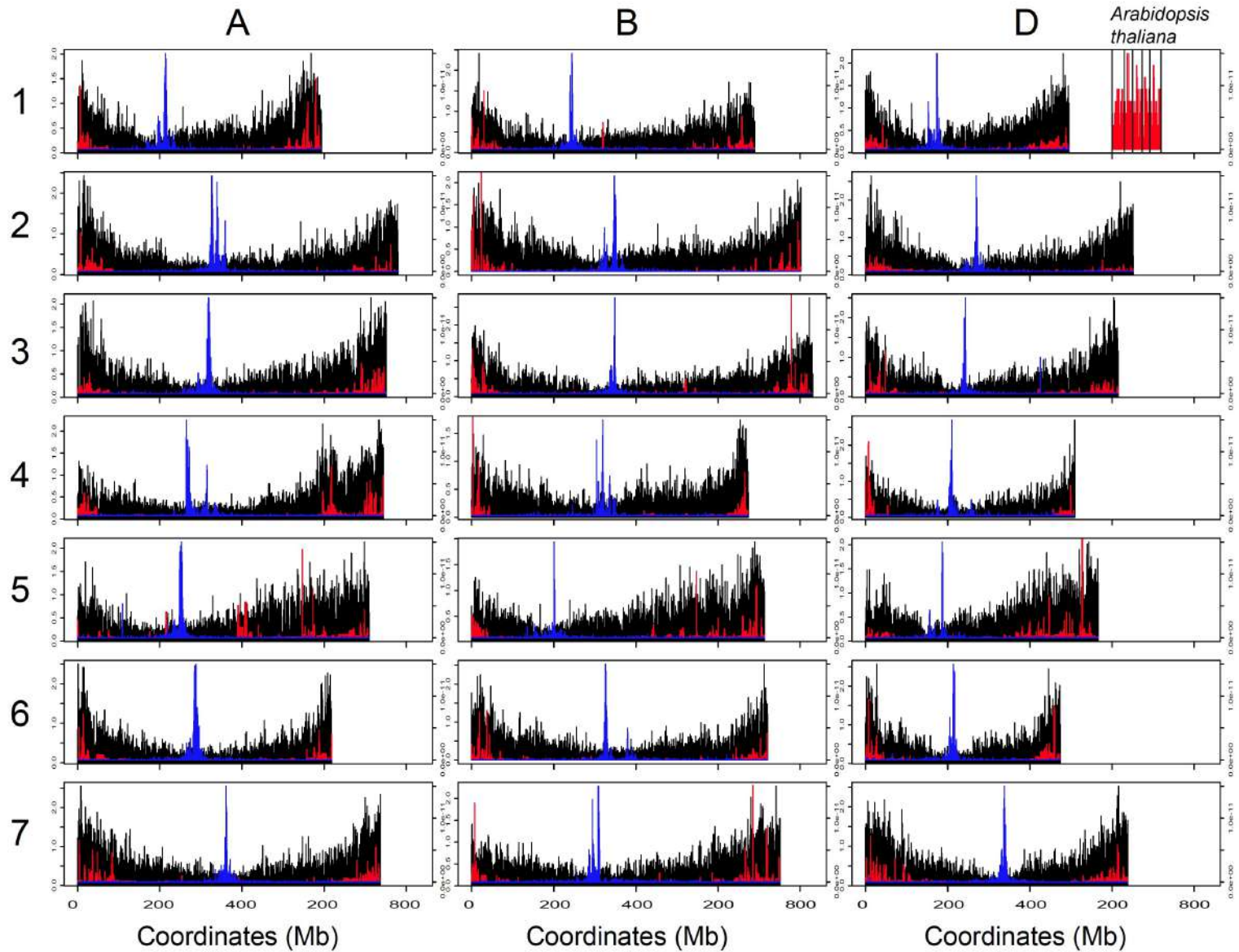


10 Crossovers

Anti-
Crossover

FANCM
 MHF1
 MHF2
 FIGL1
 TOP3a
 RECQ4a
 RECQ4b
 HCR1

Recombination in the wheat genome vs Arabidopsis

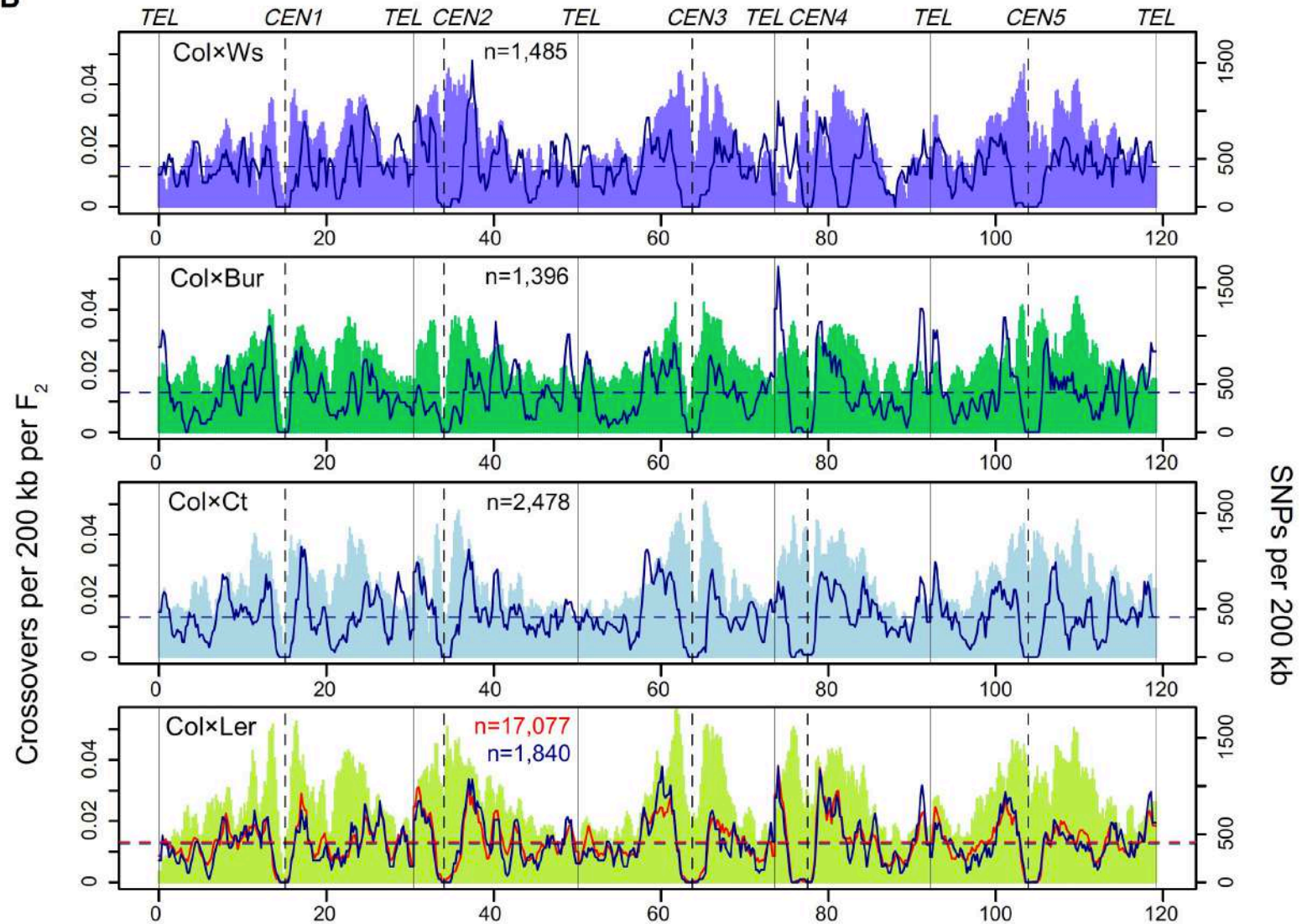


■ H3K4me3:H3K9me2
■ Gypsy LTR Cereba
■ cM/Mb

BBSRC sLola
CS x Renan cM/Mb data from Pierre Sourdille

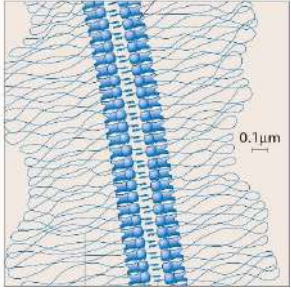
Crossover landscapes in the Arabidopsis genome

B



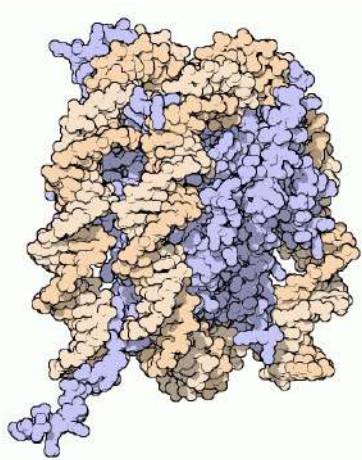
Meiotic recombination pathways

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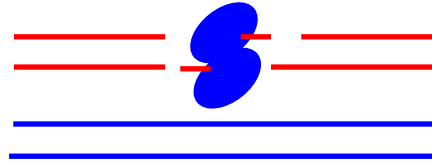


ASY1
 ASY3
 REC8
 ZYP1a
 ZYP1b

H2A.Z
 H3K4^{me3}
 5-mC
 H3K9^{me2}

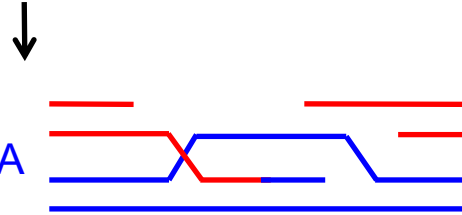


SPO11-1/SPO11-2/
MTOPVIB

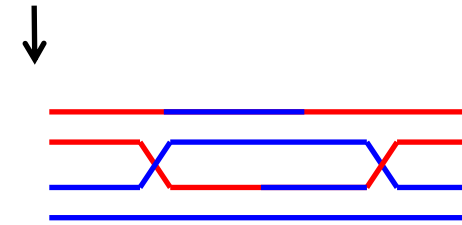


~200 DNA breaks

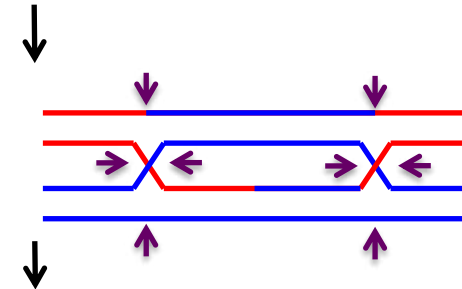
DMC1, RAD51, RPA



HEI10, ZIP4,
MSH4, MSH5,
MER3, SHOC1,
PTD



MLH1, MLH3



Crossover

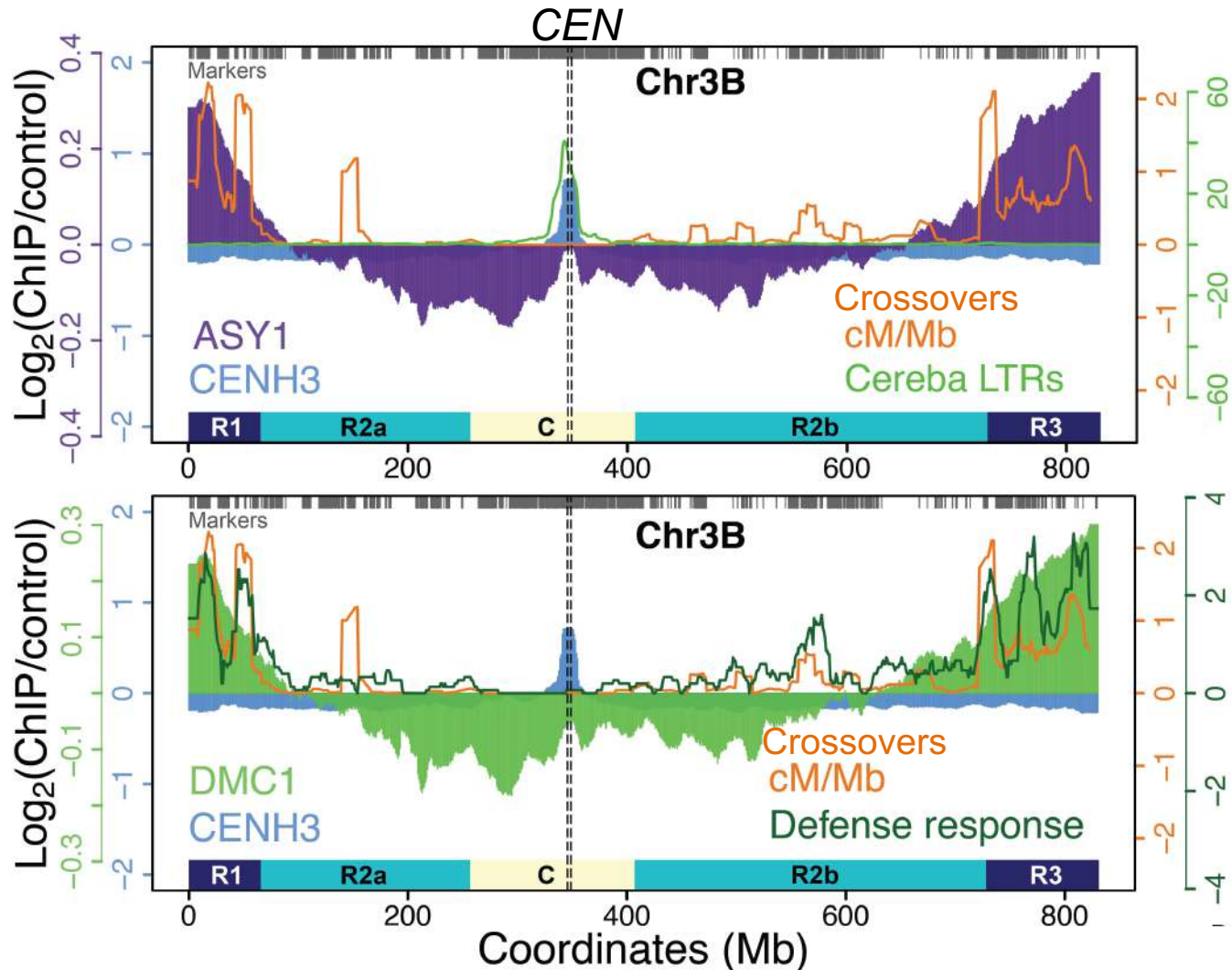


10 Crossovers

Anti-
Crossover

FANCM
 MHF1
 MHF2
 FIGL1
 TOP3a
 RECQ4a
 RECQ4b
 HCR1

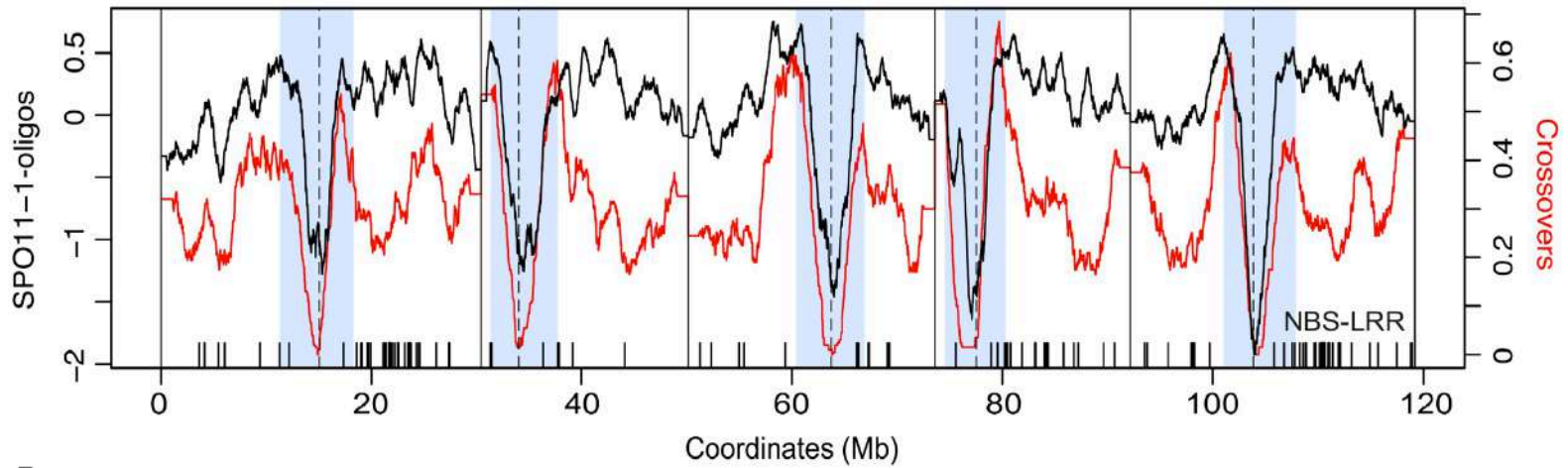
Polarized crossovers, ASY1 and DMC1 in wheat



ChIP for ASY1
and DMC1
performed
against CS
flowers

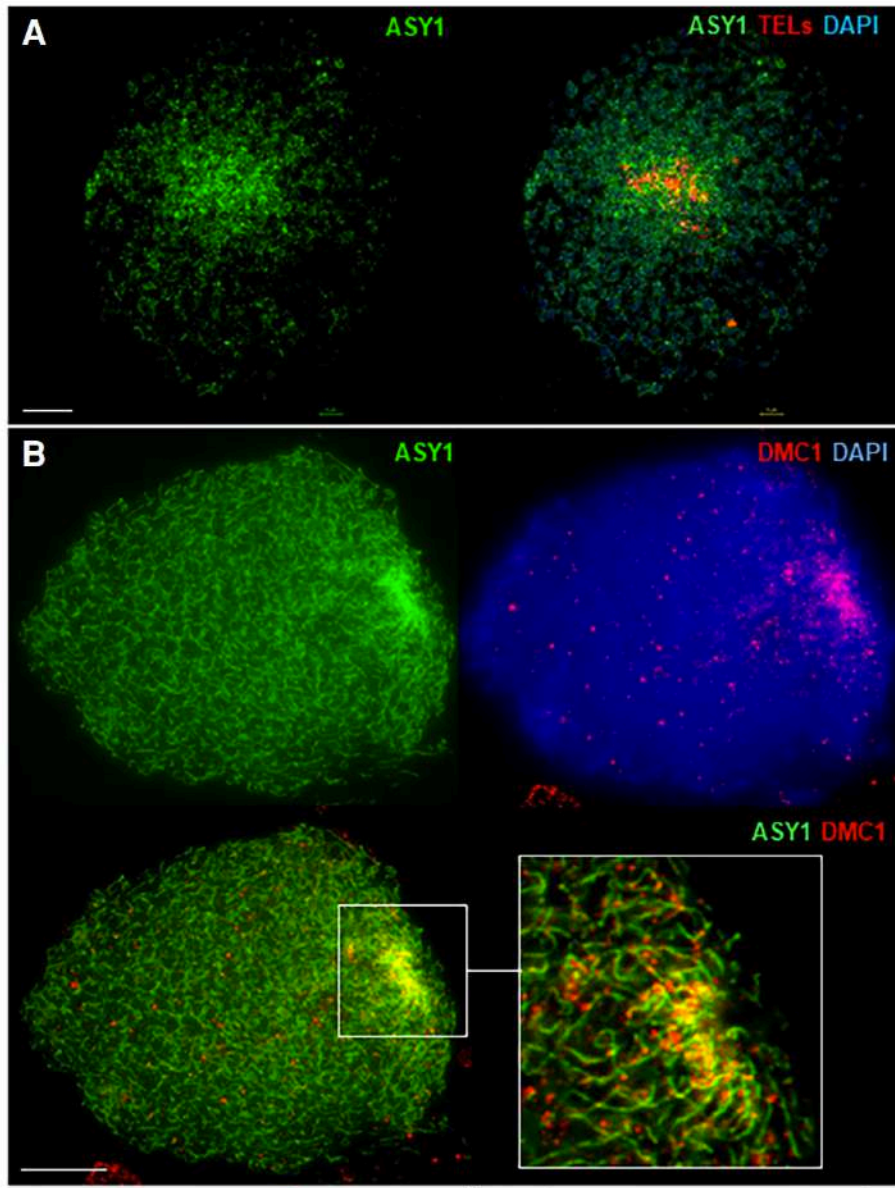
Andrew Tock & Dan Holland (2021) *Genome Res*
cM/Mb from Appels et al (2018) *Science*

ASY1 and SPO11-1-oligos in Arabidopsis



Choi et al (2018) *Genome Res*

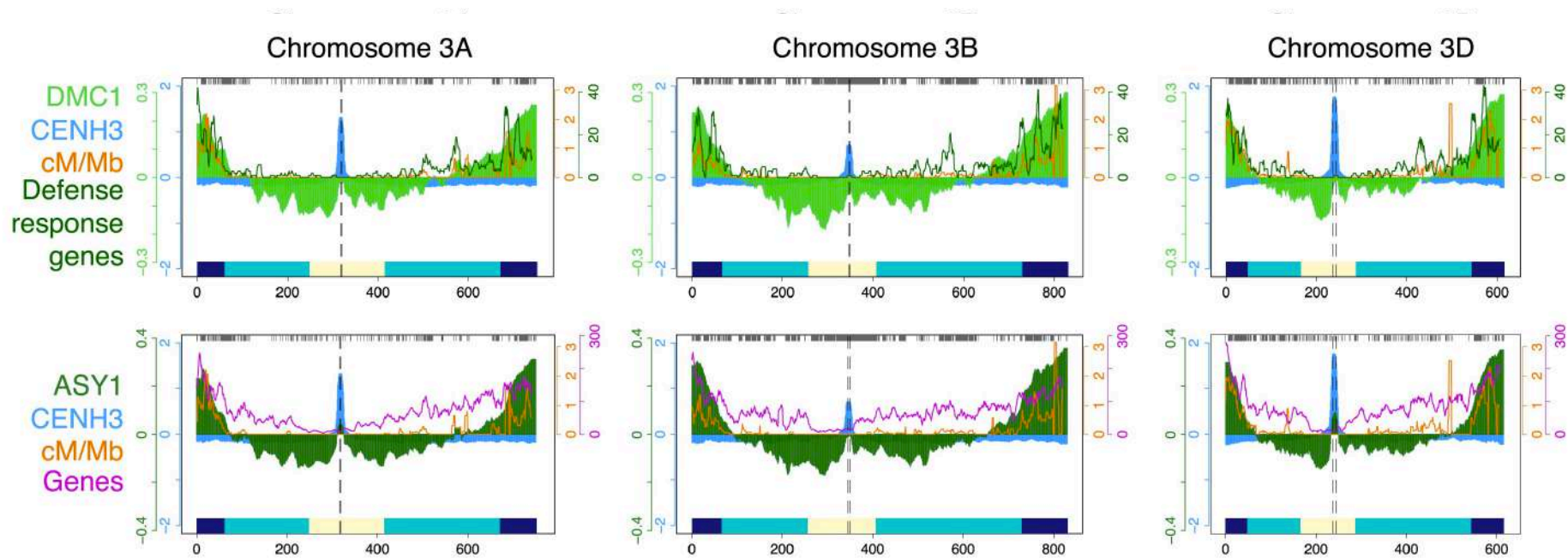
Polarized recombination in bread wheat



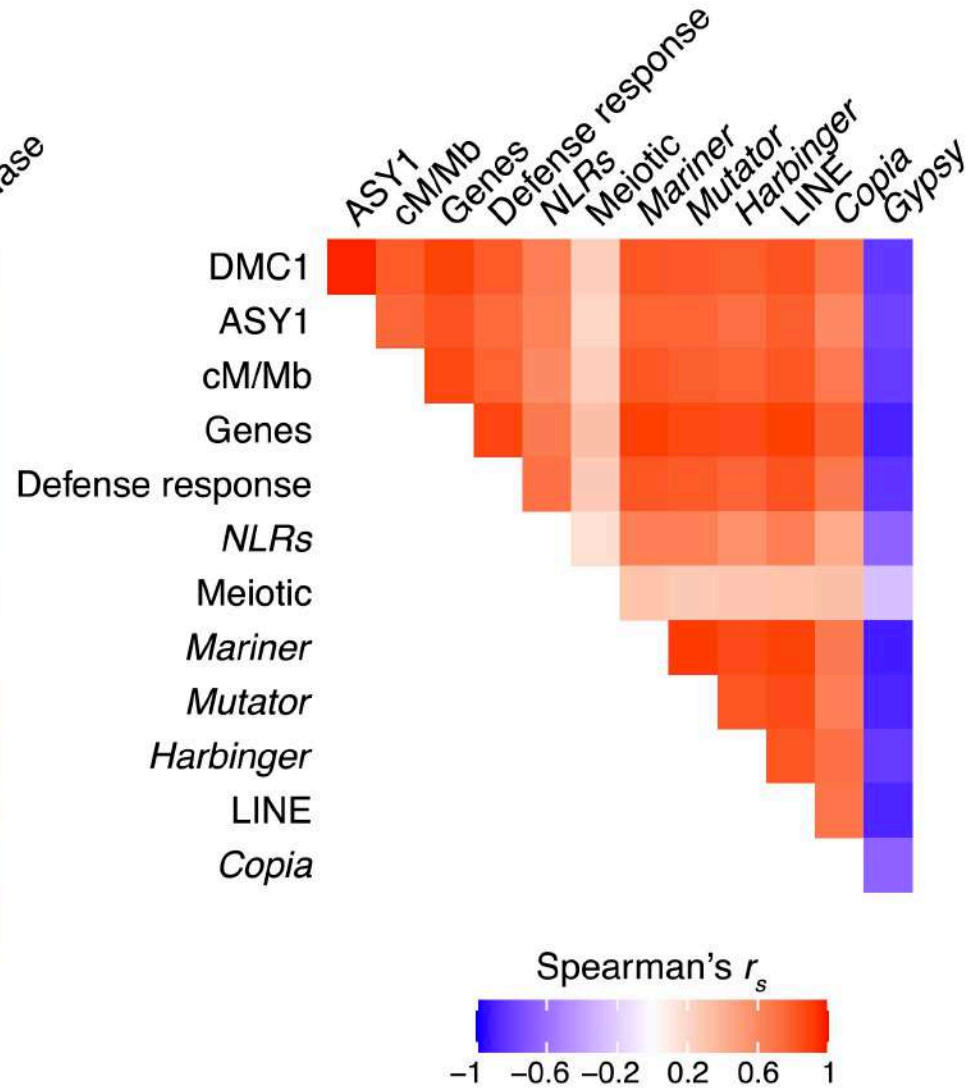
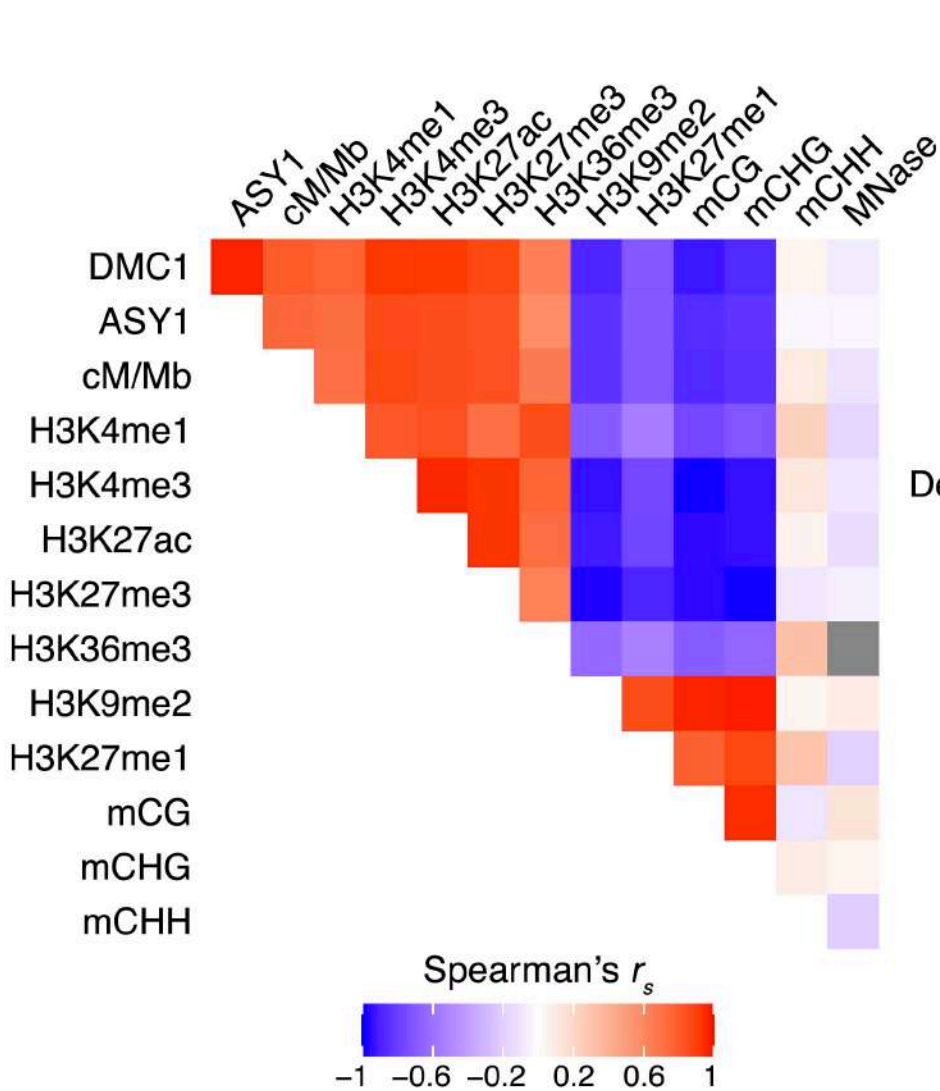
Kim Osman, Eugenio
Sanchez-oran and
Chris Franklin

BBSRC sLola

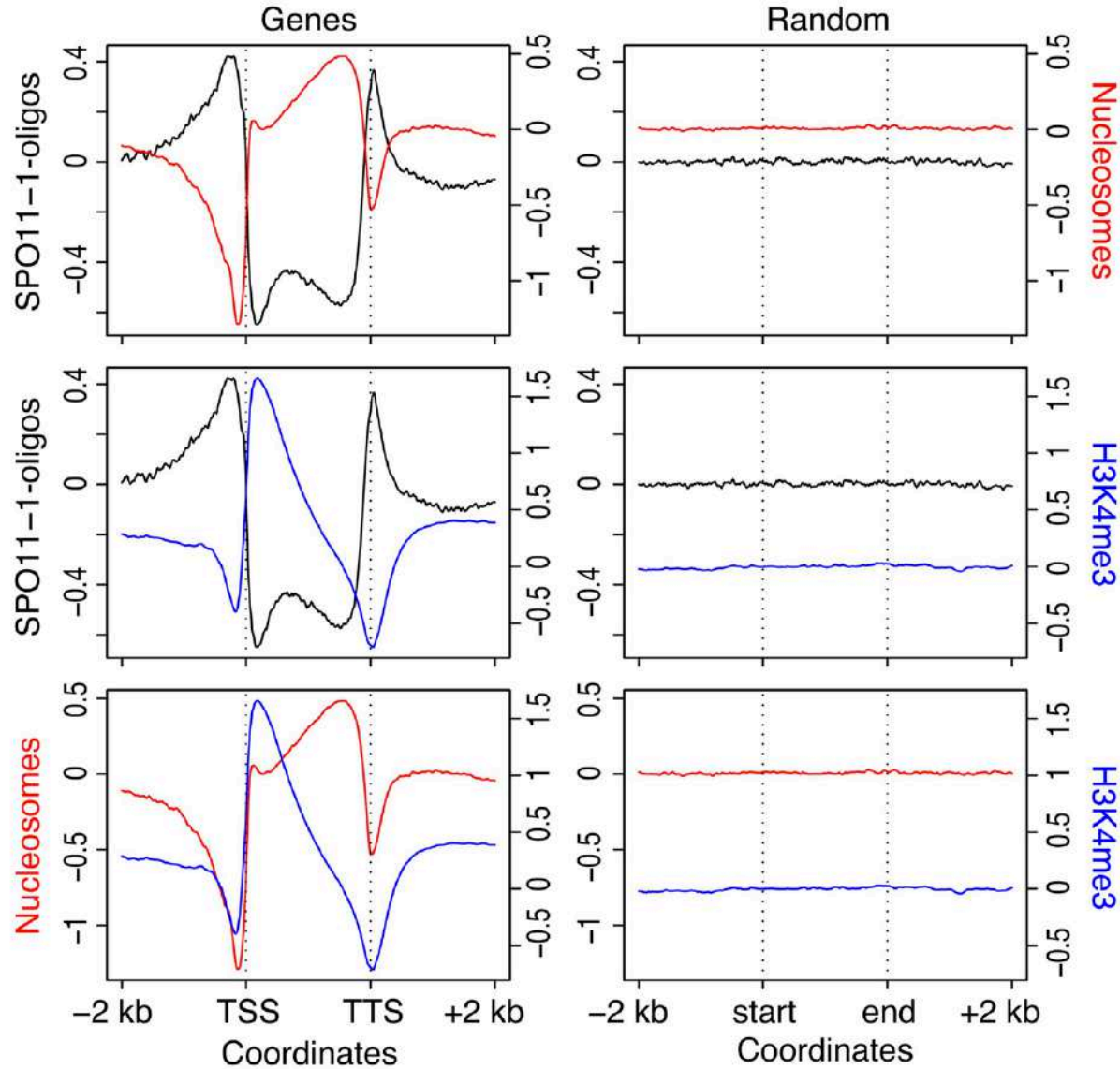
Recombination and chromatin in the wheat genome



Recombination and chromatin in the wheat genome



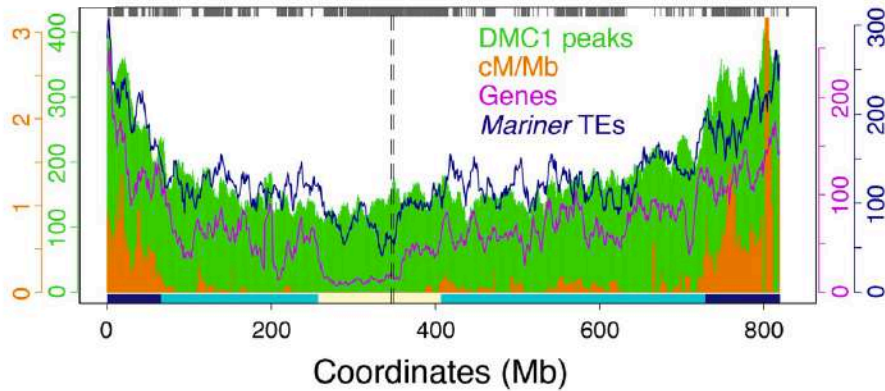
Recombination hotspots around Arabidopsis genes



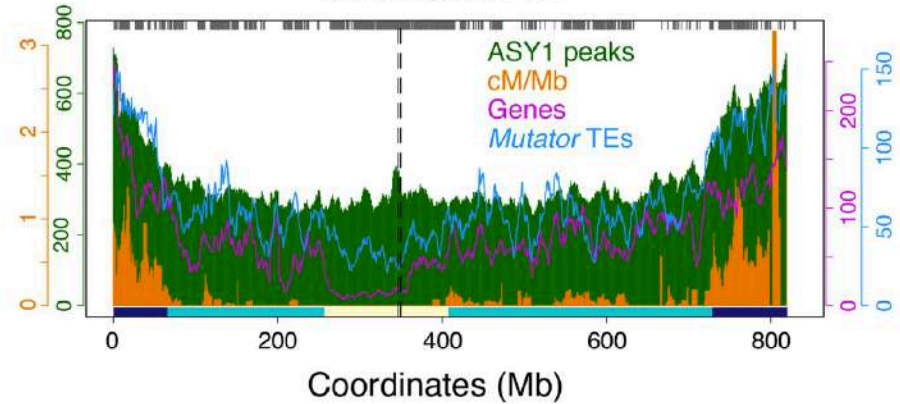
Choi et al (2018) *Genome Res*

ASY1 and DMC1 hotspots in wheat

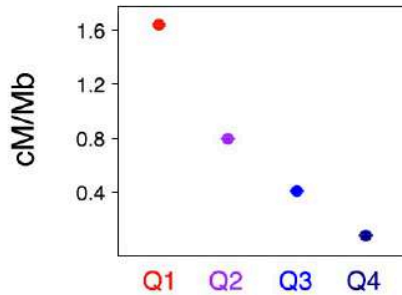
Chromosome 3B



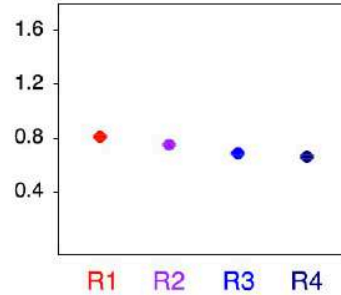
Chromosome 3B



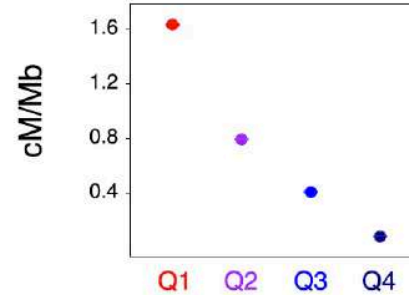
DMC1 peaks
cM/Mb



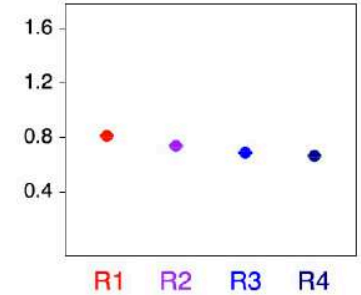
DMC1 peaks
random



ASY1 peaks
cM/Mb

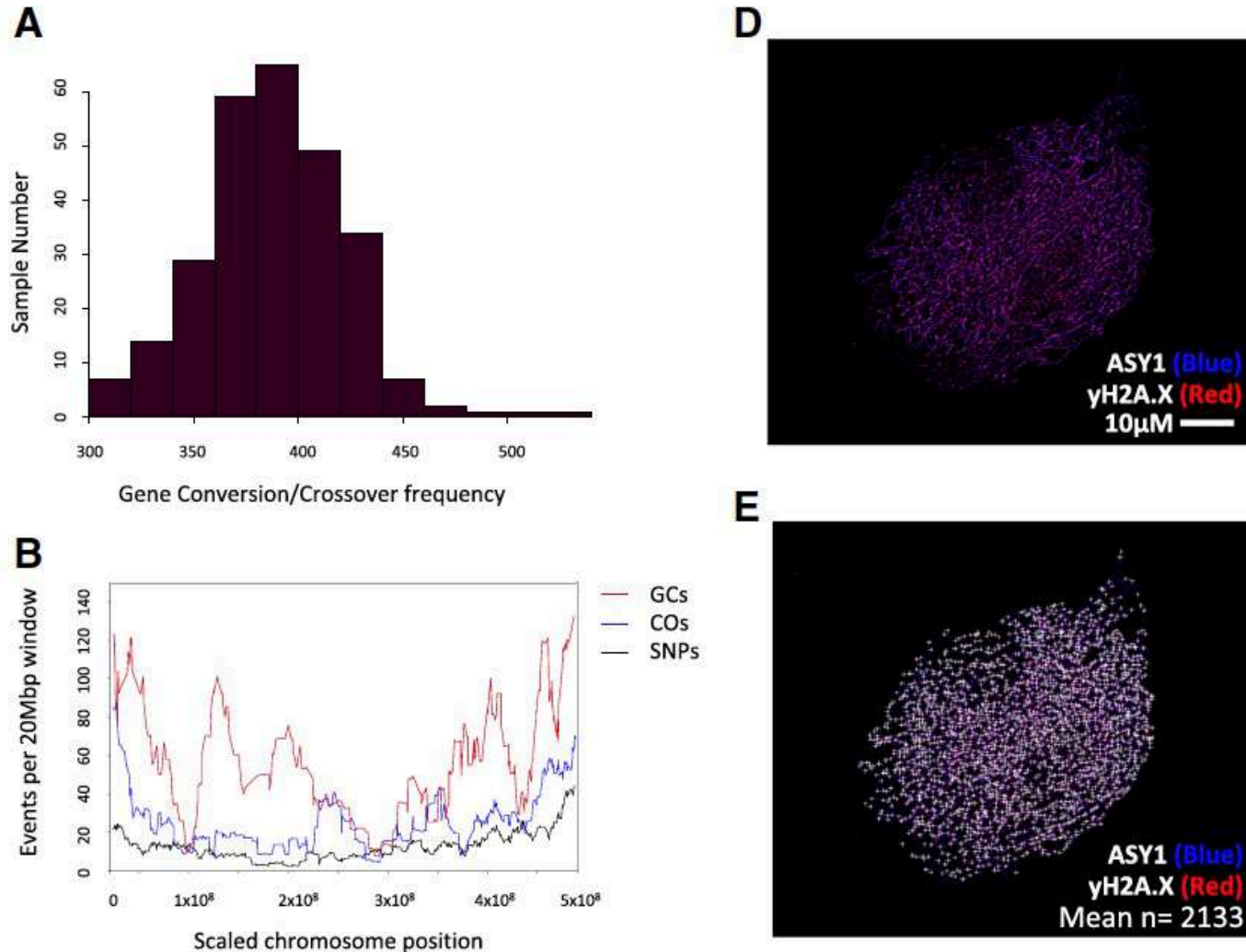


ASY1 peaks
random

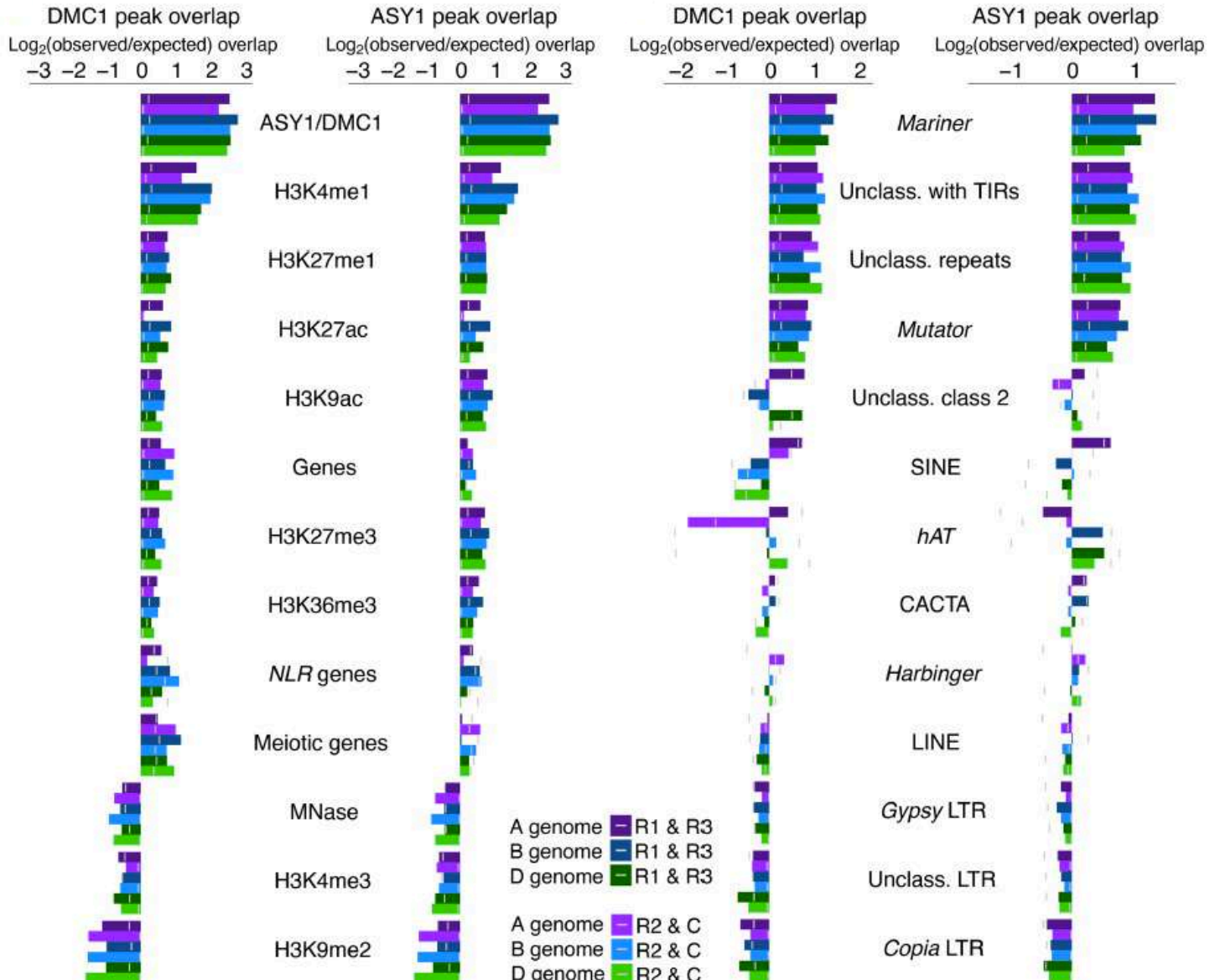


cM/Mb data from Pierre Sourdille's CS x Renan mapping experiment

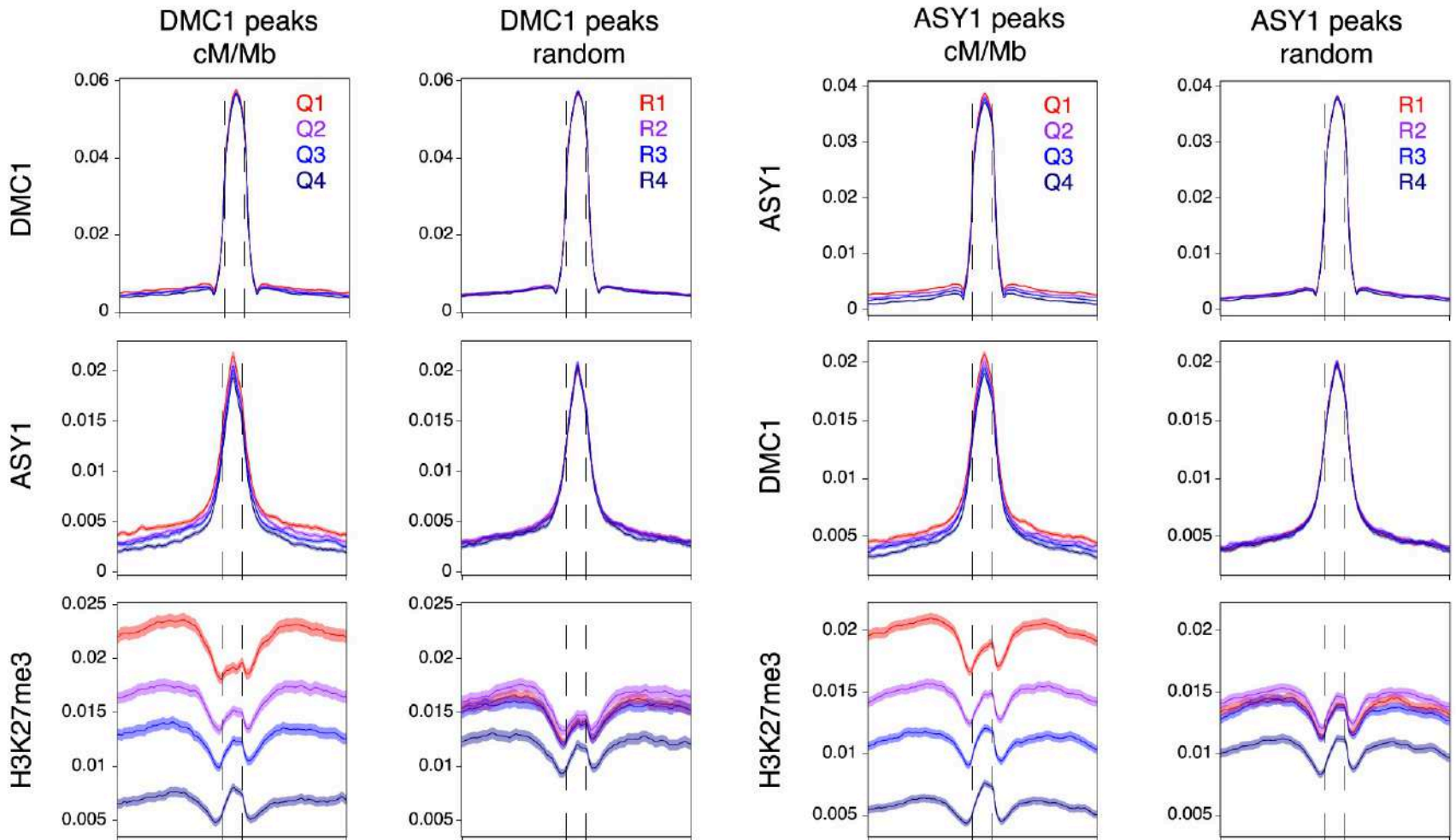
Evidence for widespread DSBs in wheat



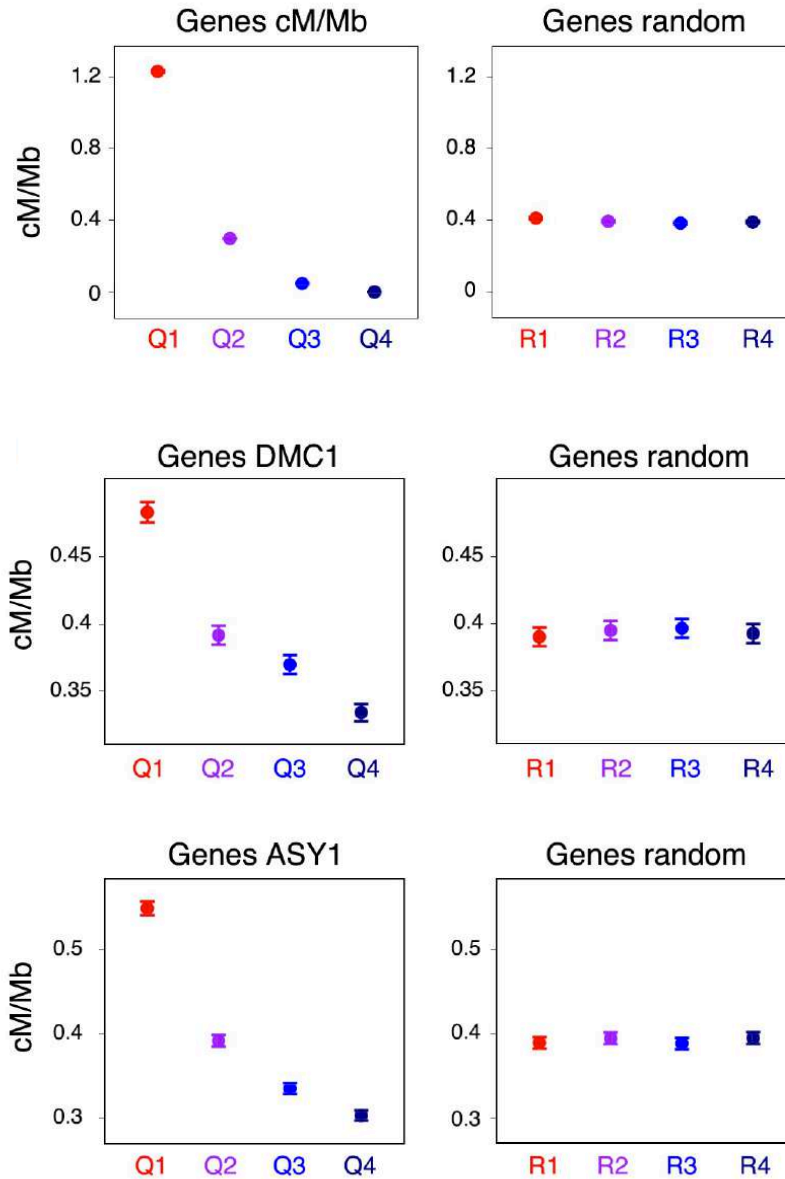
ASY1 and DMC1 hotspots in wheat



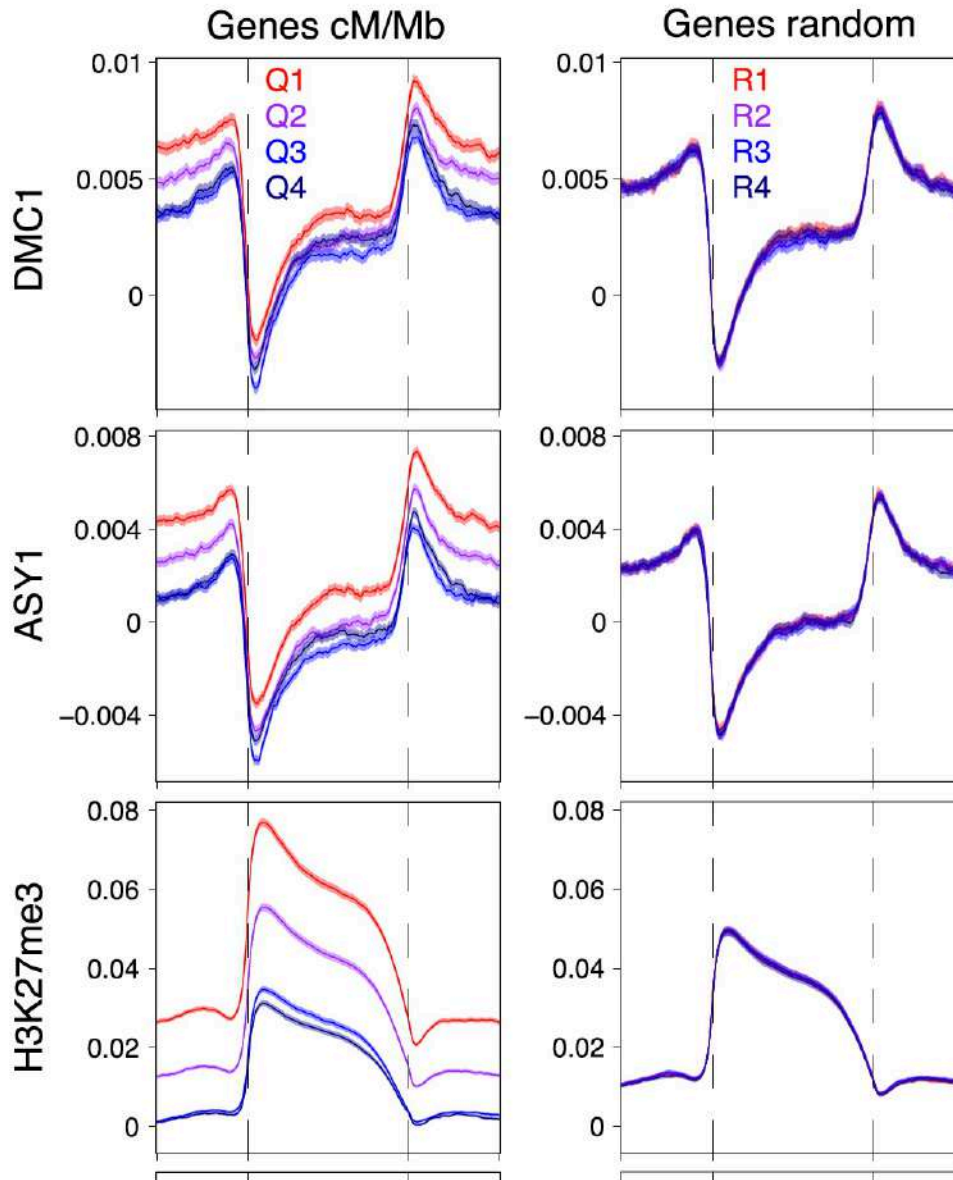
Correlation of crossovers & H3K27me3 at DMC1 and ASY1 peaks



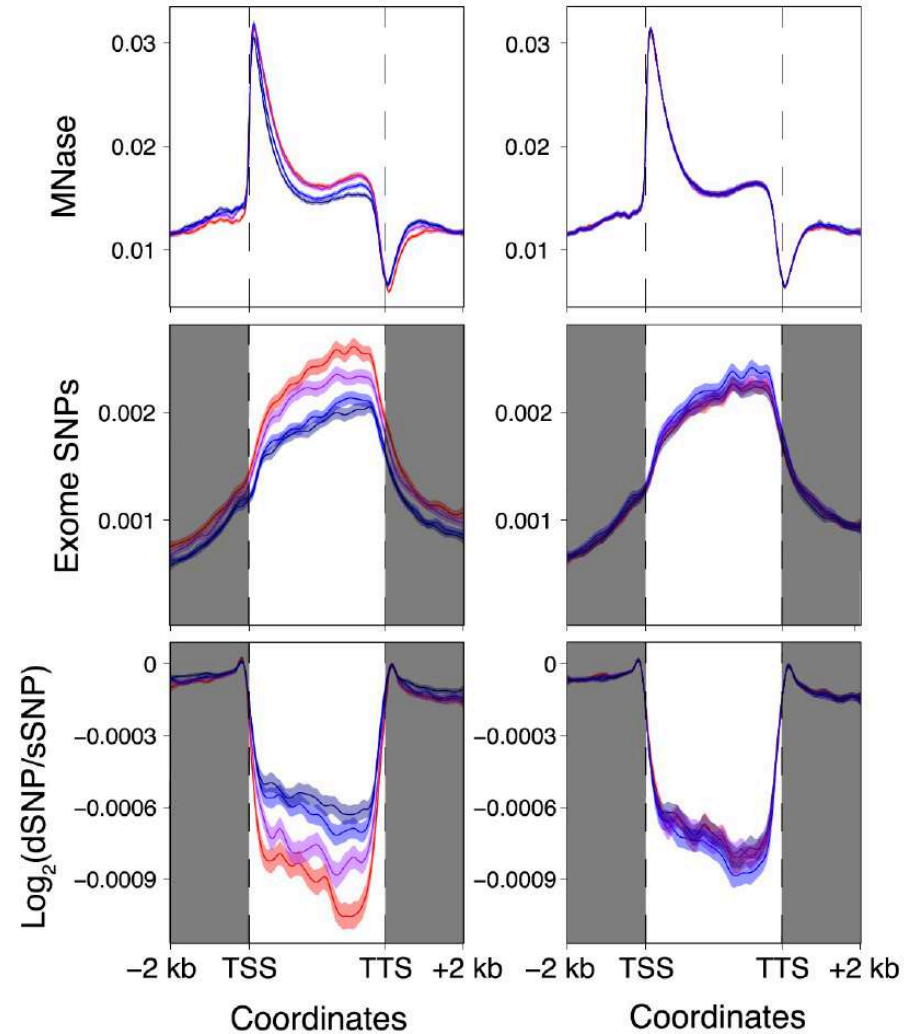
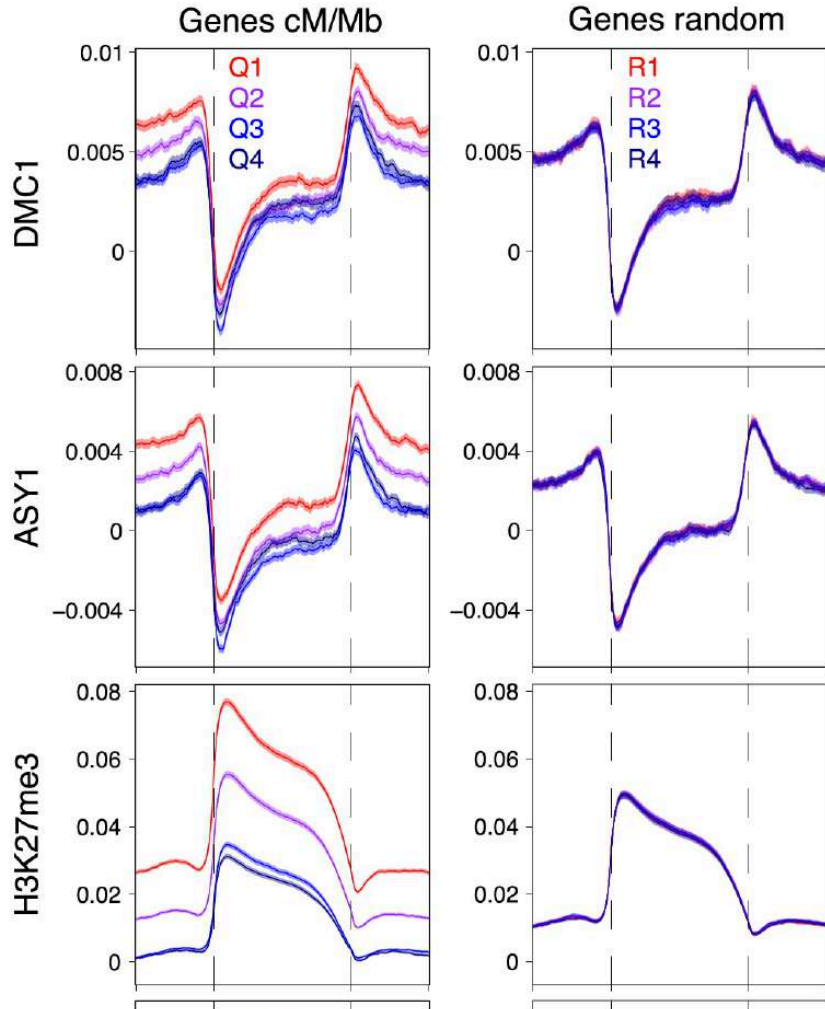
Crossovers, ASY1 and DMC1 in wheat genes



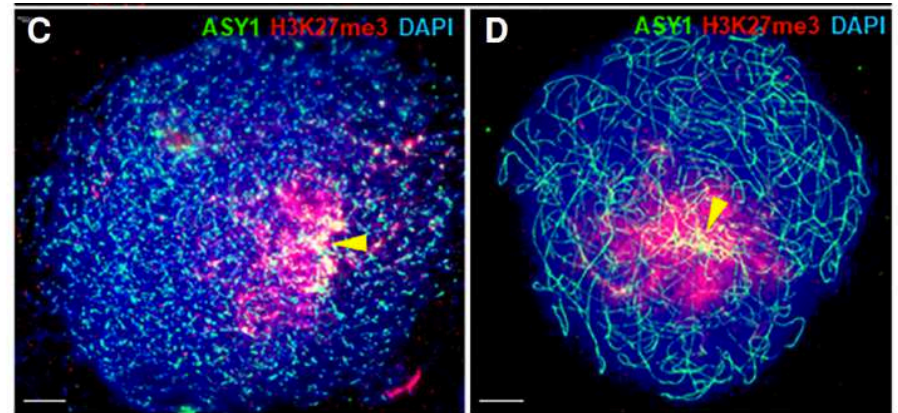
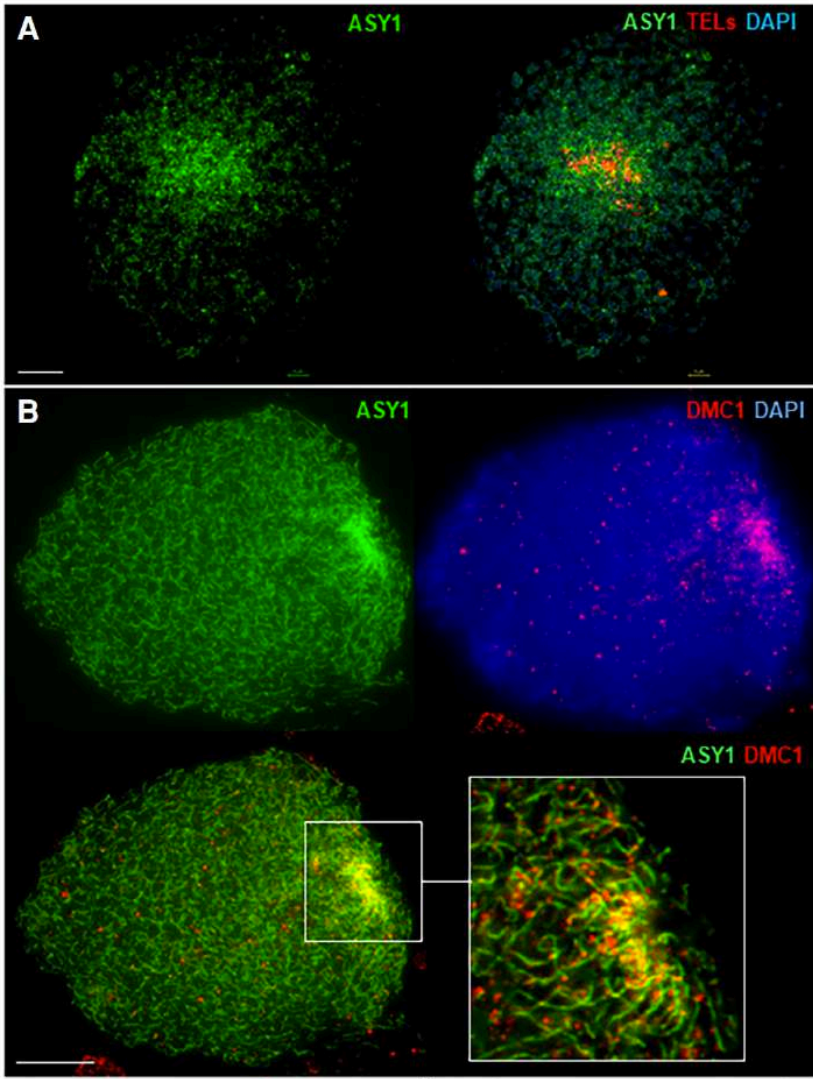
Crossovers, H3K27me3, ASY1 and DMC1 in wheat genes



Crossovers and diversity in wheat genes



Polarized H3K27me3 and recombination in bread wheat



BBSRC sLola

Kim Osman, Eugenio Sanchez-Moran and Chris Franklin

What is the Polycomb mark H3K27me3?

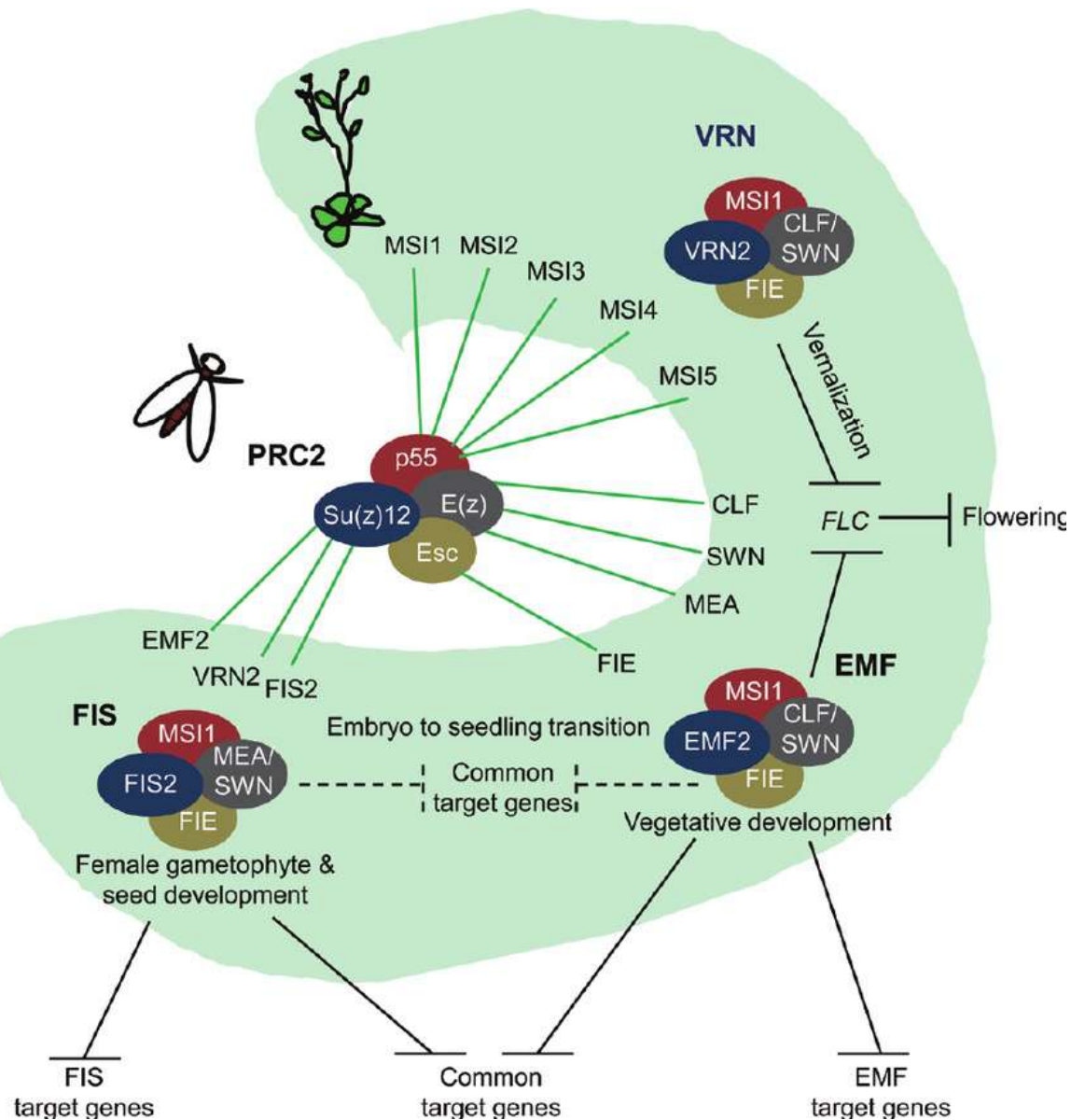
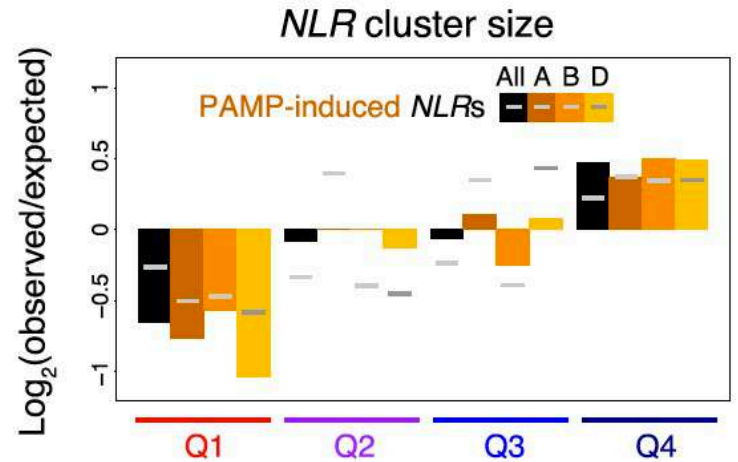
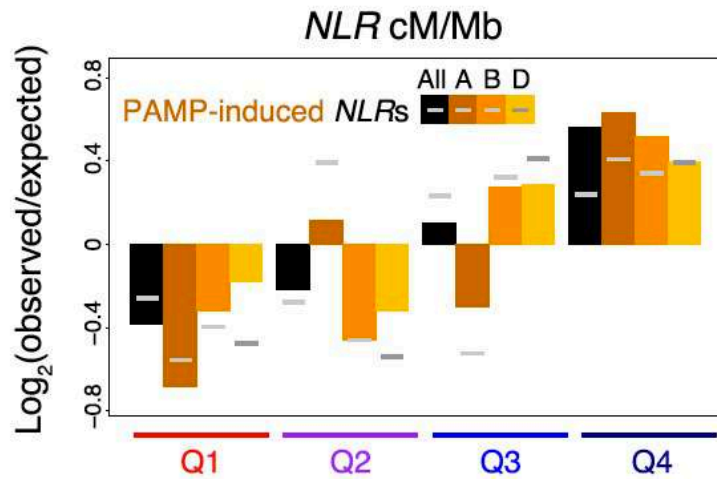
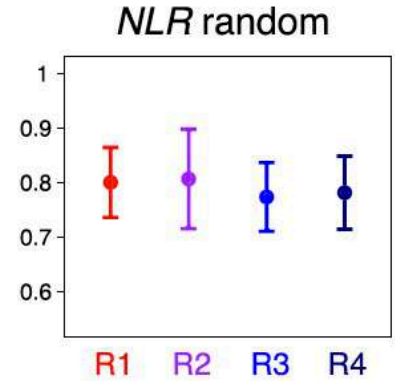
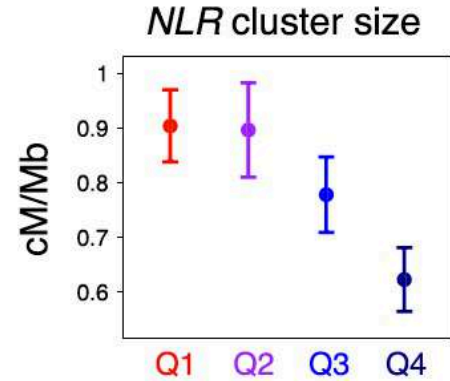
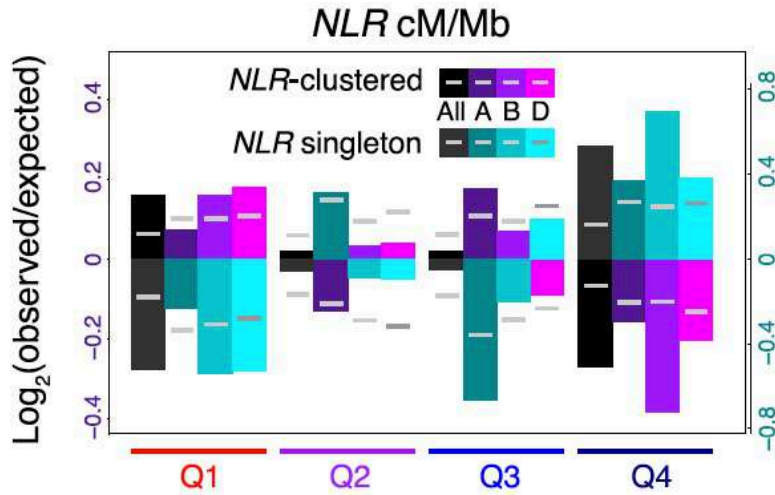


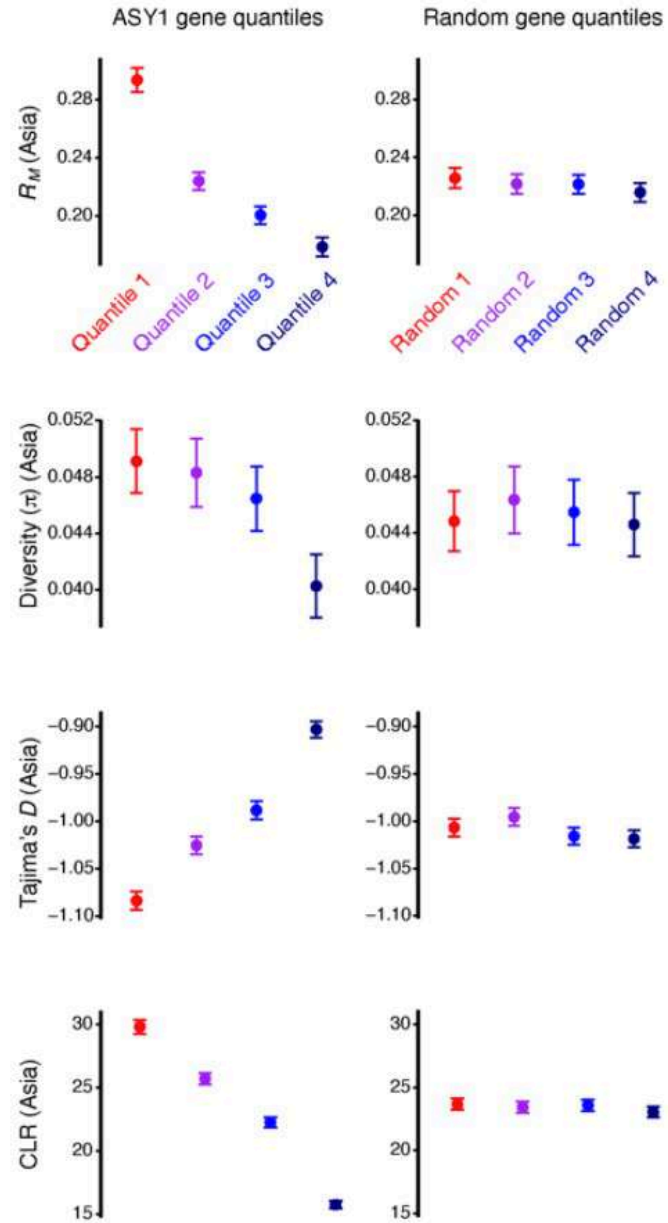
Table 4. Crossover and DNM rates. Results are presented for the sexes separately. Numbers represent the crossover rate relative to the genome average within annotated regions; values within parentheses are 95% CIs. ChromHMM categories are measured in adult ovaries. Abbreviations: Enhancers/DNase, enhancer states (EnhA1/2/AF/W1/W2/Ac) and deoxyribonuclease (DNase)-only states (DNase); Biv/Poised, bivalent and poised promoters; PRC2, polycomb-group-repressive complex 2 (ReprPC); Prom, promoter regions (PromU/D1/D2); Tx, transcribed regions (Tx5'/Tx/Tx3'/TxWk); TxEnh, enhancers within transcribed regions (TxEnh5', TxEnh3', TxEnhW, and TxReg); ZNF, enriched over zinc-finger genes and repeats (ZNF/Rpts); Het, heterochromatin.

Parameter	Paternal	Maternal
<i>Crossover recombination results</i>		
Autosomal genetic length (cM)	2602.3 (2600.4,2604.2)	4180.4 (4177.2,4183.6)
Crossovers (cM per Mb)	0.945 (0.944, 0.946)	1.518 (1.517,1.519)
Complex crossover ratio (%)	0.53 (0.50,0.56)	1.24 (1.21,1.29)
<i>Relative crossover rates in annotated regions</i>		
Pratto DSB	24.90 (24.87, 24.93)	18.93 (18.91, 18.96)
Altemose PRDM9 ± 500 bp	7.28 (7.27, 7.30)	7.12 (7.11, 7.13)
Altemose H3K4me3 ± 500 bp	3.92 (3.92, 3.93)	3.99 (3.99, 4.00)
5-Hydroxymethylation	2.82 (2.80, 2.85)	2.59 (2.58, 2.60)
THE1b regions	4.32 (4.27, 4.37)	2.53 (2.50, 2.56)
ChromHMM Biv/Poised	1.56 (1.53, 1.59)	1.69 (1.67, 1.71)
ChromHMM Enhancers/DNase	1.56 (1.55, 1.57)	1.93 (1.92, 1.94)
ChromHMM Het	0.61 (0.60, 0.63)	0.32 (0.31, 0.33)
ChromHMM Prom	1.10 (1.08, 1.12)	1.32 (1.31, 1.34)
ChromHMM PRC2	2.94 (2.92, 2.97)	2.54 (2.52, 2.55)
ChromHMM TxEnh	0.71 (0.71, 0.72)	0.63 (0.62, 0.63)
ChromHMM Tx	0.56 (0.55, 0.56)	0.46 (0.46, 0.46)
ChromHMM ZNF	0.52 (0.49, 0.55)	0.18 (0.17, 0.19)
Ovary H3K27me3	2.54 (2.52, 2.56)	2.37 (2.35, 2.38)
Ovary H3K36me3	0.60 (0.59, 0.60)	0.53 (0.52, 0.53)
Ovary H3K4me3	1.19 (1.18, 1.21)	1.43 (1.42, 1.44)
Ovary H4K20me1	0.64 (0.62, 0.65)	0.55 (0.54, 0.56)
Ovary H3K27ac	1.37 (1.36, 1.38)	1.64 (1.63, 1.65)
Ovary H3K4me1	1.41 (1.40, 1.42)	1.68 (1.67, 1.69)
<i>Sex-specific DNM rates near crossovers</i>		
Genome-wide rate (10 ⁻⁹)	9.59 (9.48, 9.70)	2.59 (2.54, 2.64)
Enrichment within 0 to 1 kb	41.5 (33.2, 52.0)	58.4 (44.0, 77.4)
Enrichment within 1 to 3 kb	6.91 (4.76, 10.1)	11.9 (7.42, 19.2)
Enrichment within 3 to 40 kb	1.05 (0.82, 1.35)	2.21 (1.60, 3.06)
Enrichment within 3 to 40 kb (only for complex crossovers)	—	49.7 (27.5, 90.0)

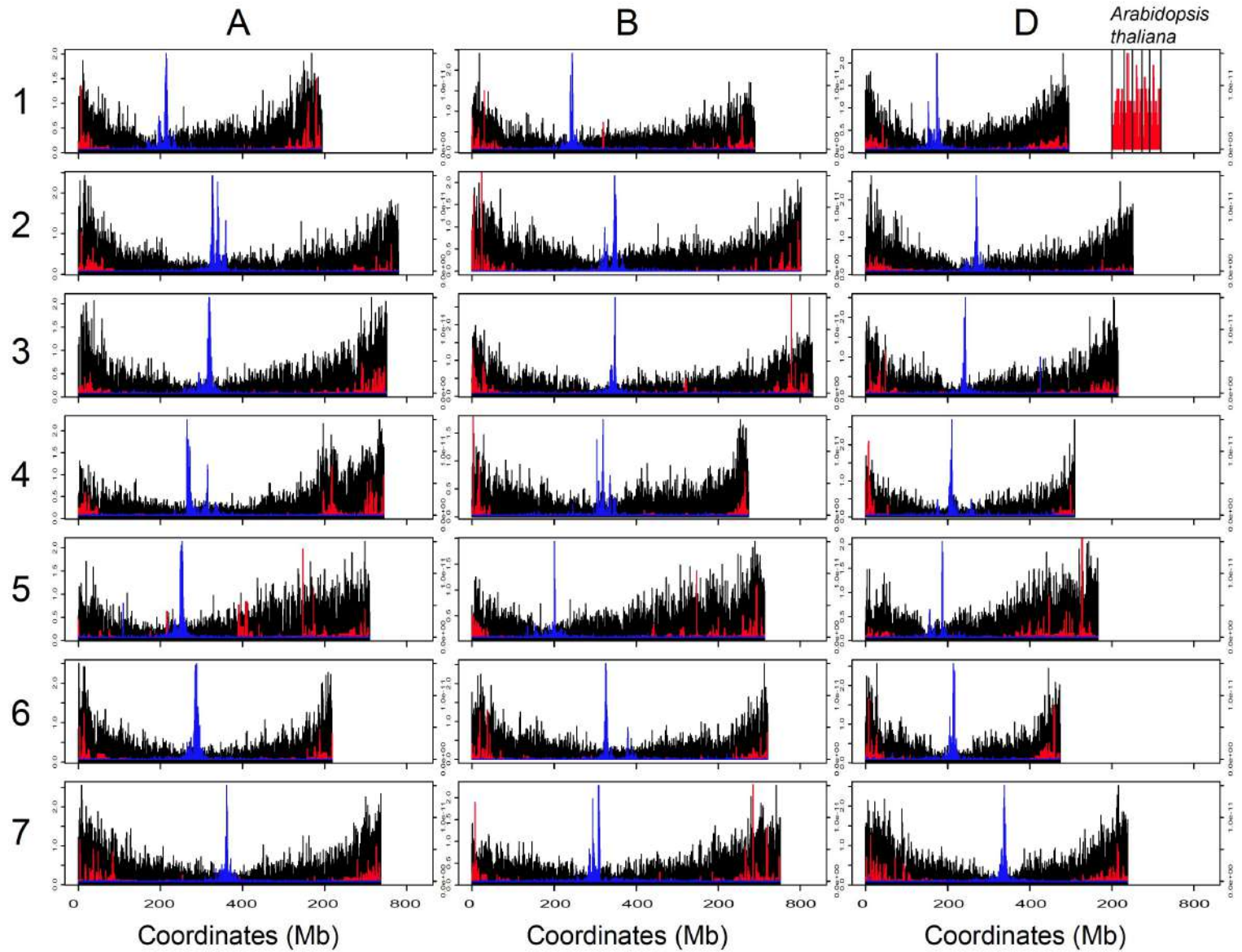
Crossovers and wheat *NLR* genes



Crossovers, diversity and selection



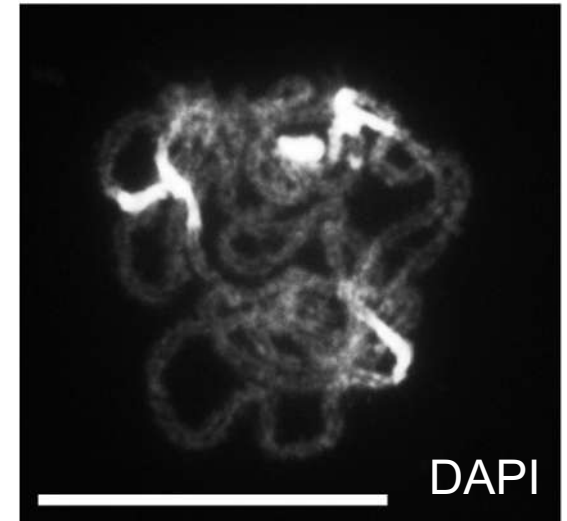
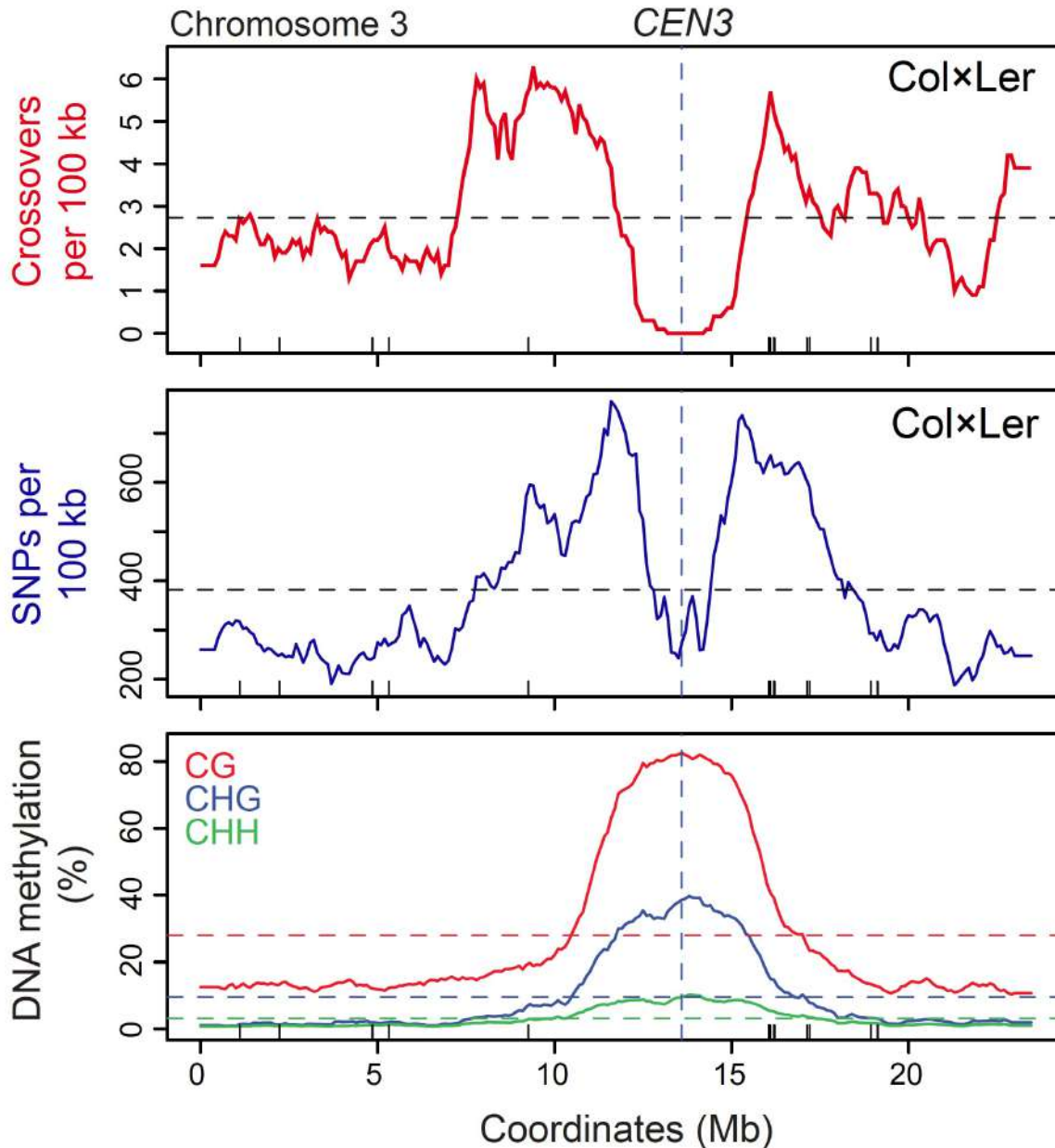
Recombination gradients in the wheat genome



■ H3K4me3:H3K9me2
■ Gypsy LTR Cereba
■ cM/Mb

BBSRC sLola

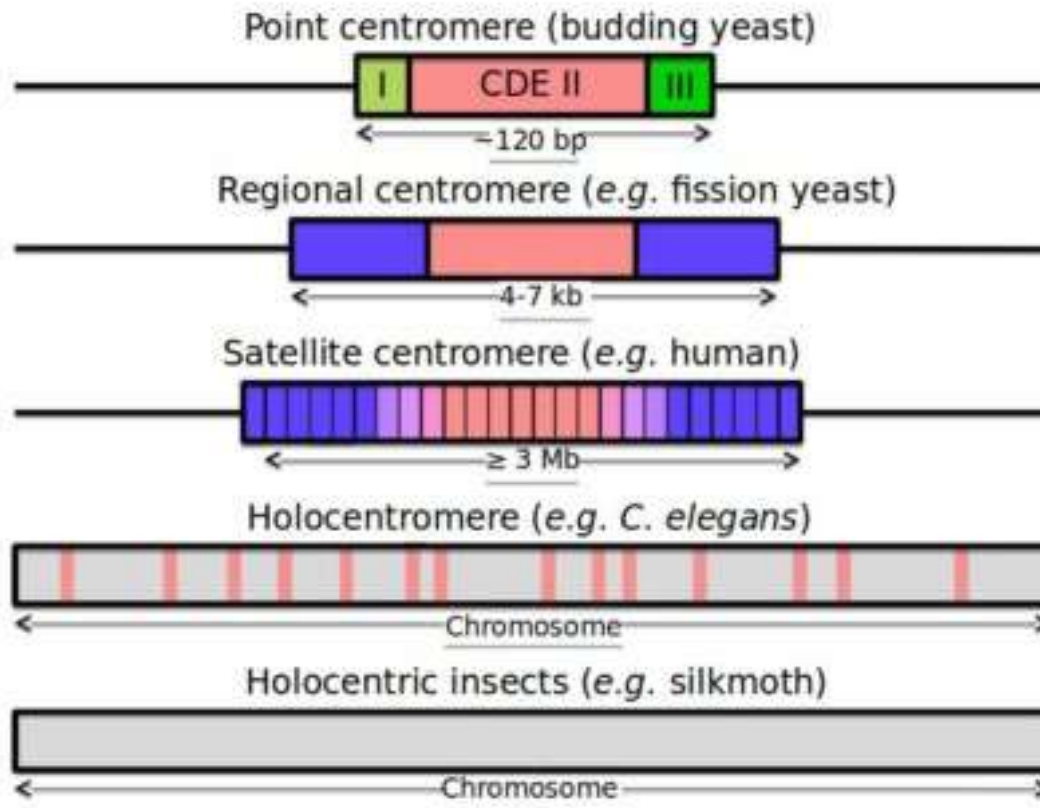
Arabidopsis crossover and polymorphism landscapes



Christophe Lambing

The Centromere Paradox: Stable Inheritance with Rapidly Evolving DNA

Steven Henikoff,* Kami Ahmad, Harmit S. Malik



← Arabidopsis

Long read DNA sequencing

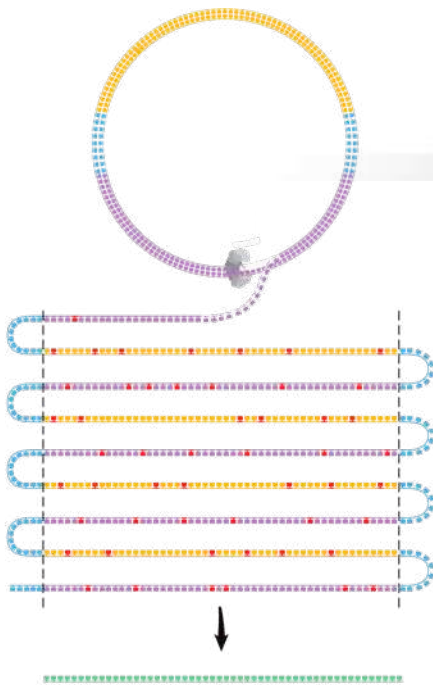


Oxford Nanopore
Technologies

>100 kb

95-99% accuracy

Methyl-detection



HiFi READ

>99.9% accuracy

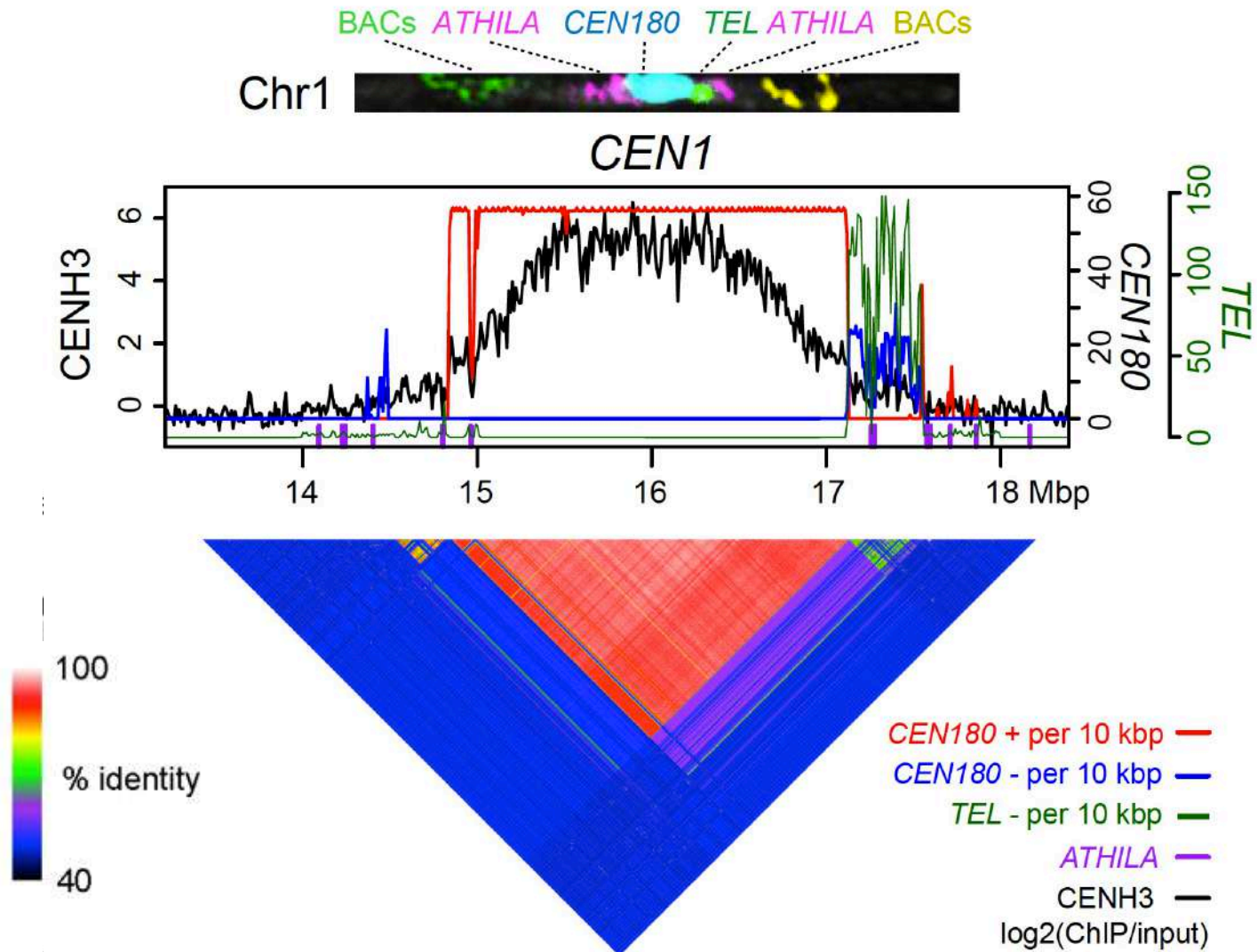
Pacific Biosystems

HiFi CCS

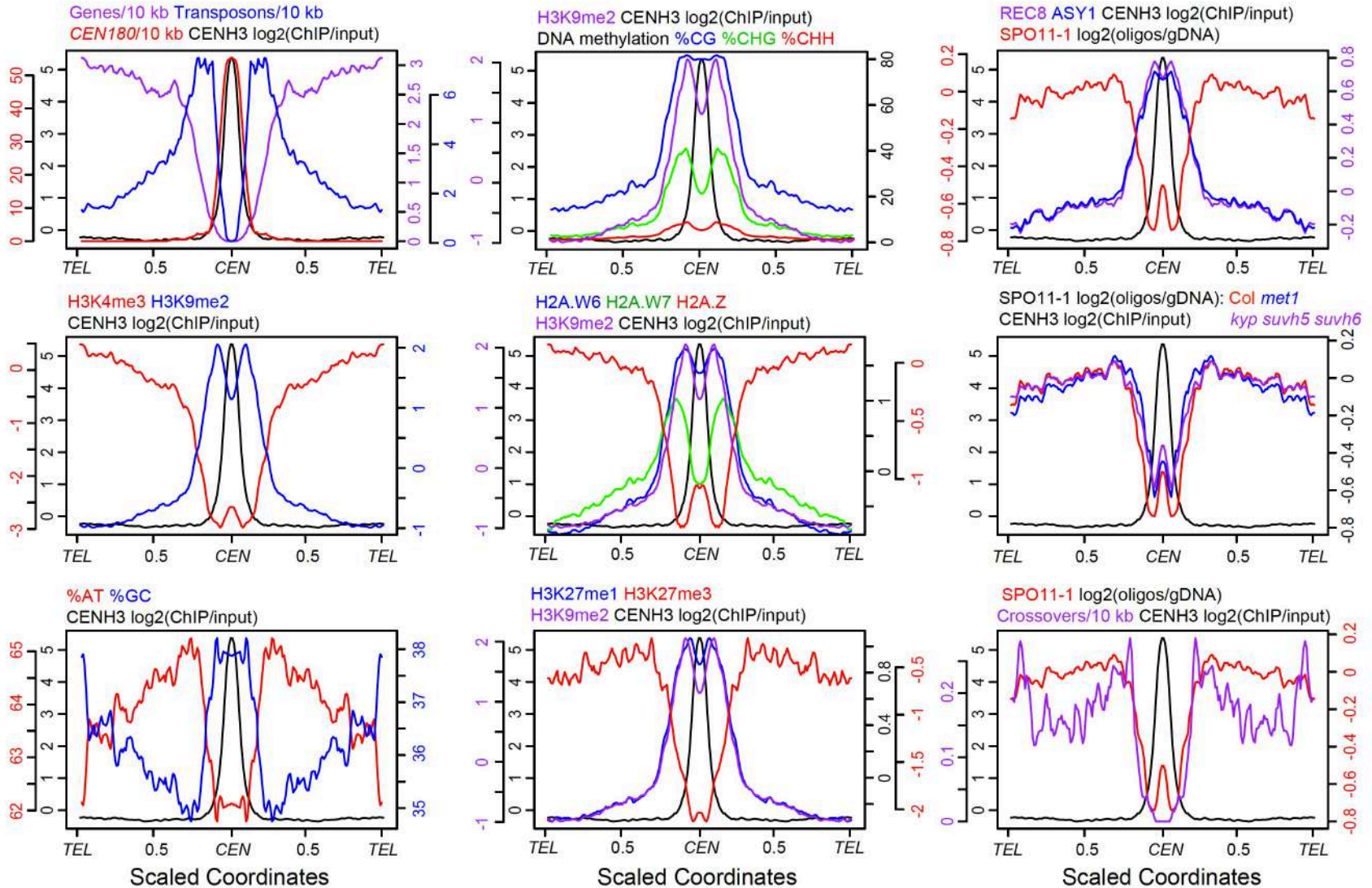
~15 kbp

>99% accuracy

Centromeres are massive tandem arrays of *CEN180* repeats that support CENH3

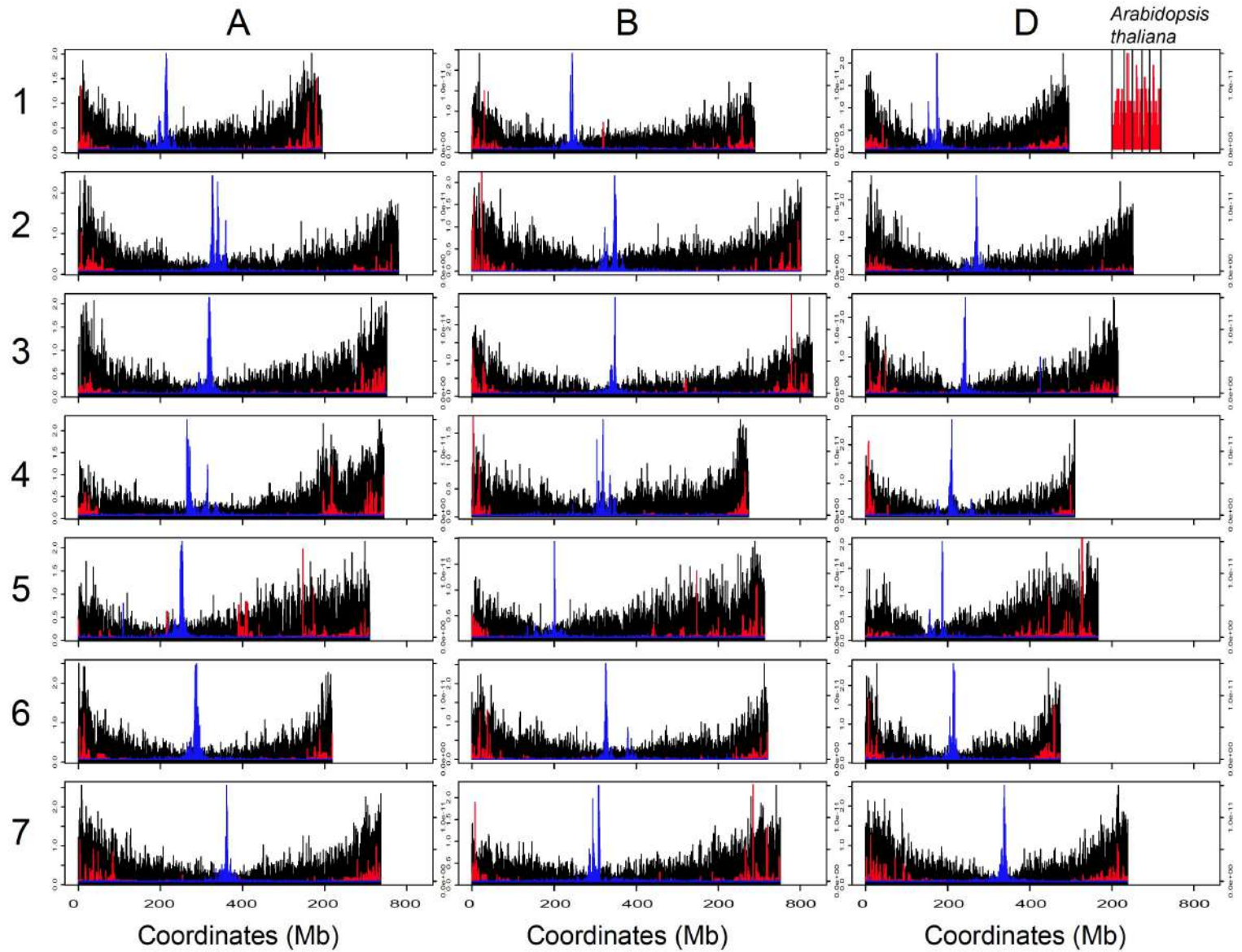


Genetic and epigenetic landscape of the centromeres



Centromeric chromatin is distinguished from pericentromeric heterochromatin

Recombination gradients in the wheat genome



■ H3K4me3:H3K9me2
■ Gypsy LTR Cereba
■ cM/Mb

BBSRC sLola

Questions:

1. The wheat genome is highly polarized in terms of recombination (crossover), DMC1 and ASY1
2. DMC1 and ASY1 hotspots are widespread
3. The polycomb mark H3K27me3 distinguishes regions of high crossover activity
4. Recombination correlates with gene diversity, signatures of selection and annotation
5. Arabidopsis may not be an adequate model for the wheat genome!

Acknowledgements



Lab members

Stephanie Topp

Andy Tock

Robin Burns

Daniel Holland

Piotr Wlodzimierz

Matthew Naish

Nicola Gorringe

Collaborators

James Higgins (Leicester)

Chris Franklin (Birmingham)

Eugenio Sanchez-Moran (Birmingham)

Kim Osman (Birmingham)

Keith Edwards (Bristol)

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Piotr Ziolkowski (AMU Poznan)

Charlie Underwood (KeyGene)

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ERC

BBSRC

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KWS

HFSP