

IWGSC Workshop and Training Session
7-8 April, 2011

Wheat Bioinformatics activities at the CCG

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UNIVERSITY
PERTH, WESTERN AUSTRALIA

CENTRE FOR
COMPARATIVE GENOMICS

Western Australia

Outline

- Contribution to the IWGSC
 - Two specific regions of chromosome 3B
 - Chromosome 7A
- Analytical environment for analysis
- Informatics issues

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Chromosome 3B

- **The ctg506 region** Selected for detailed analysis
 - Several cell wall invertase (IVR1) genes are located in this region.
 - These genes are often important in maintaining pollen viability during early development.
- **The ctg344 region** Selected for detailed analysis
 - Carries the gwm533 marker that is widely used to track disease resistance (Sr2) located on 3B
 - Potentially a region important in several disease resistances

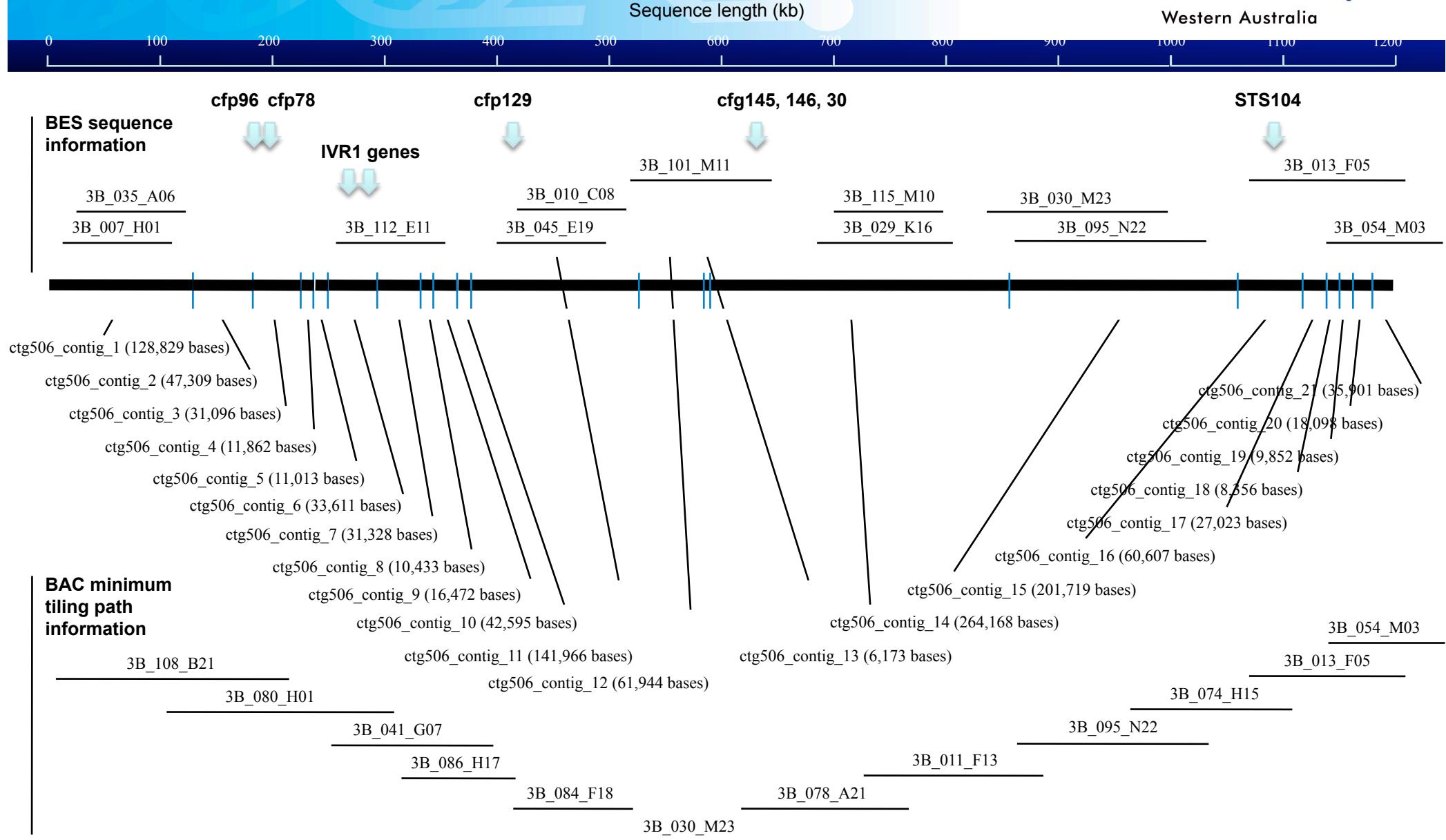
Core Ideas for Assembly

- We know wheat is “difficult” to assemble
 - mis-assemblies are and will be common
- If we see the same sequence assembled using different assemblers and data
 - this is more likely to be correct
- Points of divergence between different assemblers need to be analysed in detail
- Information from databases such as TREP need to be considered in compiling the detailed genome sequence.
- Genome sequence information needs to relate to a high quality molecular genetic map with traits included

Assembly approach

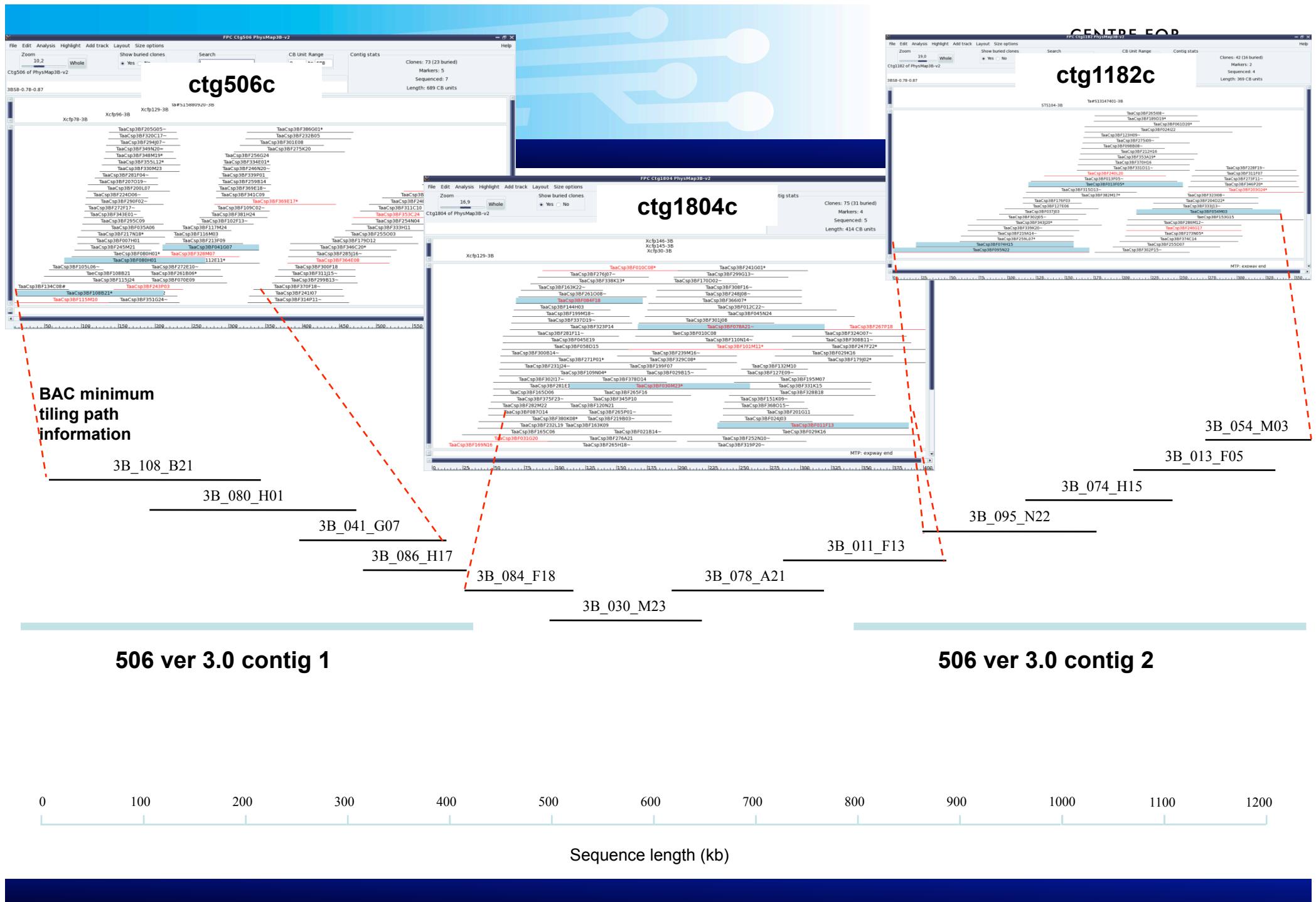
- Each BAC (Illumina short reads 70bp, paired-end, Sanger) assembled separately using Velvet
 - using multiple parameter combinations
- Different assemblies of the same BAC compared (mummer, freckle)
 - Potential mis-assemblies identified (eg: assemblies that disagree at certain points)
- Using the fingerprint assembly as a guide, identifying sequence present in overlapping BACs – take contigs that agree between different BACs to be “confident”. Start with large contigs and work down to shorter contigs.
- Look at paired-end alignments (BWA + genomeview, Hawkeye) and identify potential mis-assemblies
- Use paired-end information (illumina short reads 500bp insert + Sanger 4kb insert) to try to extend contigs (in general, this is difficult)
- Use LTR information from eg: TREP to order contigs

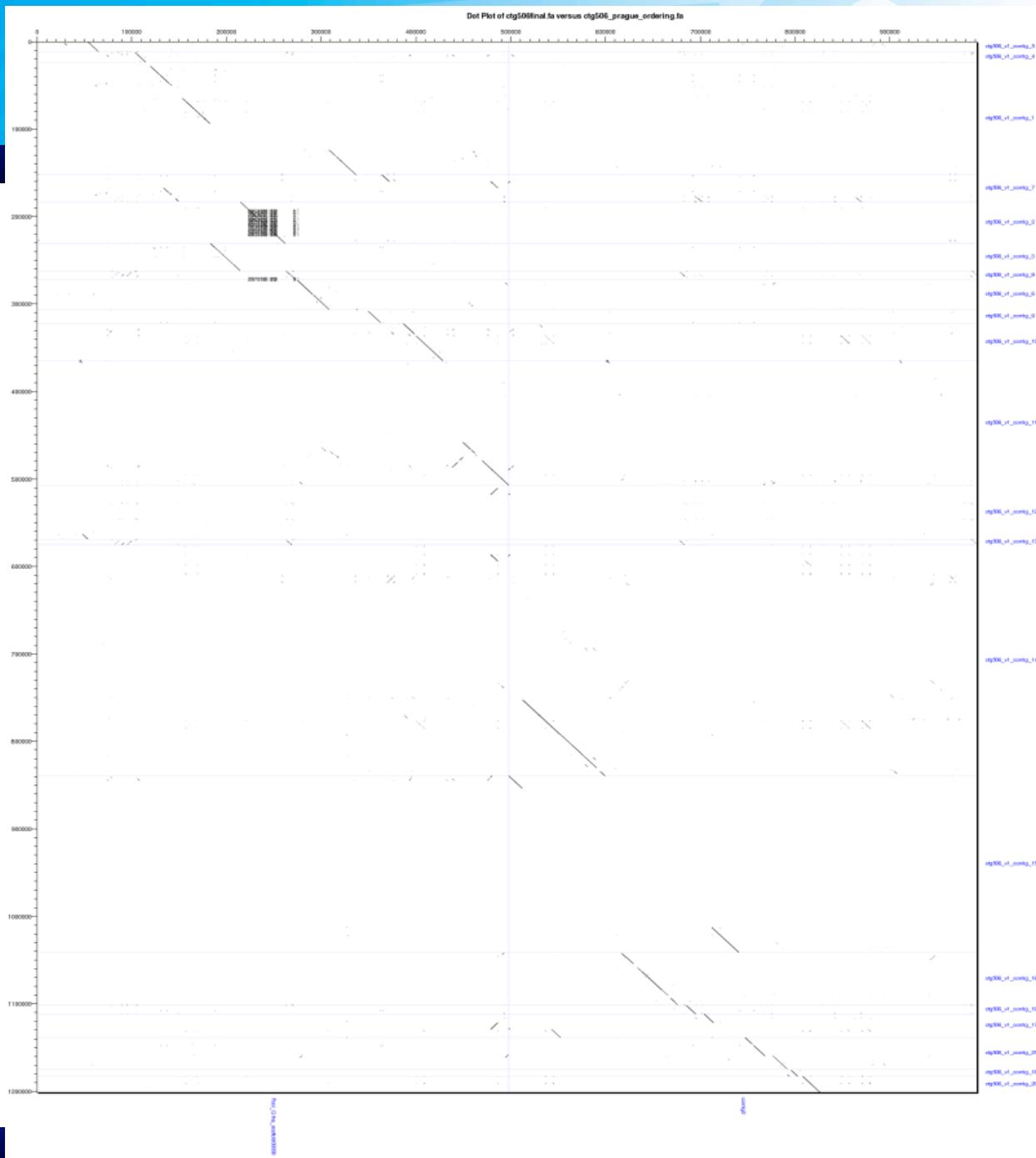
Original ctg506



INRA 454/8kb of Ctg506

- CCG-illumina/Sanger sequencing compared to INRA 454/8kb mate pairs sequencing:
- The INRA scaffolds for ctg506
 - Closed 6 small gaps of Ns in CCG assembly
 - Improved the ordering of contigs in the CCG sequencing
 - Highlights miss-assemblies (at least 6)
- The CCG sequencing
 - Orient large INRA scaffolds
 - Closed 30/83 gaps in INRA sequencing assembly
 - Most of these gaps were in the introns of regions annotated as coding sequences.

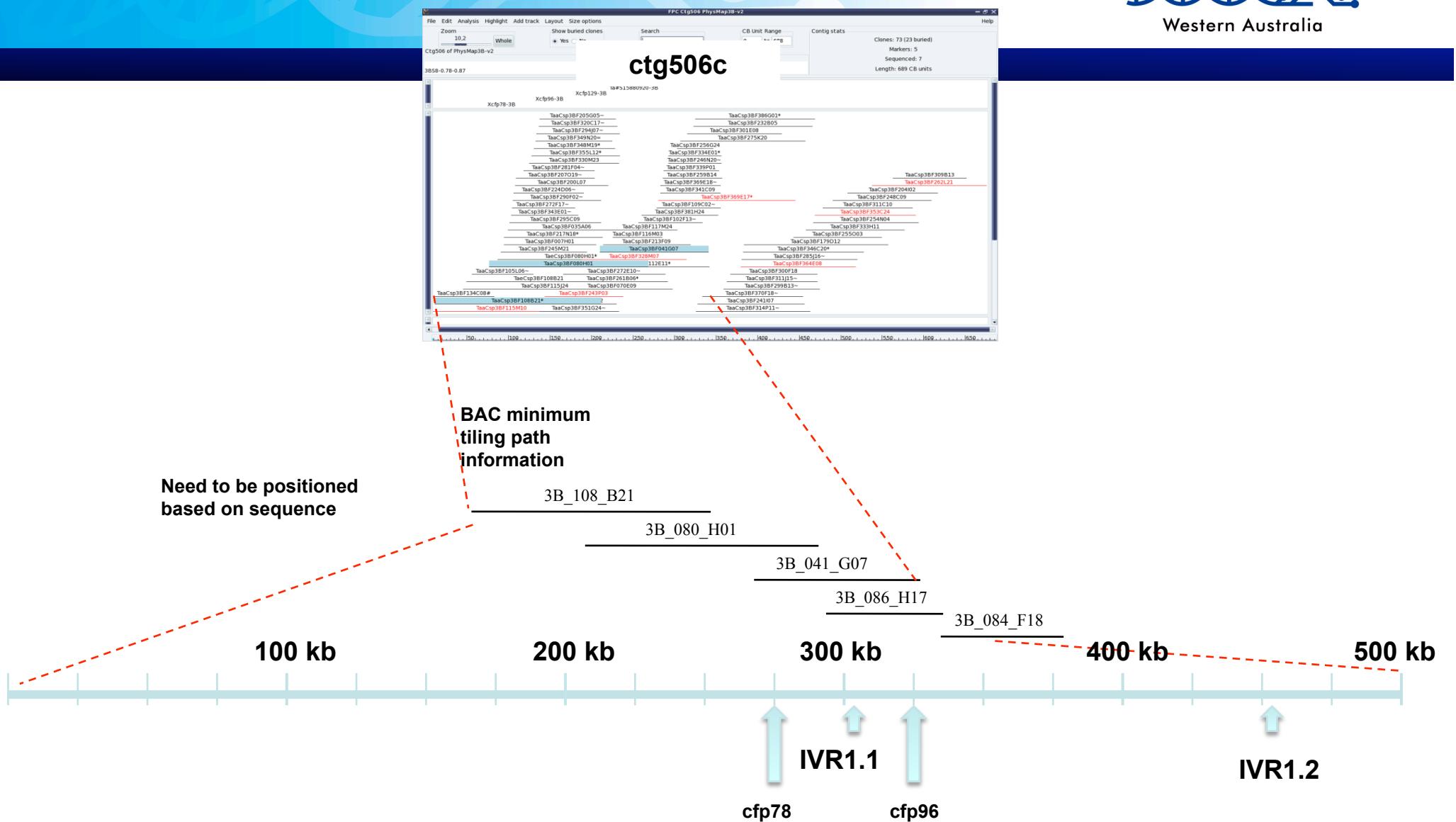




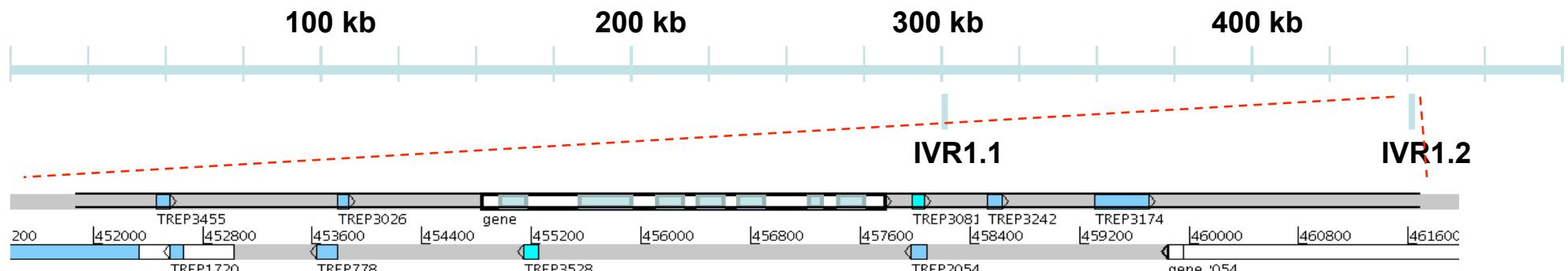
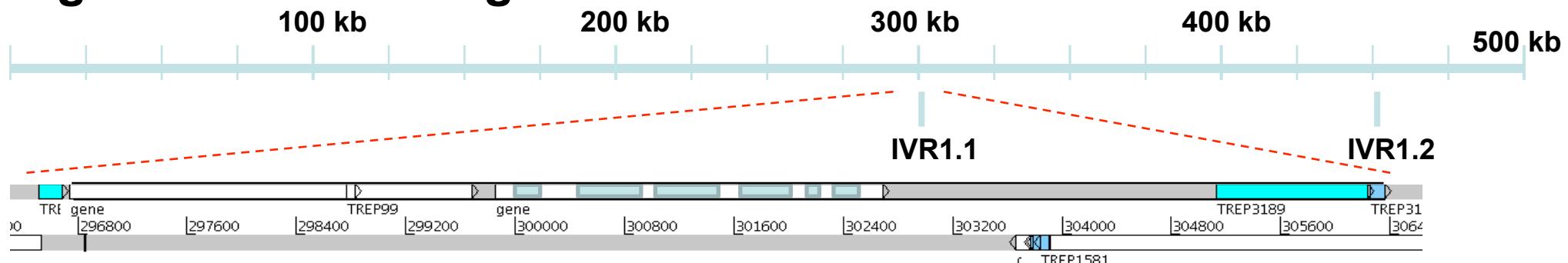
Alignment of CCG short read +
Sanger assembly (y-axis) against
version 3 (combined CCG + INRA)
(x-axis)

The CCG contigs have been re-
arranged in an attempt to fit the order
revealed by INRA scaffolds

Detailed analysis of IVR gene region



ctg506c ver3.0 contig1



Summary – Chromosome 3B

- Have a good appreciation of sequence assembly issues for wheat genome
- Comparative genomics of a repetitive protein kinase locus in ctg344 and biological studies of cell wall invertase genes on ctg506
- Waiting for ctg1804c and ctg344 scaffolds from INRA

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Chromosome 7A

Overall aims

- Detailed analysis of QTL regions that are important to Australian agriculture
- Trait information linked directly to genome sequence
- ISBP identification of new markers for wheat breeding and selection for traits of interest

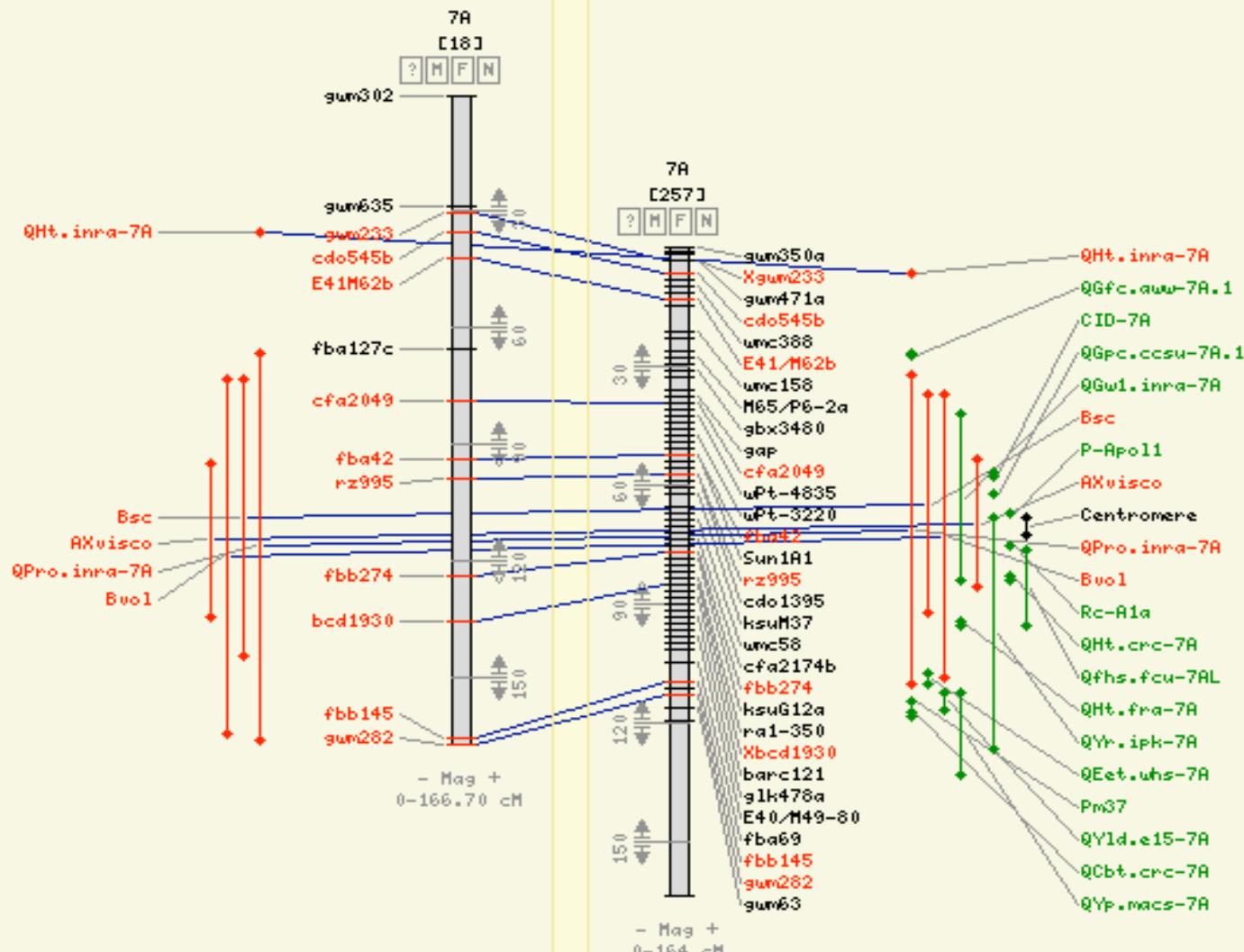
Tasks

- BAC-based physical map assembly of 7AS and 7AL
- BAC end sequencing
- Survey sequencing of 7A
- In-depth sequencing of QTL regions
- Anchor to genetic map

7A genome
sequence

Comparative
Wheat
Renan*Recital Groos 07

Reference
Wheat
7A Consensus May 09

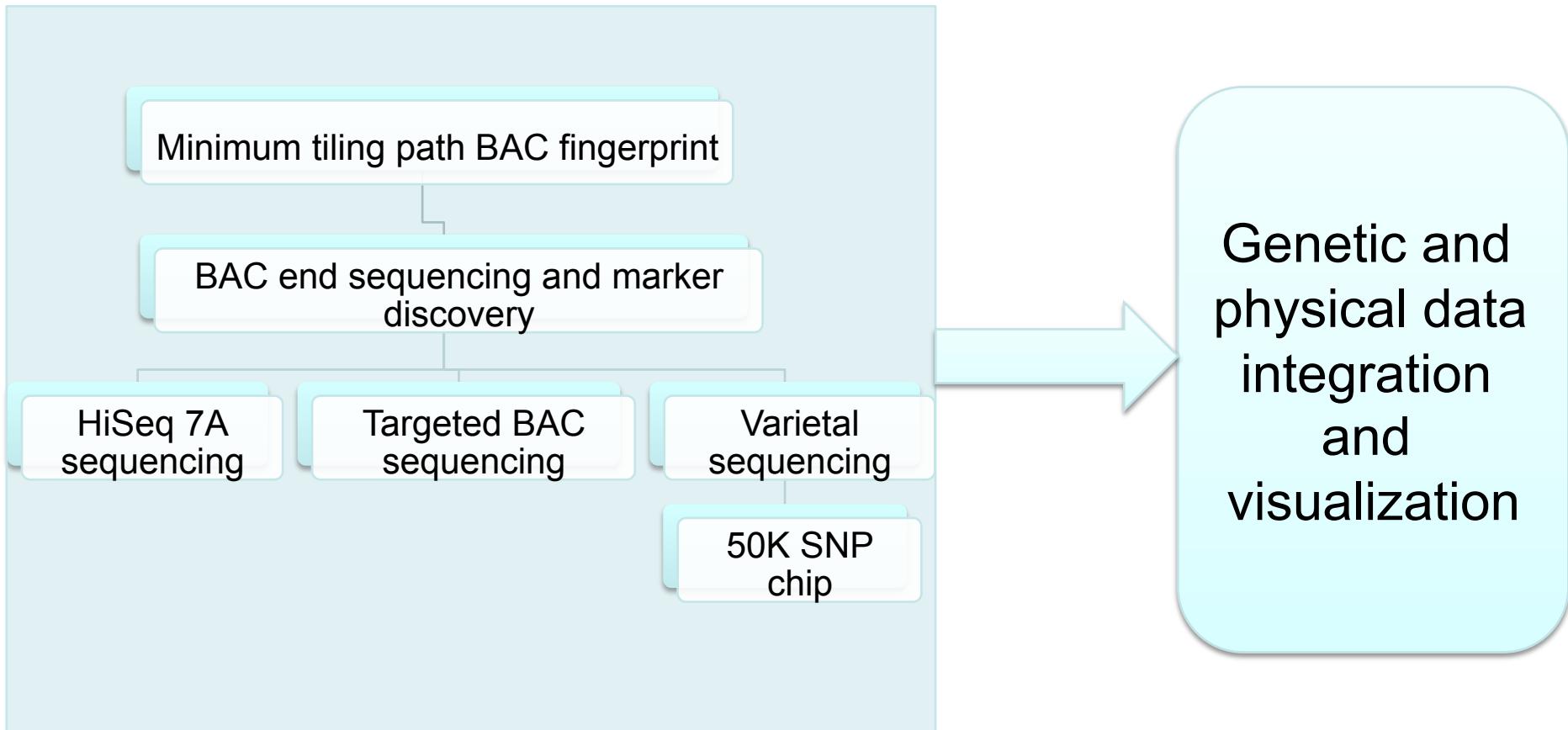


- Cmap (at CCG) compiles published QTL/trait data onto composite map.
- The QTL/trait data includes the information from the Wheat Gene Catalogue (McIntosh et al).
- The composite map is built from sections of published maps that share common markers to allow their integration into a master map. Trait data is incorporated based on flanking molecular markers. The molecular markers allow projection on to the genomic DNA sequence

7A timeline

- **2010** Funding finalised (GRDC and BioPlatforms Australia)
 - 7A physical map, support to the IWGSC, survey sequencing, QTL region sequencing
- **March 2011** 7AS BAC library (58,000 BAC clones) produced by Dolezel lab
 - UC Davis for DNA fingerprinting (Mingcheng Luo)
- **May 2011** LTC/FPC 7AS fingerprint assembly
- **June 2011** compile BAC contigs and define minimum tiling paths (MTP) for physical map
- **May/June 2011** 7AL library from Dolezel lab (underway) shipped to UC Davis for DNA fingerprinting
- **June** commence BAC end sequencing (BACs from MTPs)
- **2011** Targeted BAC sequencing
- **2012** anchoring of BAC contigs to genetic maps

CCG: Wheat Bioinformatics 7A (Data integration)



Outline

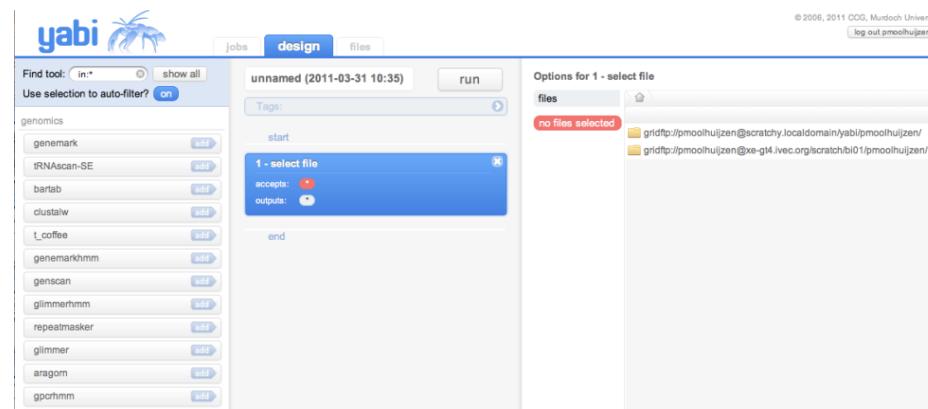
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CCG: Wheat Bioinformatics activities 3B

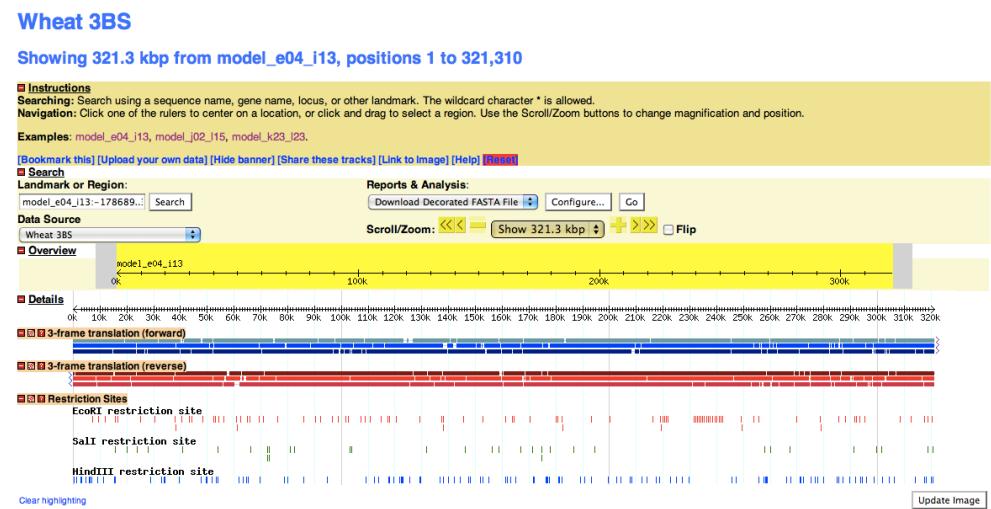
HTP Assembly (Ctg344 and Ctg506)

Annotation (HTP pipeline)

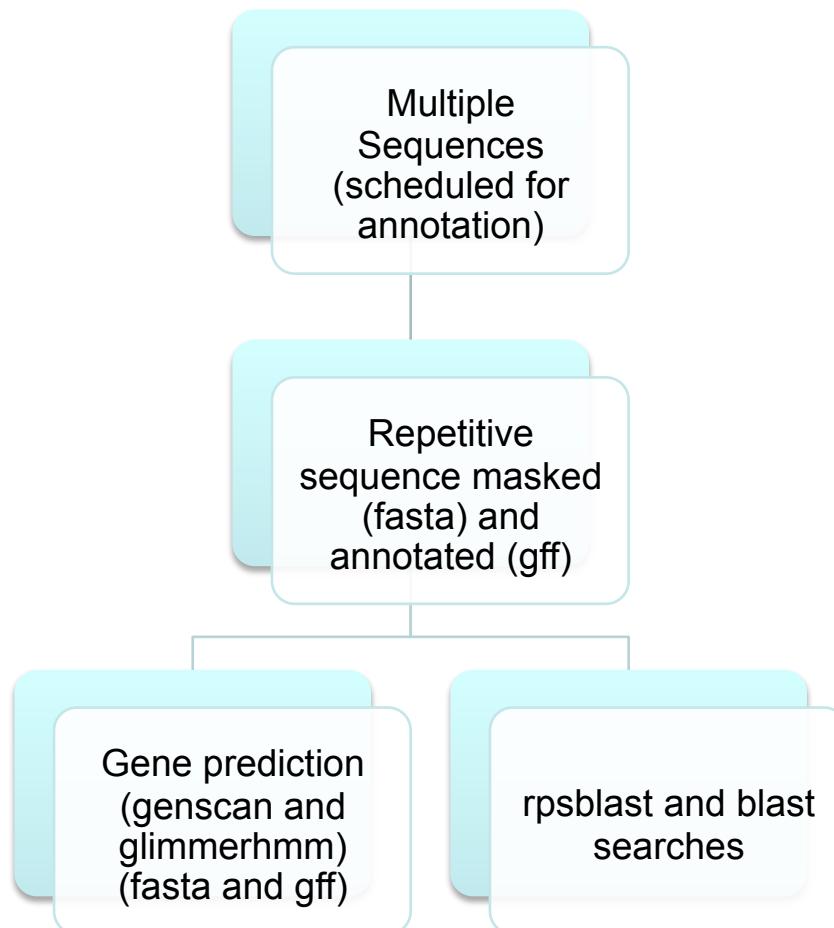
Visualization (Gbrowse)



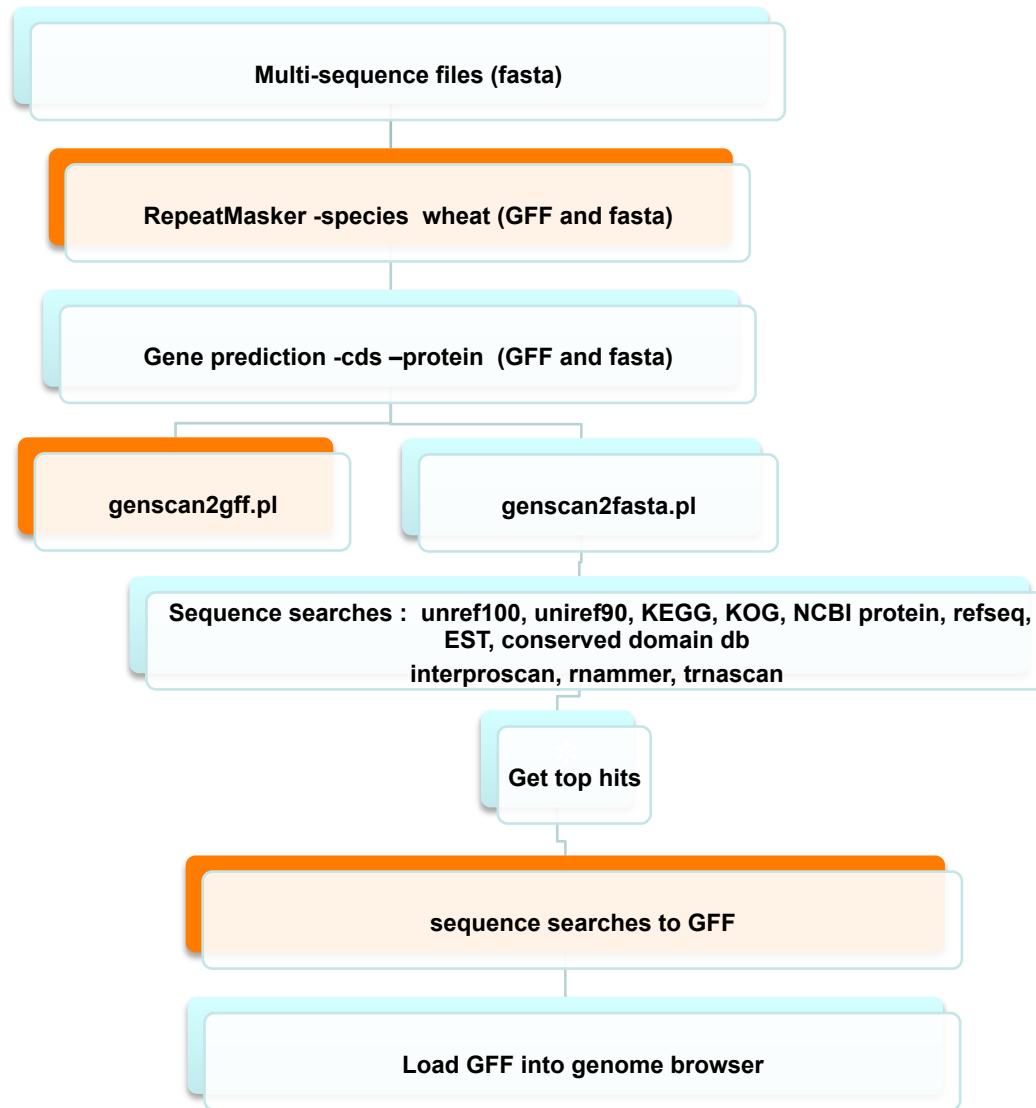
The screenshot shows the yabi bioinformatics workflow interface. The top navigation bar includes 'jobs', 'design' (which is selected), and 'files'. A sub-menu for 'unnamed (2011-03-31 10:35)' is open, showing options for 'start', 'accepts', and 'outputs'. On the left, a sidebar lists various genomic tools: genemark, iRNAscan-SE, bartab, clustalw, t_coffee, genemarkhmm, gencan, glimmerhmm, repeatmasker, glimmer, aragom, and gpchmm. The main workspace displays a 'Options for 1 - select file' dialog with a 'files' section containing two entries: 'gridftp://pmoolhuijzen@scratty.localdomain/yabi/pmoolhuijzen/' and 'gridftp://pmoolhuijzen@xe-g14.ivec.org/scratchbi01/pmoolhuijzen/'. The bottom right corner of the interface shows the copyright notice '© 2006, 2011 CCG, Murdoch University' and a 'log out pmoolhuijzen' link.



Bioinformatics annotation workflows URGI (triannot) and YABI



Bioinformatics workflows - Annotate sequences (YABI and Triannot)



YABI - Front-end

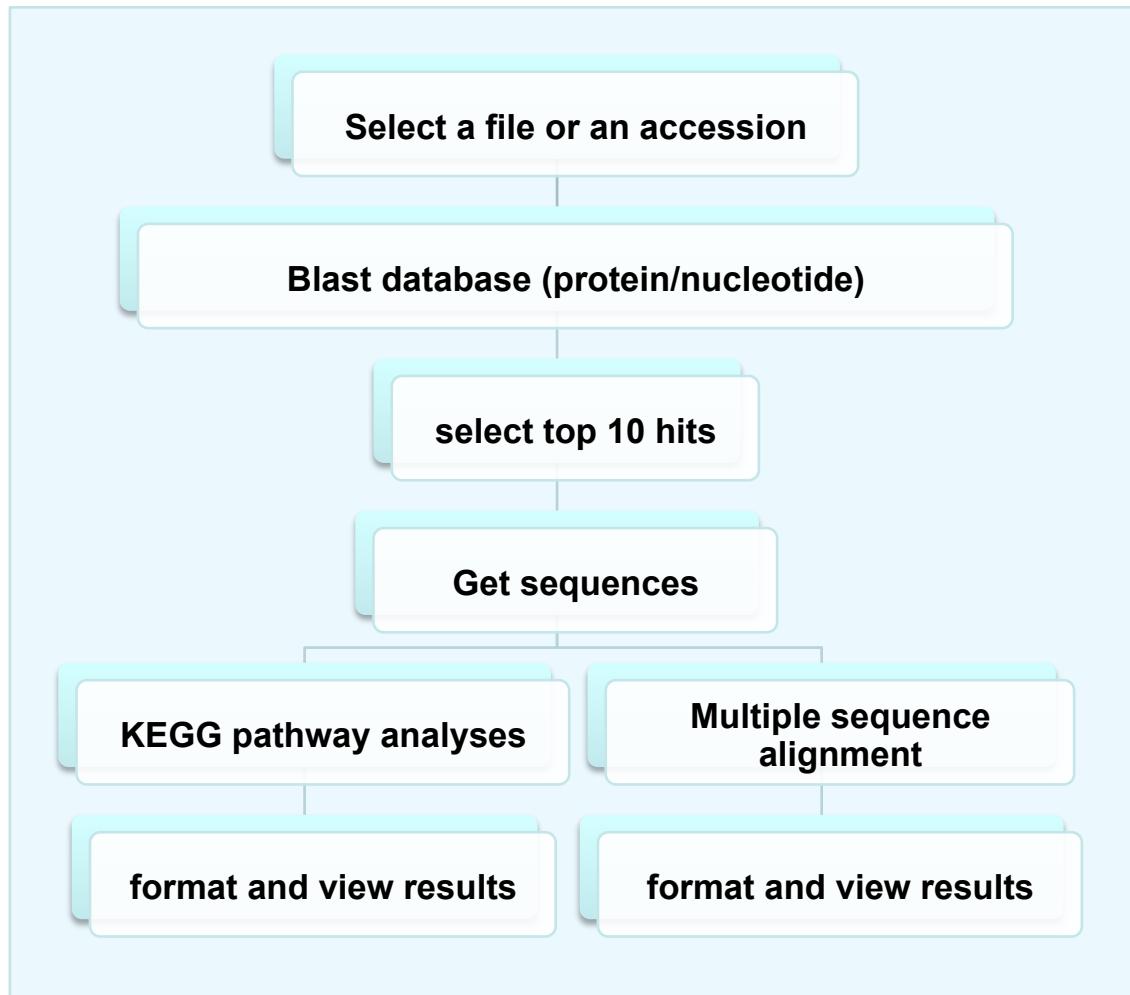
- Designed for a non-technical users
 - jobs tab, design tab, files tab
- Simple and easy to use as possible
- Reuse workflows
- User access
 - When you log in, user sees what they have access to
 - Allow scientists to work together
- Drag and drop tools

YABI - Front-end (2)

- Usability
 - Users warned if tools dragged out of place
 - Anticipate file extensions require for a given tool
 - Tools filtered
 - Tags – capture meta data
 - Errors trapped by system
- File manager
 - Drag and drop files
 - File copying via streaming
 - not via front-end
- Command-line
 - Power users

YABI demo

Simple Blast search and further analyses



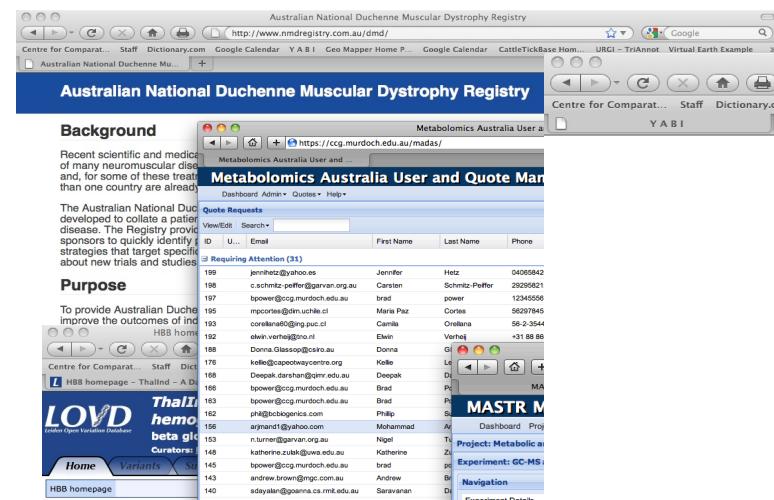
CCG: Hardware



- Stage 1A Pawsey Centre
- Ranked 87 in the world
- 9600 cores



CCG: Software development



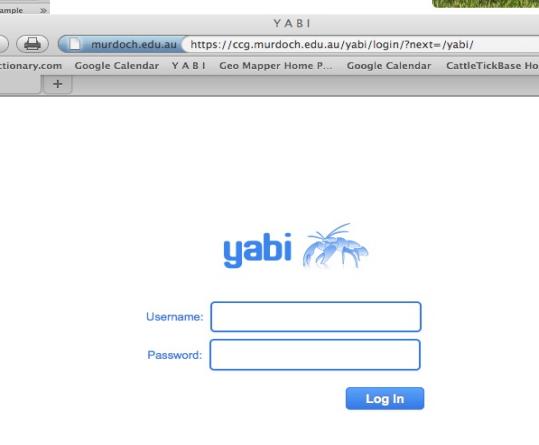
Australian National Duchenne Muscular Dystrophy Registry

Background
 Recent scientific and medical advances have led to the development of many neuromuscular diseases, and, for some of these treatable diseases, there are now treatments available. The CCG's aim is to support the sponsors to quickly identify strategies that target specific diseases.

Purpose
 To provide the Australian Duchenne muscular dystrophy community with a central place to improve the outcomes of individuals with the disease. The CCG's aim is to support the sponsors to quickly identify strategies that target specific diseases.

LOVD Thalidomide Database Curators: Home Variants HBB homepage

Centre for Comparative Genomics Western Australia



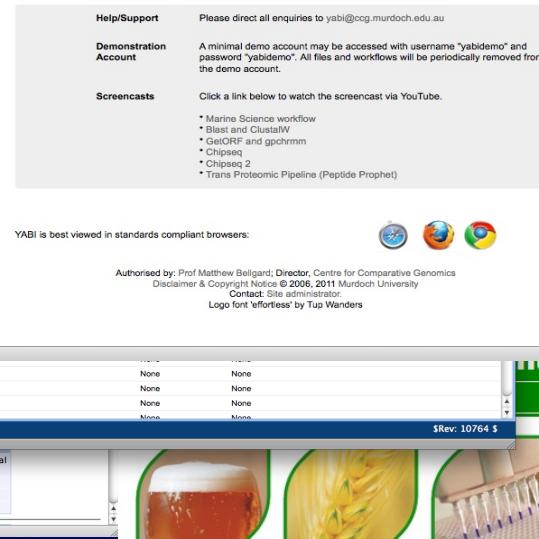
Metabolomics Australia User and Quote Management System

Quote Requests

ID	U...	Email	First Name	Last Name	Phone
199	s.mcmillan@unsw.edu.au	Jennifer	Hetz		04050545
198	s.mcmillan@unsw.edu.au	Carsten	Scheutz-Pfeifer		20250501
197	tpower@dm.murdoch.edu.au	Iain	power		13245558
196	mpcorrie@dm.uchile.cl	Maria Paz	Conce		56207045
193	corralde@ing.puc.cl	Camila	Ovalle		56-2-3544
192	ewin.verheij@zotu.nl	Ewin	Verheij		+31 88 88
188	Donna.Glassop@csiro.au	Donna	Glassop		
176	kelle@apepdcowentre.org	Kelle	L		
168	Deepak.darshani@vanderbilt.edu	Deepak	D		
166	tpower@ccg.murdoch.edu.au	Brad	PT		
163	tpower@ccg.murdoch.edu.au	Brad	R		
162	phat@cbogenomics.com	Philip	S		
156	ajrand@ yahoo.com	Mohammad	A		
153	nathan@unsw.edu.au	Nigel	T		
148	kathleen.zuk@unsw.edu.au	Adrienne	Z		
145	tpower@dm.murdoch.edu.au	Iain	Power		
143	andrew.brown@mpg.de	Andrew	B		
140	sdoyanar@panca.cs.rmit.edu.au	Saravanan	D		
124	tpower@dm.murdoch.edu.au	Brad	PD		
123	tpower@dm.murdoch.edu.au	Brad	PD		
115	tpower@dm.murdoch.edu.au	Brad	PD		
113	ricktak@gmail.com	test	T		
111	sravaranan.dayalan@gmail.com	Saravanan	D		
108	tpower@ccg.murdoch.edu.au	Brad	PD		
107	sravaranan.dayalan@gmail.com	Saravanan	D		
101	sravaranan.dayalan@gmail.com	Saravanan	D		
100	sdoyanar@panca.cs.rmit.edu.au	Saravanan	D		

MASTR M

Dashboard **Proj**



yabi

Log In

Help/Support Please direct all enquiries to yabi@ccg.murdoch.edu.au

Demonstration Account A minimal demo account may be accessed with username "yabidemo" and password "yabidemo". All files and workflows will be periodically removed from the demo account.

Screencasts Click a link below to watch the screencast via YouTube.

- Marine Science workflow
- Blast and ClustalW
- GetORF and gpcrmm
- Chipsseq
- Chipsseq 2
- Trans Proteomic Pipeline (Peptide Prophet)

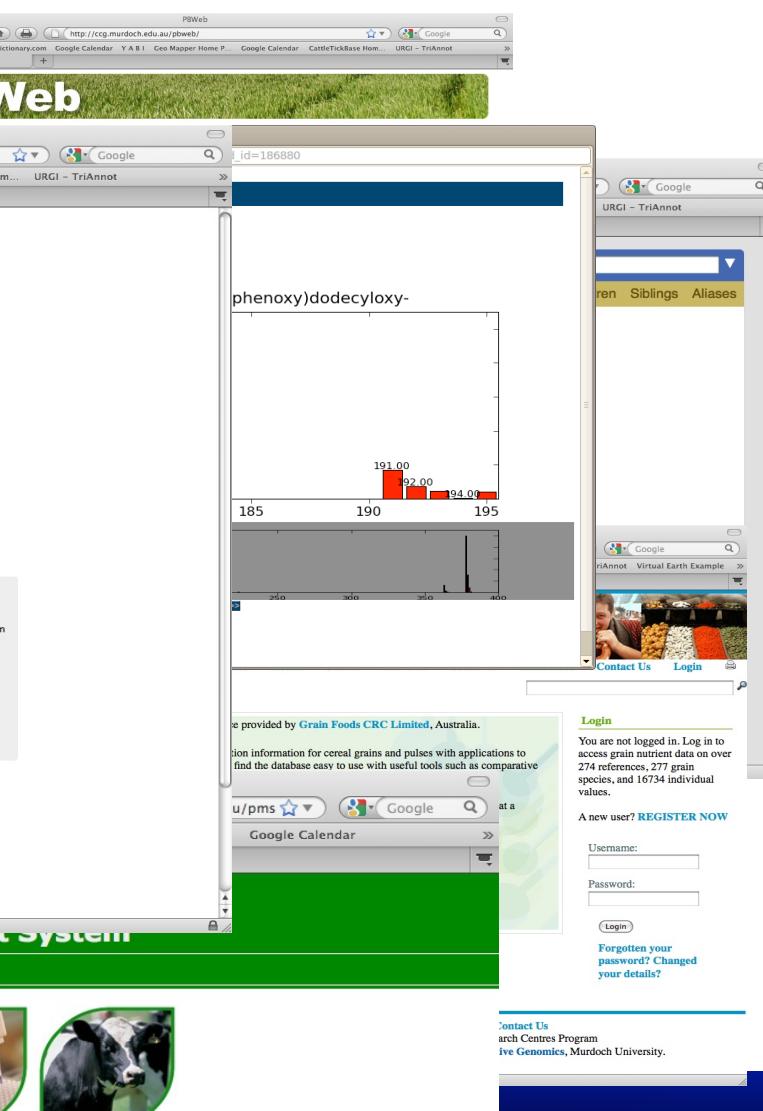
YABI is best viewed in standards compliant browsers:   

Authorised by: Prof Matthew Bellgard, Director, Centre for Comparative Genomics
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 Logo font 'effortless' by Tup Wanders

Copyright information

1586	6	None	None
1585	5	None	None
1584	4	None	None
1583	3	None	None

Done



PB Web

YABI

Phenotype id=186880
 phenoxy)dodecyloxy-

Mass Spectrum 185 190 195
 191.00 92.00 94.00

Google

URGI - TriAnnot

Contact Us **Login**

LOVD Gene homepage

LOVD Thalidomide Database Curators: Home Variants HBB homepage

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Graphical displays and utilities

Summary tables Summary of all sequence variants in the HBB database, sorted by type of variant (with graphical displays and statistics)

UCSC Genome Browser Show variants in the UCSC Genome Browser ([compact view](#))

Ensembl Genome Browser Show variants in the Ensembl Genome Browser

NCBI Sequence Viewer Show distribution histogram of variants in the NCBI Sequence Viewer

Done

Next generation sequencing analysis projects

- De novo genome/transcriptome assembly and annotation
 - Wheat, barley, rat mutant, Wine yeast, cane toad, Rhizopertha, Campylobacter, Euphorbia, Cattle tick, dog tick, rhizobium, spirochetes
- Transcriptomics/epigenomics/metagenomics
 - microRNAs (human, Arabidopsis, cattle tick)
 - Epigenomics in Arabidopsis
 - Metagenomics of environmental samples (ancient DNA)
- Disease association
 - LPK rat mutant, human disorders, diagnosis assays

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Informatics considerations

- Process/timing to integrate 3B CCG results back to IWGSC
- Protocols for BAC physical assembly
- Process/timing to integrate 7A results back to IWGSC
 - Linkages to CCG resources
- CCG could assist in scoping LTC software porting requirements

Acknowledgements

CCG Team

- Rudi Appels
- Gabriel Keeble
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- Andrew McGregor
- Paula Moolhuijzen

Collaborators

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- Mingcheng Luo (UC Davis)

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- BioPlatforms Australia