

*The Reference Sequence of Wheat  
and IWGSC Phase II:  
Ensuring a Full Genomic Toolbox for Wheat*

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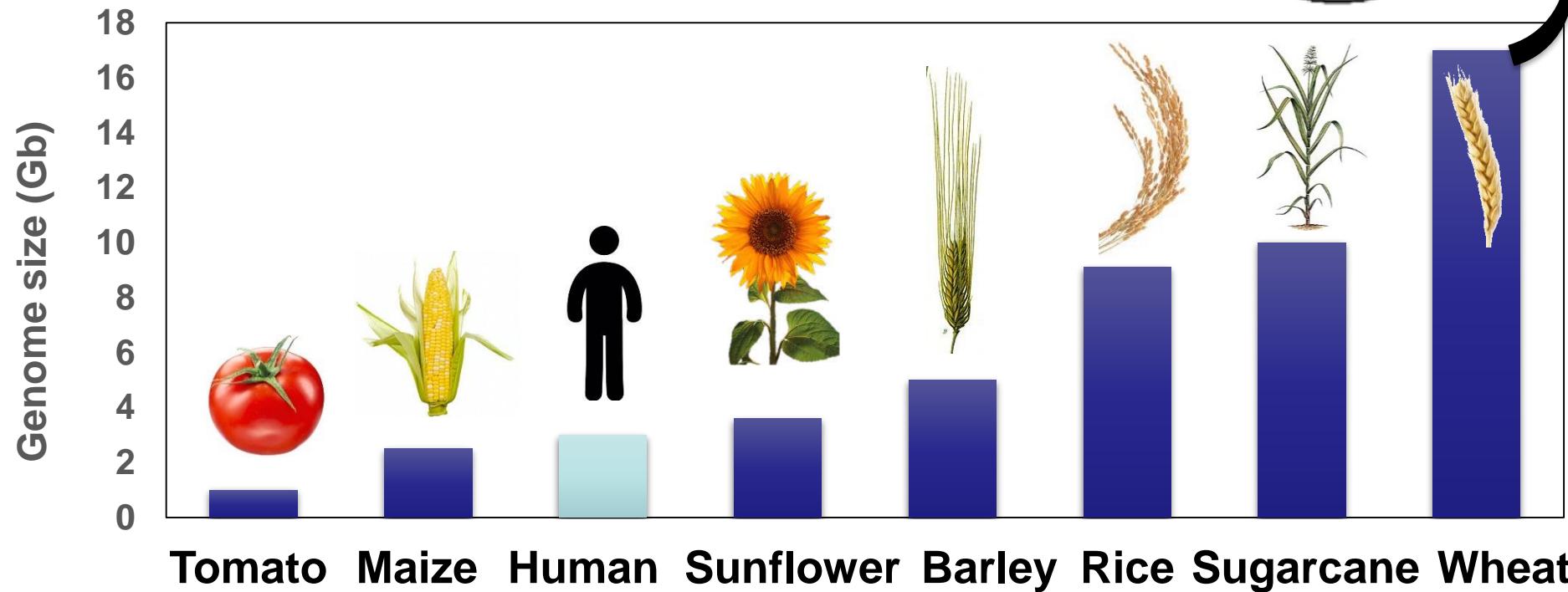
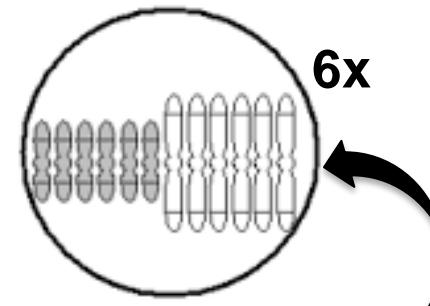


WHEATS&WOMEN INTERNATIONAL CONFERENCE  
Roma – June 2018



# Plant's genome exhibits high levels of complexity

- Large genome size
- High level of transposable elements
- Polyploidy

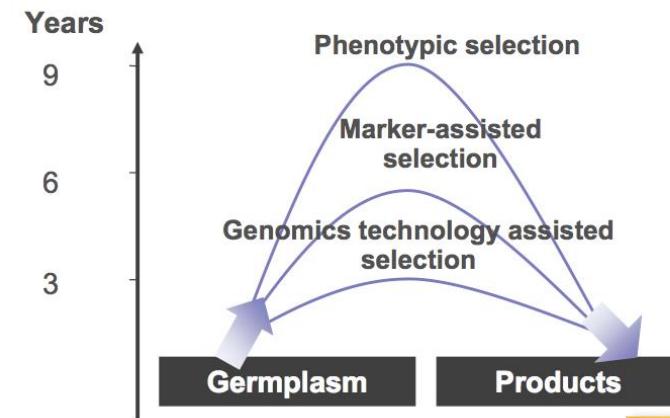
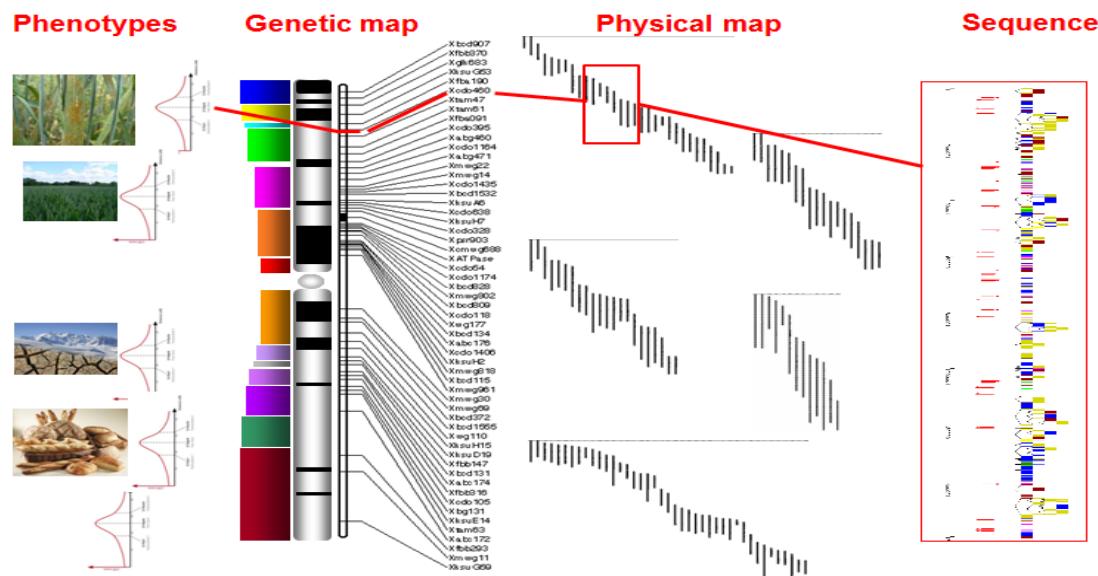


# 2005 – IWGSC Goal & Vision



- Lay a foundation to accelerate wheat improvement
  - Increase profitability throughout the industry

- High quality annotated genome sequence, comparable to rice
  - Physical map-based, integrated and ordered sequence



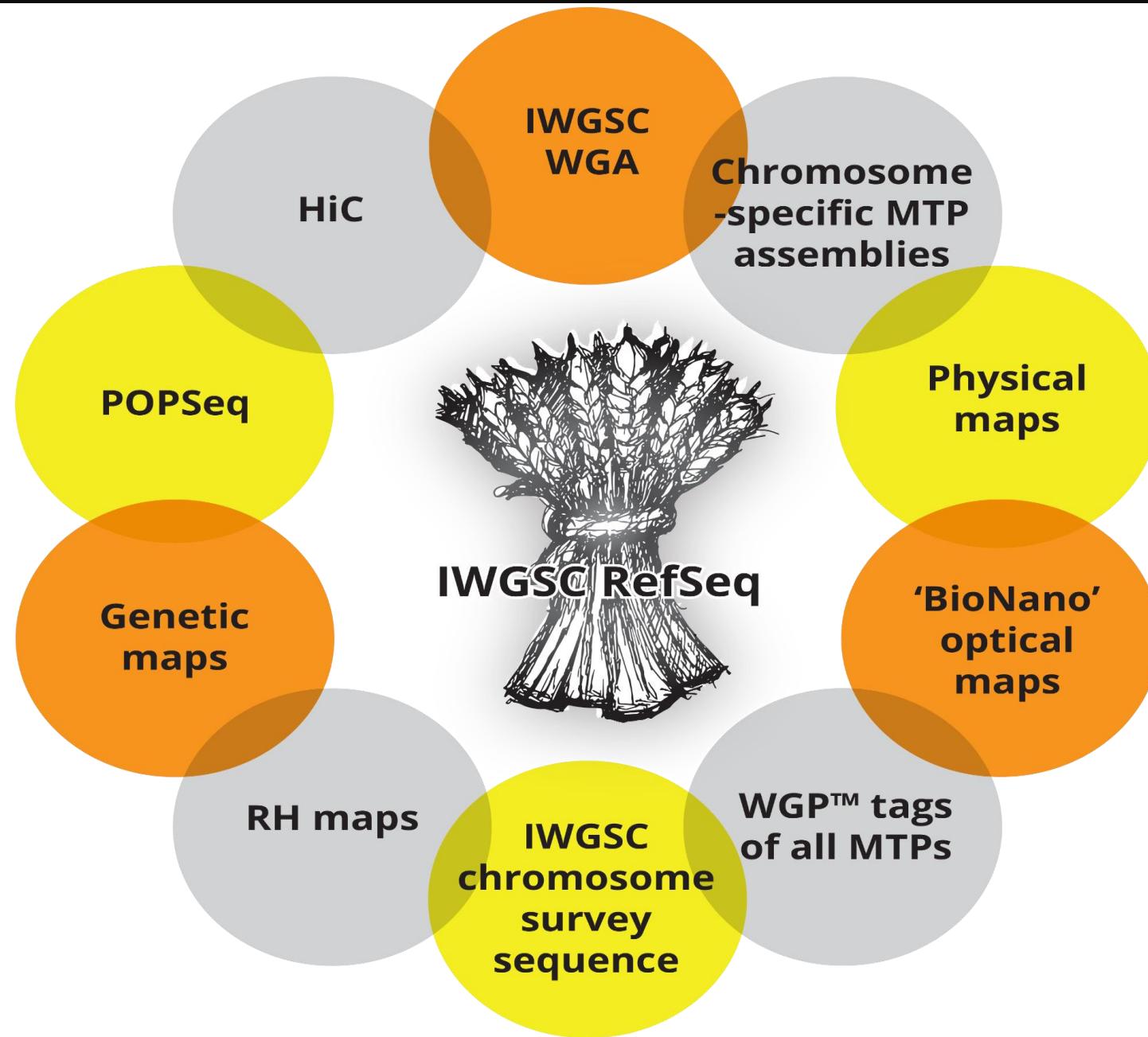
# The Reference Sequence for the Bread Wheat Genome



Frédéric Choulet  
*GDEC, INRA, UCA, Clermont-Ferrand, France*

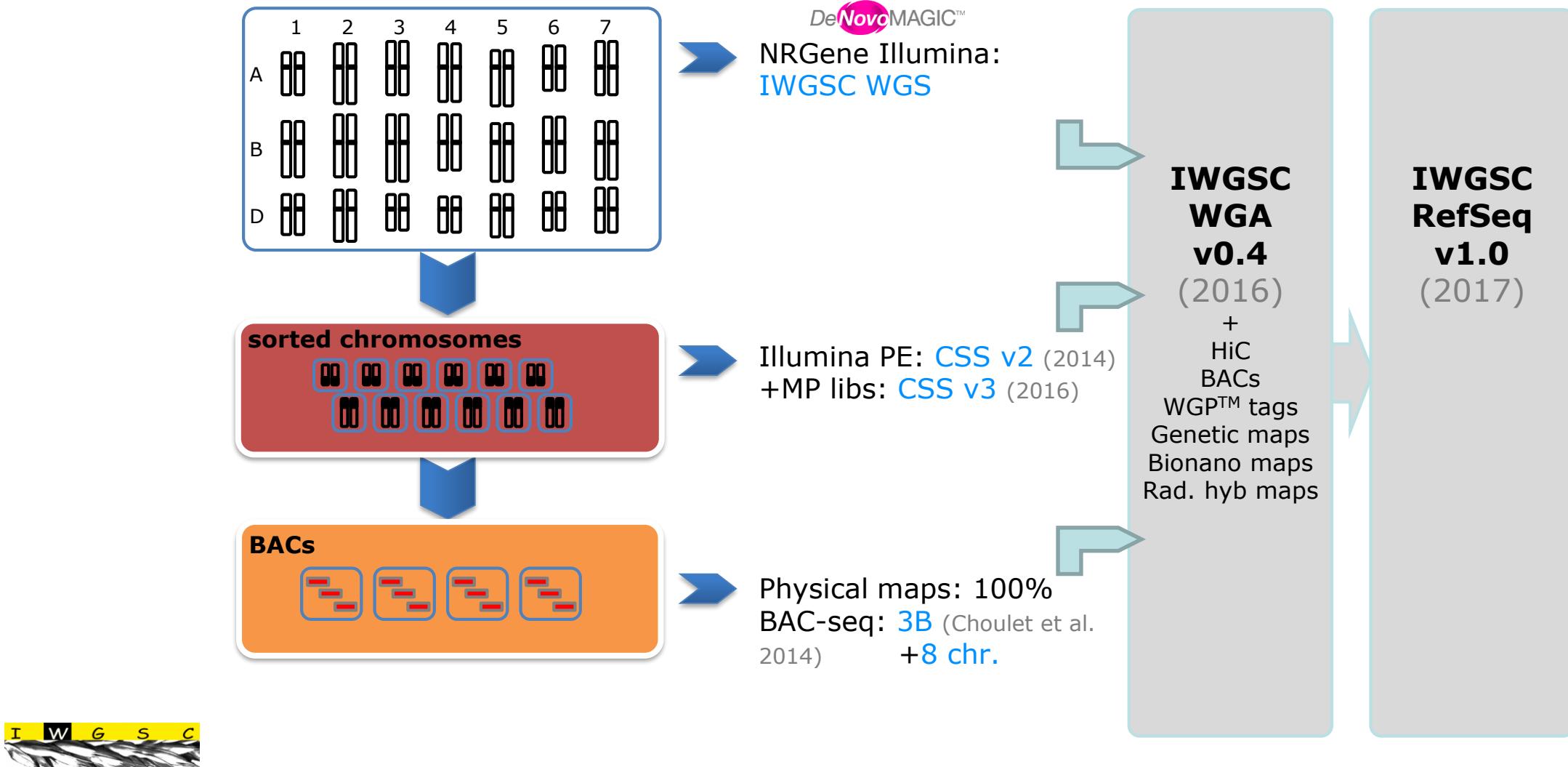


# Concerted integration of resources: RefSeq v1.0



# RefSeq v1.0

A fully annotated and anchored reference sequence assembly of the wheat genome



## **IWGSC RefSeq v1.0** – *Community resources*

- Physical maps for all chromosomes
  - ▶ 1,839,128 BACs
- WGP tags (mostly from MTP BACs) for all chromosomes except 3B
  - ▶ 4,305,249 unique tags for 693,697 BACs
- BAC sequence assemblies
  - ▶ 8 chr: 1A, 1B, 3B, 3D, 6B, 7A, 7B, 7D + 2 chr arms: 4AL, 5BS
  - ▶ 52,890 BACs (9.7 Gb), N50=68 kb
- BioNano maps for 7A, 7B and 7DS
  - ▶ 1,335 BioNano contigs
- 4.7 M molecular markers



## □ IWGSC RefSeq v1.0 – Metrics

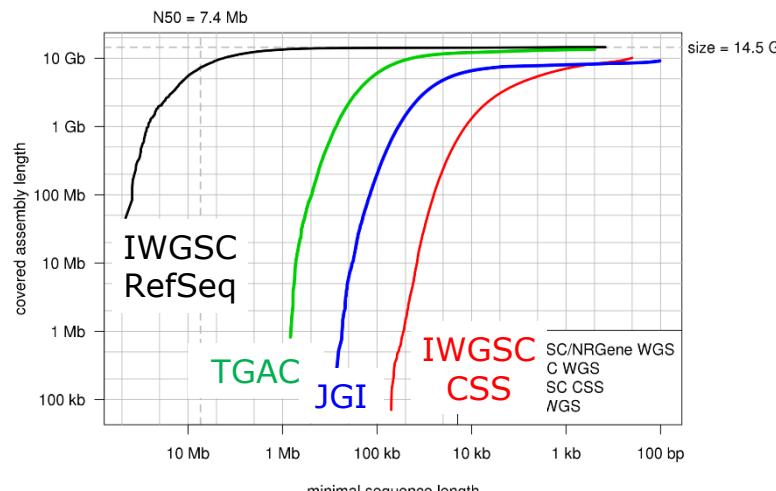
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total size	14.5 Gb (genome size ~15.5 -> <a href="#">94%</a> )
completeness	<a href="#">97-99%</a>
contig N50	52 kb
scaffold N50	7 Mb
superscaffold N50	23 Mb

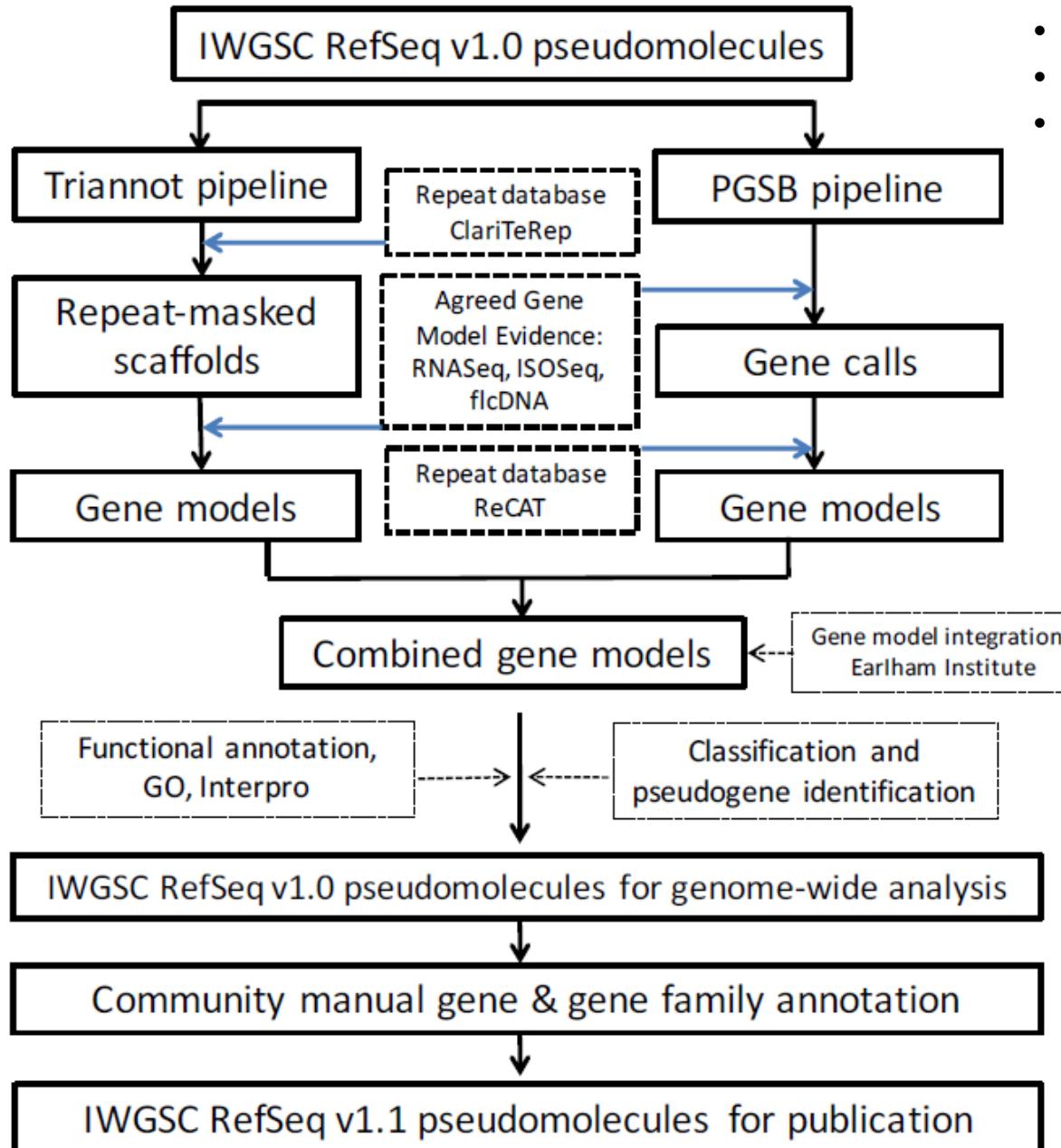
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### **21 pseudomolecules:**

total size	14.1 Gb (97%)
superscaffolds	1601 (avg 76 per chr.)



# □ IWGSC RefSeq v1.0 – Genome annotation pipeline

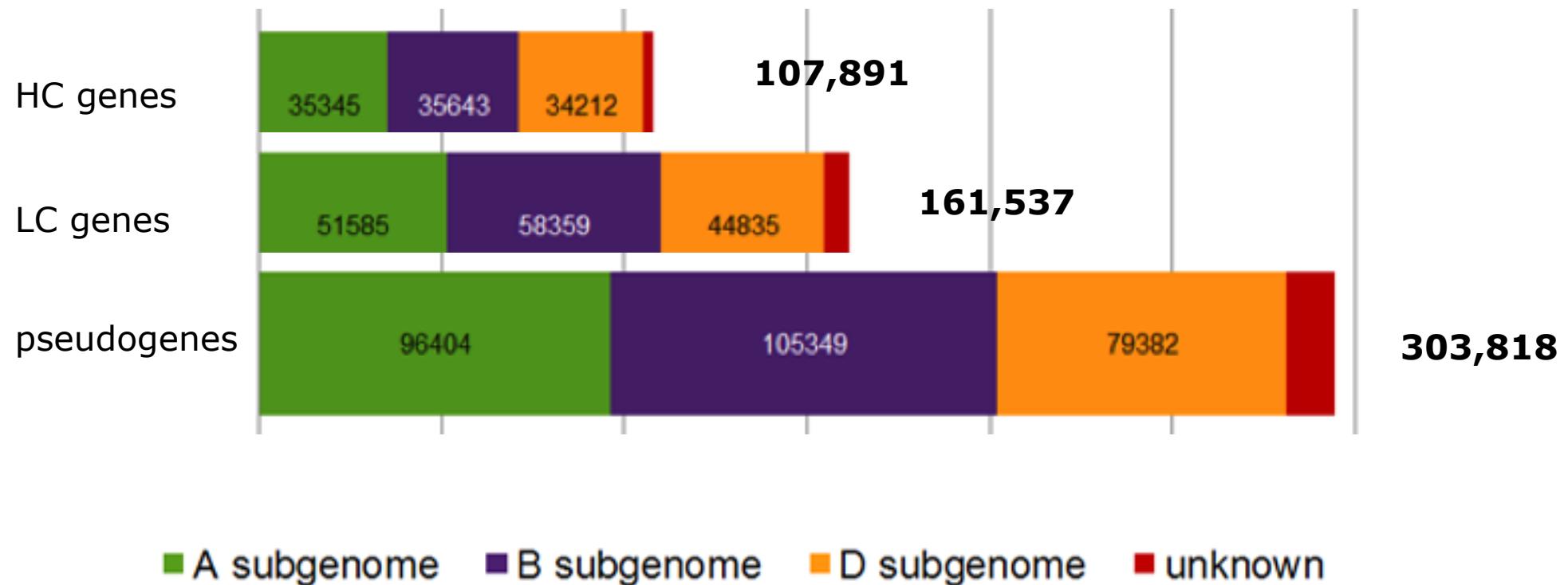


- **GDEC INRA** Clermont, France – *Rimbert Leroy Choulet et al.*
- **PGSB** Munich, Germany – *Spannagl Twardziok et al.*
- **Earlham Inst.** Norwich, UK – *Swarbreck Venturini et al.*

## □ Annotation – gene models

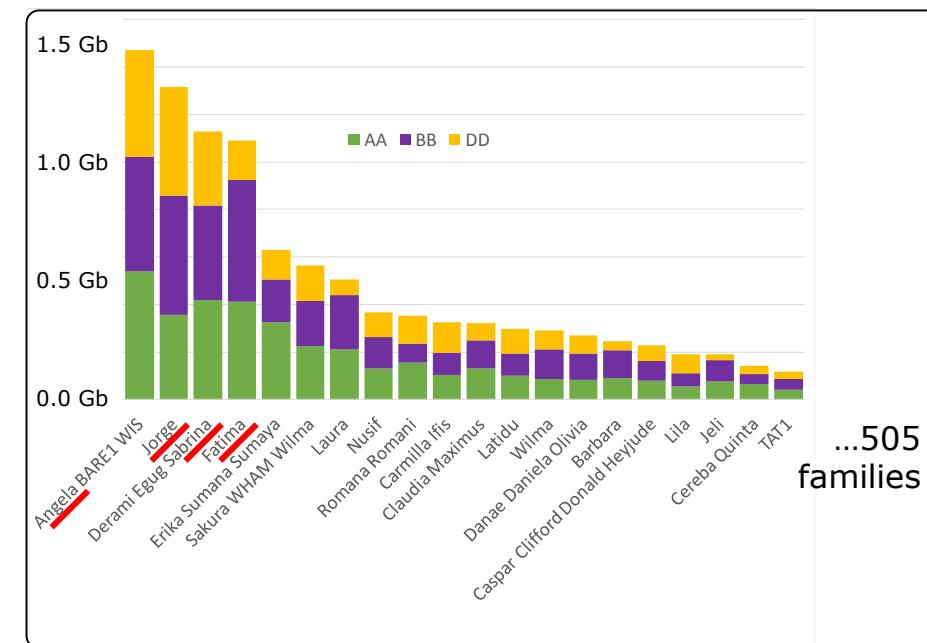
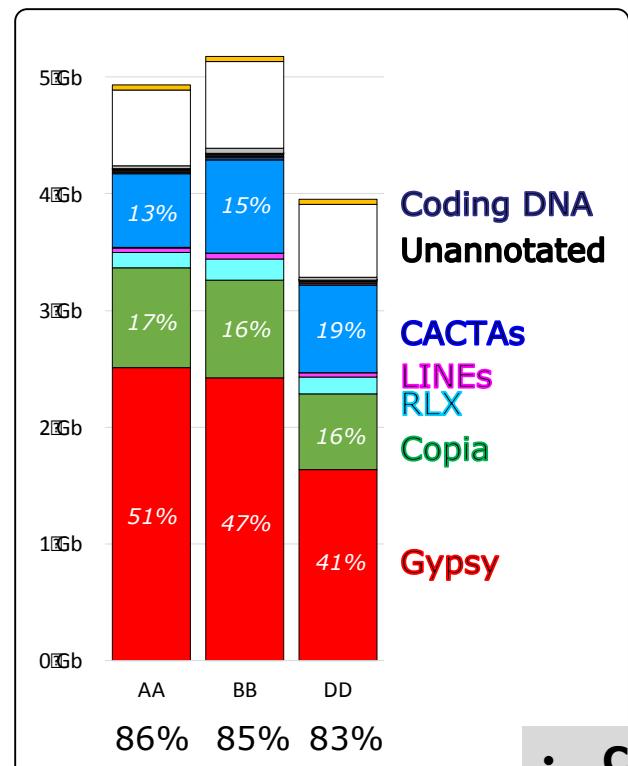
v1.0: automated

v1.1: incorporate 3685 manually annotated genes (available upon publication)



## □ Annotation – transposable elements

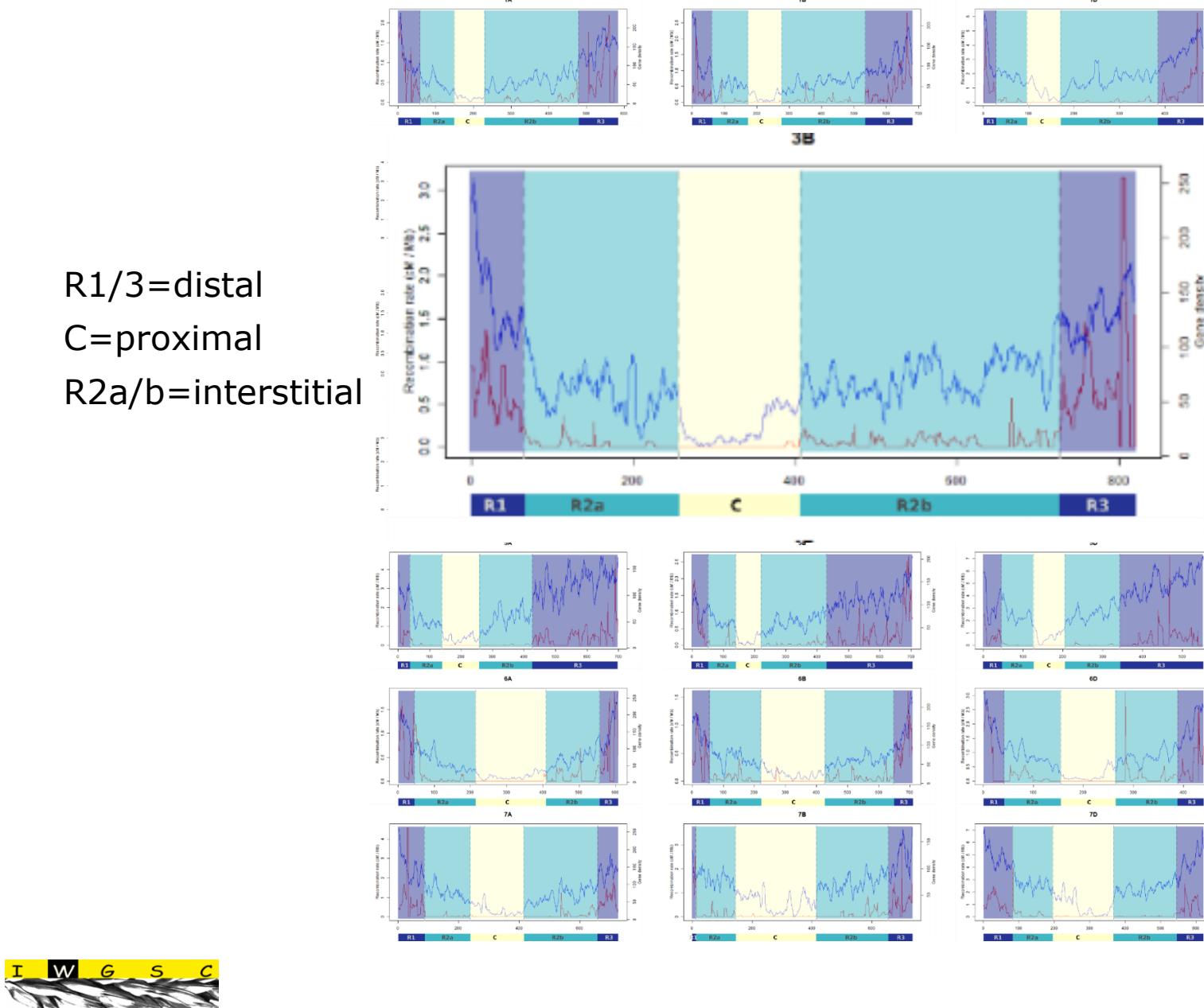
- CLARITE, TREP, *denovo* fl-LTR-RTs detection...
- 3,968,974 elements belonging to 505 families ( $84\% <=> 11.9 / 14.1 \text{ Gb}$ )



- **Conserved TE composition of A-B-D (no family specific to 1 subgenome)**
- **Complete reshuffling of TEs between homeologous loci**
- **TE activity after polyploidization very limited**

# □ Genome partitioning

R1/3=distal  
C=proximal  
R2a/b=interstitial



R1/3= high gene / low TE

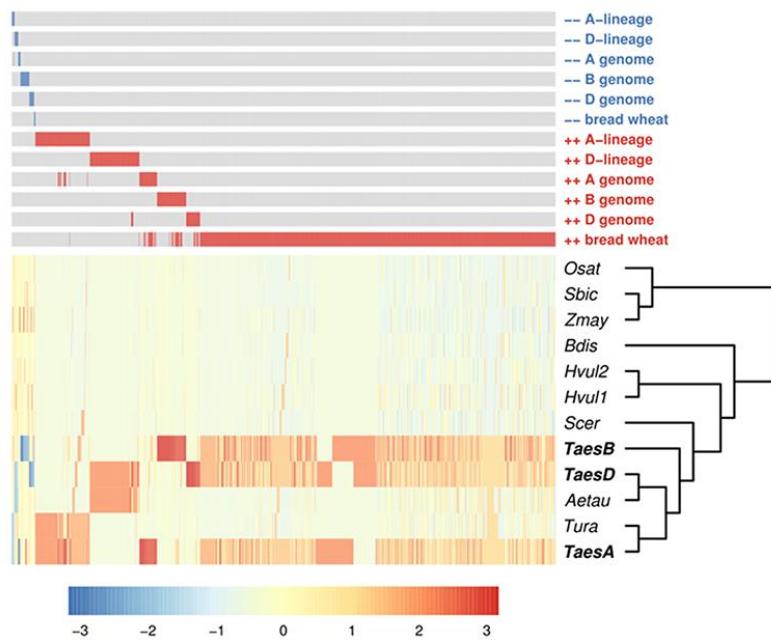
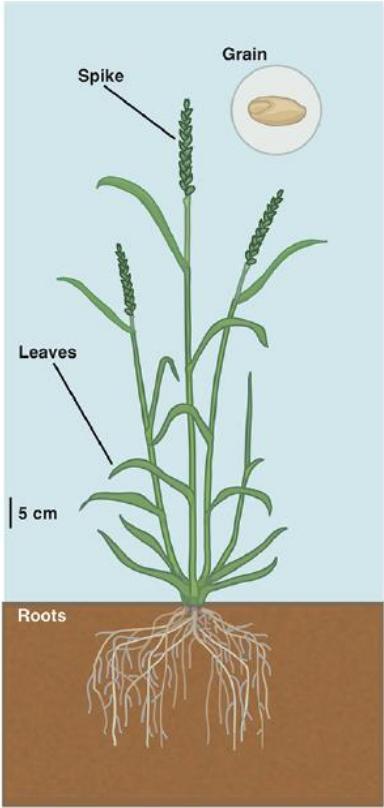


## □ A-B-D comparative analyses

Analysis of gene families of wheat

**Gene family expansion/contraction associated with traits targeted by breeder selection**

- 26080 gene families
- 33% expanded in the wheat lineage / 0.1% contracted families  
72% expanded in A+B+D



GO/PO/TO term enrichment and expanded gene families:

- **morphological traits** related to leaf, spike and root development, leaf size, tillering, vegetative growth and development, seed, endosperm, and embryo-development and morphology
- **grain yield and quality** (seed maturation, dormancy and germination)
- **abiotic stress** tolerance (salt stress, cold stress)

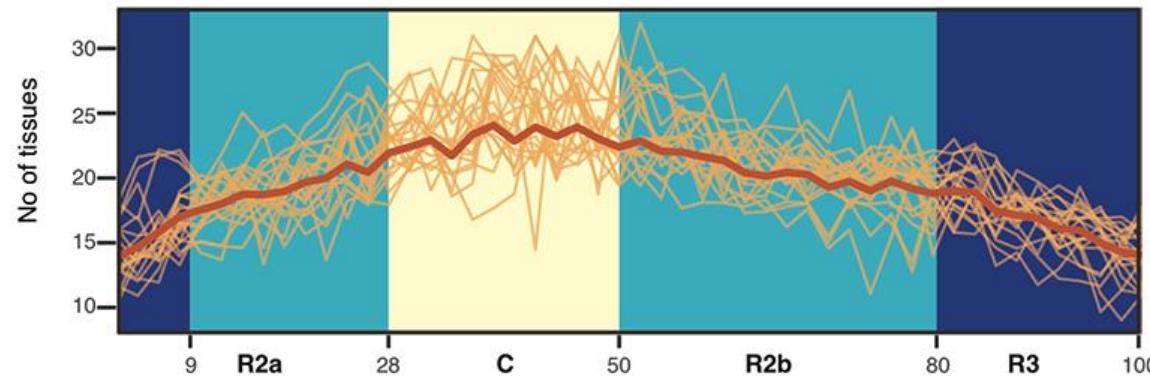


**B sub-genome expanded families enriched for genes related to plant and leaf size and development, tillering and vegetative growth time:  
predominant impact of the B genome on vegetative plant growth and development**

## □ Transcriptome atlas

### Atlas of transcription reveals trait associated gene co-regulation networks

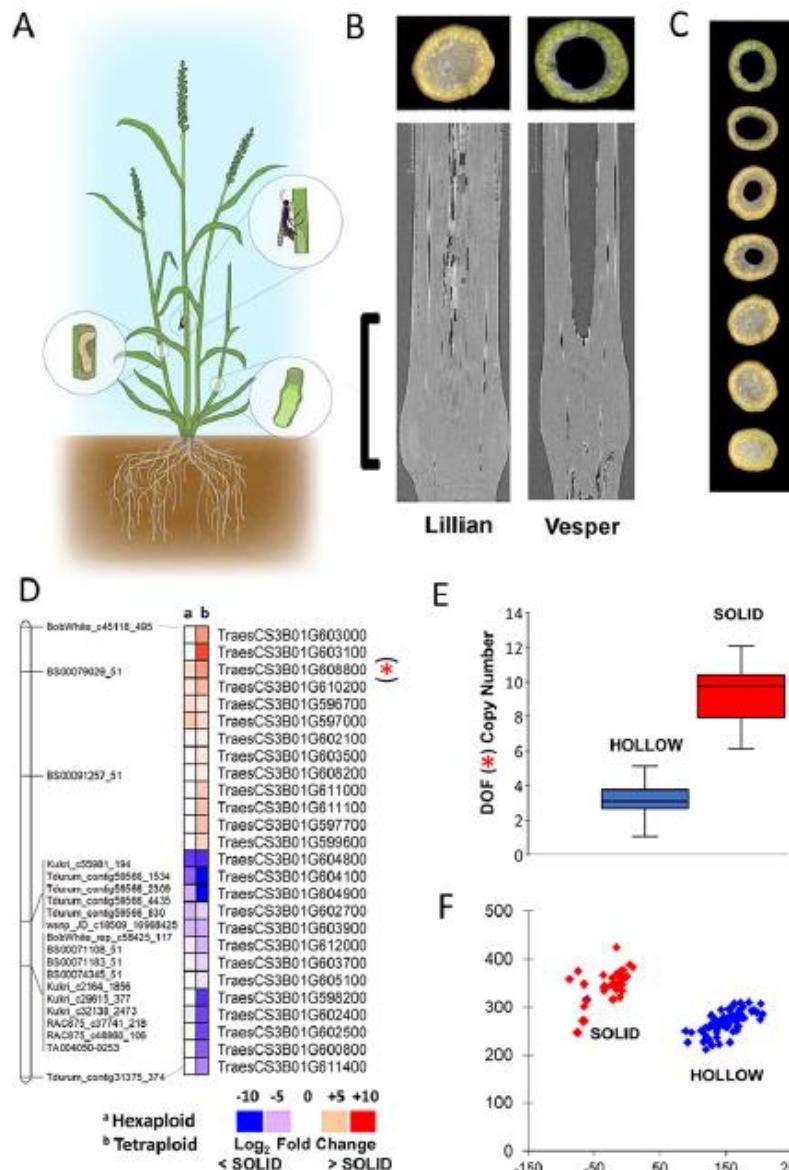
- 850 RNASeq samples – 32 tissues
  - 85% HC genes expressed
  - 49% LC genes expressed
- Chromosome partitioning
  - Distal region genes expressed in fewer tissues and enriched for responses to stress



Genes located in the distal R1 and R3 regions : lower expression breadth than those in the proximal regions



# ☐ Reference-guided dissection of an insect/abiotic stress resistance QTL for improved marker assisted selection



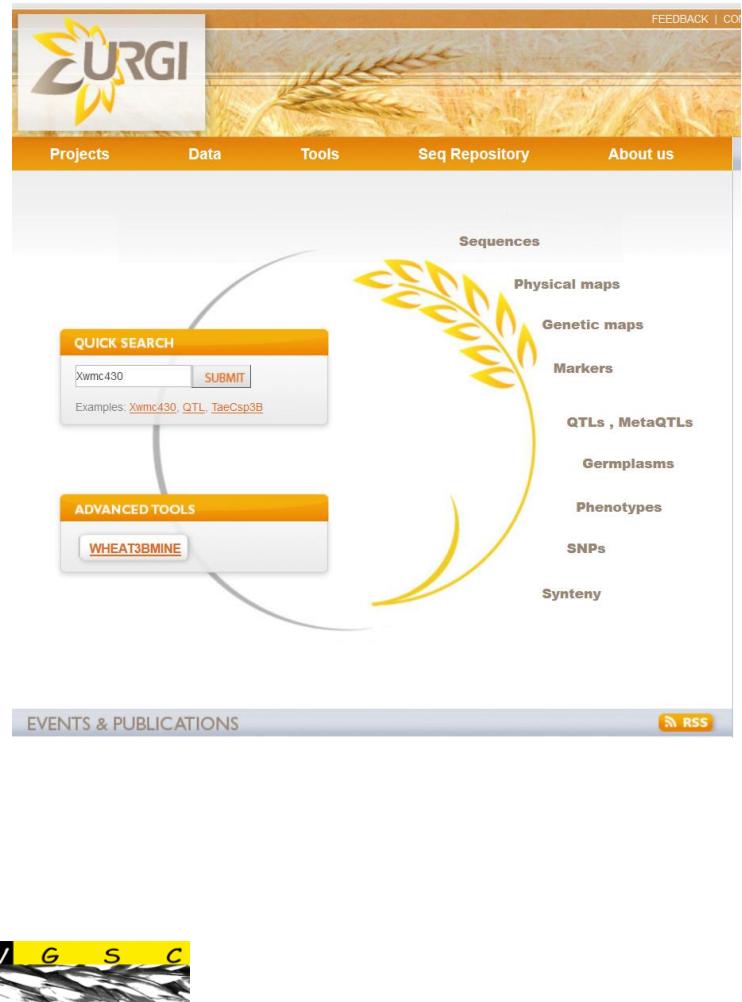
- traits of agronomic importance are inherited as QTL
- informative DNA markers in strong linkage disequilibrium with QTL are essential
- IWGSC RefSeq v1.0 provides template for assigning molecular markers to coordinates in linear DNA molecules anchored to high density molecular maps
- Example described : dissecting a QTL associated with insect/abiotic stress resistance

## ➤ Diagnosis marker

➤ Combining this knowledge with major loci selected for breeding programs provides a new framework for breeders to tackle the challenges of the new agriculture (balance the selection processes for adaptation to biotic and abiotic stress, end-use quality, and yield improvement)

# □ IWGSC RefSeq Data Access & Availability

<https://wheat-urgi.versailles.inra.fr/Seq-Repository/>



Pre-publication data access under Toronto Agreement:

- IWGSC **WGA v0.4**: June 2016
- IWGSC **RefSeq v1.0**: January 2017
- IWGSC **Annotation v1.0**: May 2017
- IWGSC **Annotation v1.1**: upon publication
- Manuscript under review

<http://www.wheatgenome.org/Tools-and-Resources>



# IWGSC Phase II: What's Next for the IWGSC



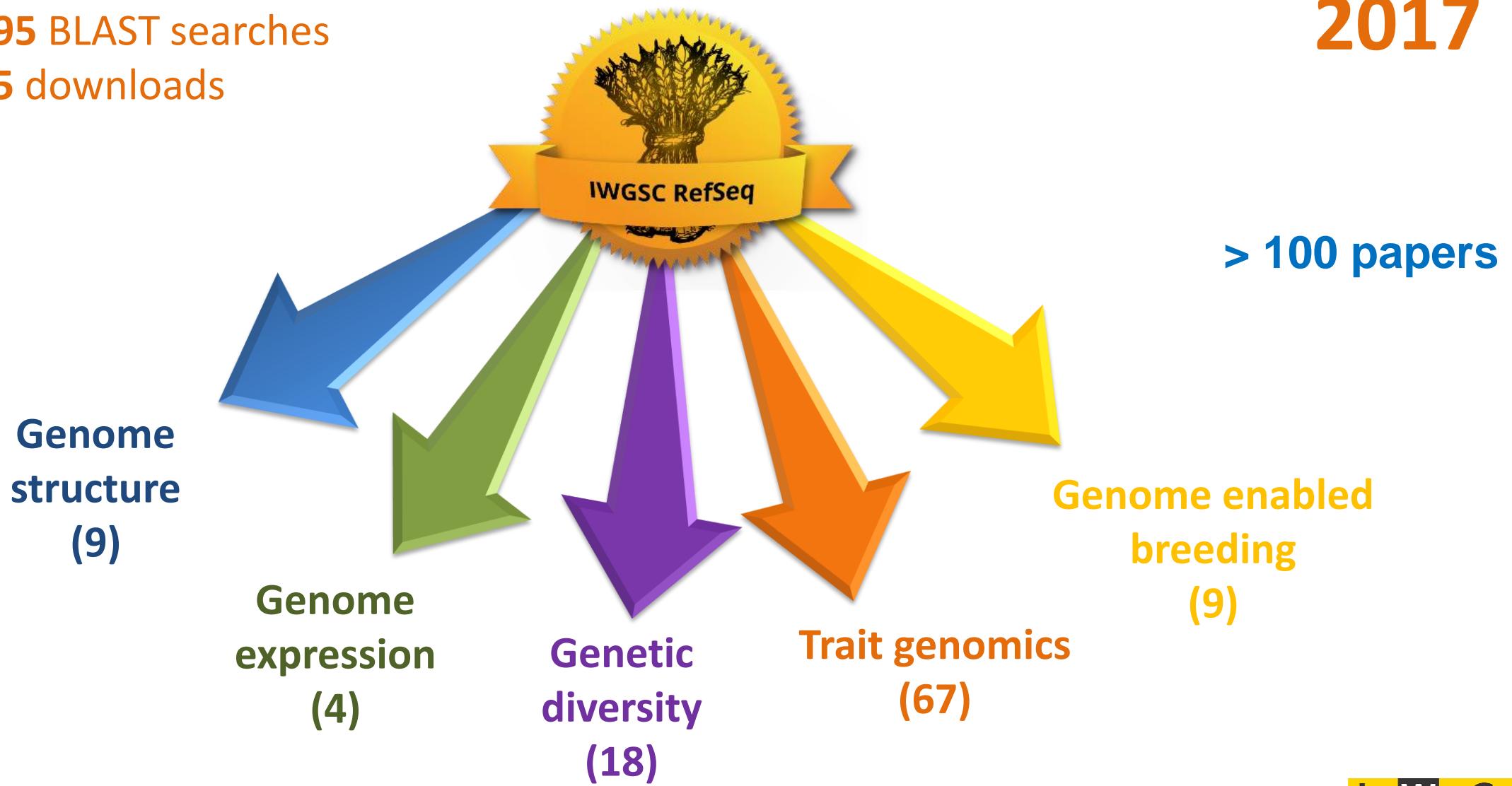
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# Measuring the Impact of IWGSC RefSeq v1.0

476,595 BLAST searches  
22,935 downloads

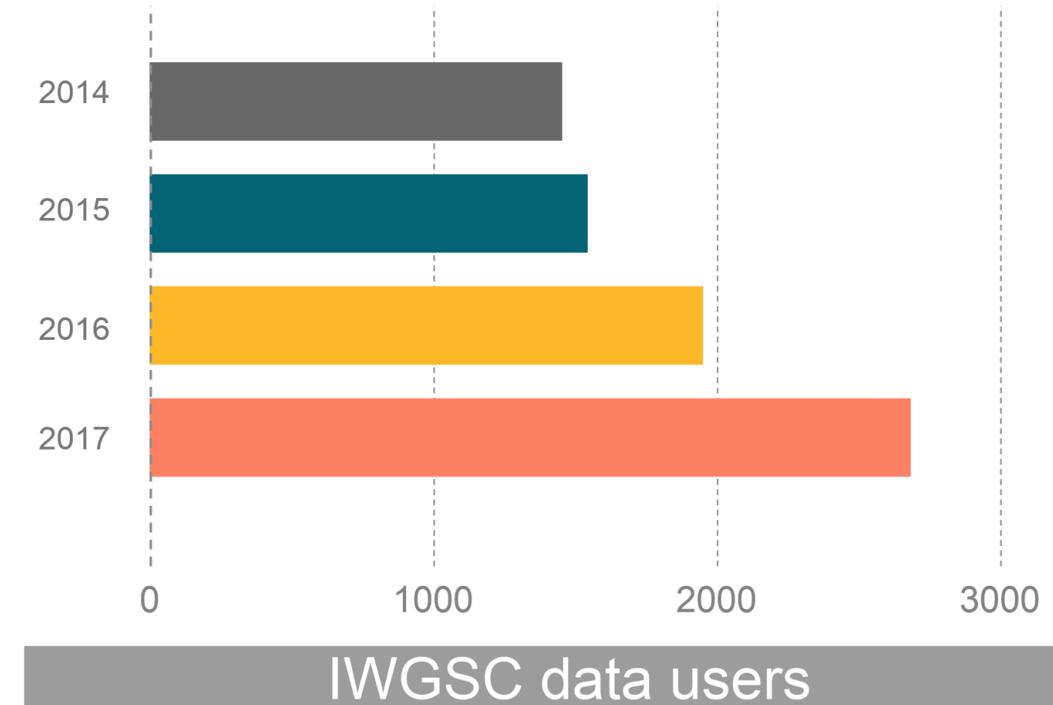
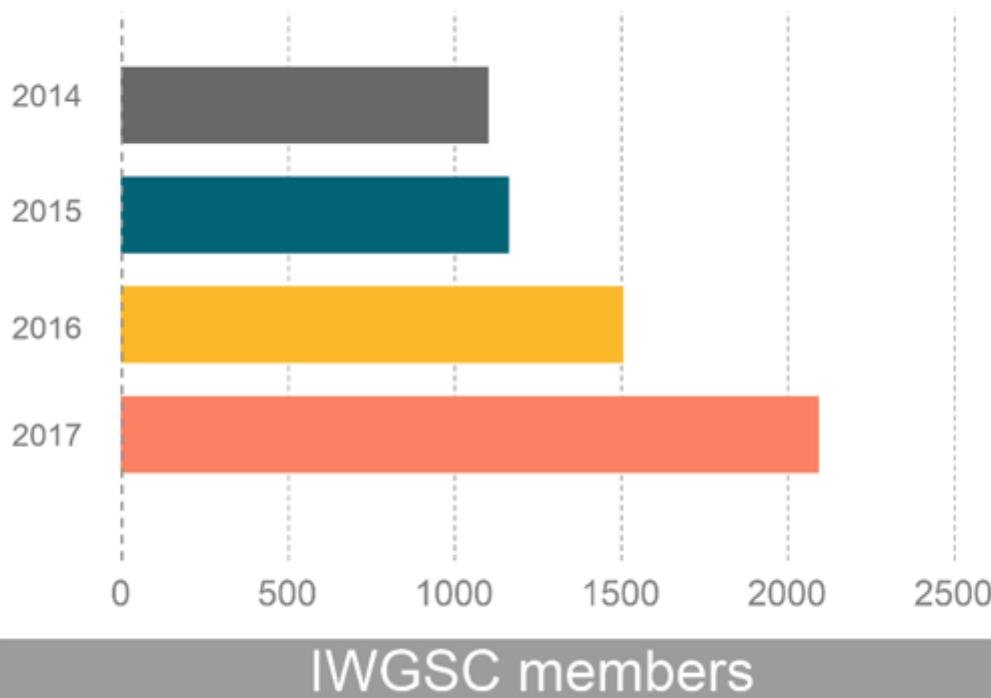
2017



# of papers referencing use of IWGSC RefSeq related resources since January 2017



# Measuring the Impact of IWGSC RefSeq v1.0



# Progress

2005  
Vision

- High quality annotated genome sequence, comparable to rice
- Physical map-based, integrated and ordered sequence



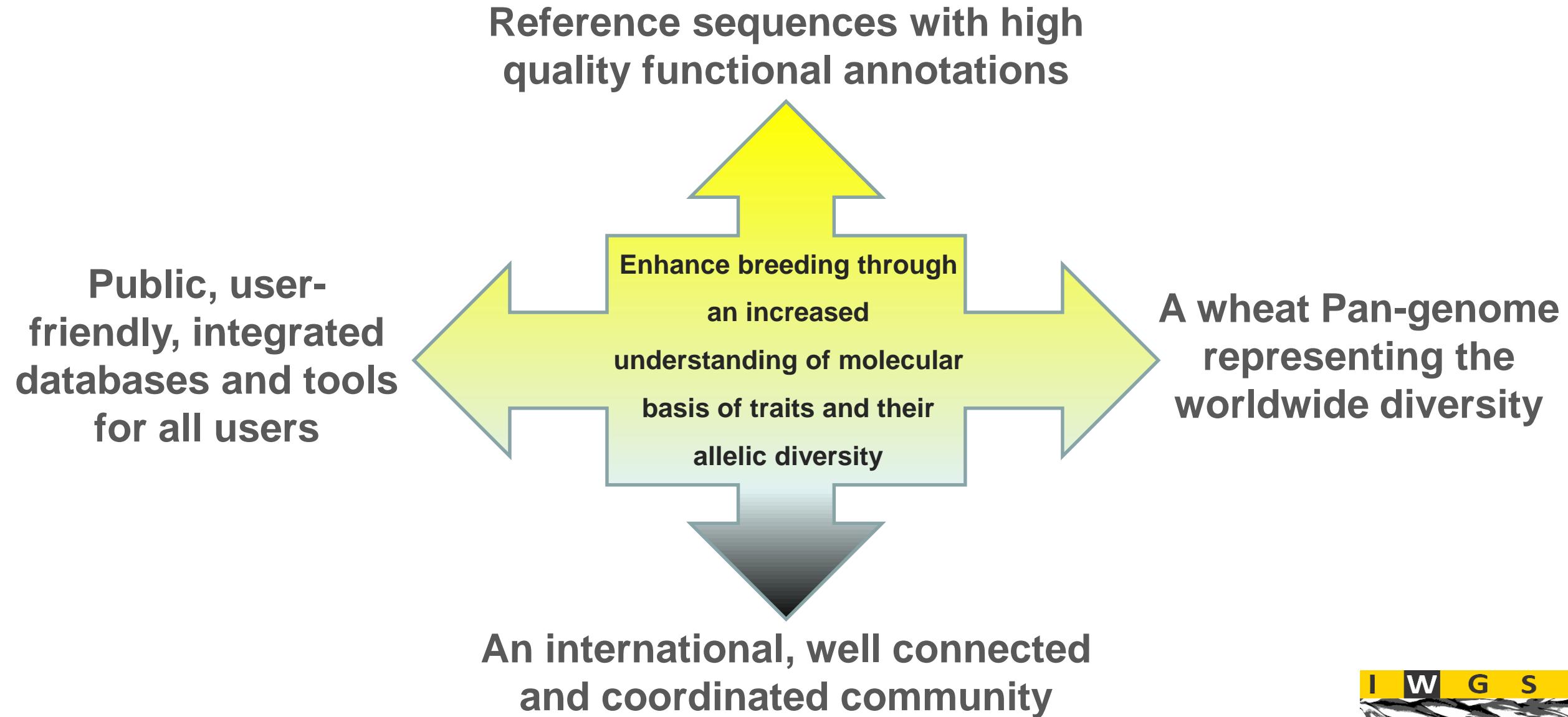
# Looking into the Future

- Achieve Gold Standard Reference
- Reach beyond reference to ensure a full genome-sequence based toolbox for wheat

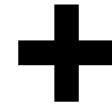


Goal: Accelerate wheat improvement

# Vision and strategy for the IWGSC 2.0



# From RefSeq v1.0 to IWGSC Gold Standard



Manual &  
Functional  
Annotation



- Pipeline for community generated manual annotation with future annotation releases
- Functional annotation
  - Gene family leaders
  - Applying machine learning techniques
- Closing gaps by incorporating new resources to build a IWGSC RefSeq v2.0
- IWGSC leaders: Rudi Appels, Fred Choulet.....

# IWGSC 2.0 Activities: Pan-Genome

- Develop a wheat pan-genome that represents the breadth of worldwide wheat diversity
- Reference quality for 8-12 landraces
- Diversity panels at draft quality
- Haplotype database
- Skim-sequencing and high throughput genotyping of many lines
- IWGSC team: Etienne Paux, Sébastien Praud,.....

# IWGSC 2.0 Activities: Database

- Develop user-friendly, integrated databases and tools
- IWGSC Pan-genome Database Task Force
  - Fred van Ex, Bayer CropScience
  - Magalie Leveugle, Biogemma
  - Matthieu Conte, Syngenta
  - Michael Alaux, URGI-INRA
  - .....
- What tools do you want?

# IWGSC 2.0 Activities: IWGSC Exome Array

- Wheat exome capture array based on IWGSC RefSeq annotation v1.1.
- Arbor Biosciences will offer enrichment reagents and an enrichment service
- Exome Team: Burkhard Steuernagel, Sreya Ghosh, Sébastien Praud, Hikmet Budak, Etienne Paux, Ute Bauhman
- Discussion: What modules would you like to see?

## □ Conclusion

- ▶ IWGSC goal for high quality reference achieved
- ▶ IWGSC RefSeq v1.0 incorporates highly diverse community resources (WGS assembly + Hi-C links + physical maps + CSS + BAC sequences + millions of markers)
- ▶ Draft v0.4 + pseudomolecules v1.0 + annotation v1.0 available for download prior to publication
- ▶ IWGSC Phase II
  - Continued improvements from manual and functional annotation
  - IWGSC Exome Array based on the sequence
  - Pan-genome that covers the breadth of wheat diversity (landraces and elite var.)
  - Pan-genome database, visualization, and tools for breeders

# ❑ Acknowledgments: Leaders & Major Contributors

**IWGSC RefSeq PIs:** [Rudi Appels](#), [Kellye Eversole](#), [Nils Stein](#), [Jane Rogers](#), [Catherine Feuillet](#), [Beat Keller](#)

**IWGSC Whole Genome Assembly PIs:** [Curtis Pozniak](#), [Nils Stein](#), Frédéric Choulet, Assaf Distelfeld, Kellye Eversole, Jesse Poland, Jane Rogers, Gil Ronen and Andrew G. Sharpe

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**Dehydrin gene family:** [Pilar Hernandez](#) and [Sergio Galvez](#)



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**1B BAC Sequencing & Assembly:** Frédéric Choulet and Etienne Paux

**1D, 4D, 6D Physical Mapping:** Bikram Gill

**2AL Physical Mapping:** Kuldeep Singh

**2AS Physical Mapping:** Nagendra K. Singh and Jitendra P. Khurana

**2B, 2D, 4B, 5BL & 5DL IWGSC-Bayer Whole Genome Profiling(WGP™) Physical Maps:** Jane Rogers and John Jacobs

**3AL Physical Mapping:** Bikram Gill

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**6B BAC Sequencing & Assembly:** Hirokazu Handa

**7A Physical Mapping & BAC Sequencing:** Rudi Appels

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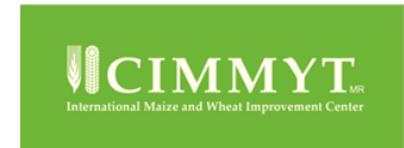
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